Tuberculosis Interventions

Interventions for Tuberculosis Control and Elimination

2002

International Union Against Tuberculosis and Lung Disease

Interventions for Tuberculosis Control and Elimination

2002

Hans L Rieder

International Union Against Tuberculosis and Lung Disease

68, boulevard Saint Michel, 75006 Paris, France

The publication of this guide was made possible thanks to the support of the United States Centers for Disease Control and Prevention, the British Columbia Lung Association, the French Ministry of Foreign Affairs, and the Norwegian Royal Ministry of Foreign Affairs.

Editor

International Union Against Tuberculosis and Lung Disease (IUATLD), 68 boulevard Saint Michel, 75006 Paris, France

Author : H. L. Rieder

© International Union Against Tuberculosis and Lung Disease (IUATLD)

March 2002

All rights reserved

No part of this publication may be reproduced without the prior permission of the authors, and the publisher.

ISBN : 2-914365-11-X

Table of contents

Acknowledgments.	1
Preface	3
Introduction	5
Summary – the role of specific interventions	9
1. Chemotherapy	15
Essential drugs	15
Isoniazid.	17
Rifampicin	26
Pyrazinamide	35
Ethambutol	37
Streptomycin	41
Thioacetazone	45
Fixed-dose combinations	49
Principal prerequisites for an efficacious anti-tuberculosis drug	50
Early bactericidal activity	51
Sterilizing activity.	52
Ability to prevent emergence of resistance to the companion drug	53
Emergence of anti-tuberculosis drug resistance	54
Effective or functional monotherapy	55
Monotherapy during sterilization of special populations	56
Differences in bactericidal activity	57
Sub-inhibitory concentrations.	57
Differences in post-antibiotic effect (lag phase)	59
Clinical trials in the treatment of pulmonary tuberculosis	59
Streptomycin monotherapy	61
Streptomycin plus para-aminosalicylic acid	61
Streptomycin plus para-aminosalicylic acid plus isoniazid.	62
Isoniazid plus ethambutol	64
Isoniazid plus thioacetazone	65
Isoniazid plus rifampicin	66

Isoniazid plus rifampicin plus pyrazinamide (plus a fourth drug)	66
Rifampicin-containing continuation phase	67
Non-rifampicin-containing continuation phase	68
Intermittent regimens	68
Treatment regimens of less than six months' duration	70
Clinical trials in extrapulmonary tuberculosis	71
Tuberculosis of peripheral lymph nodes	71
Tuberculosis of the spine	72
Tuberculosis of the central nervous system	73
Influence of HIV infection on the choice of a regimen	74
Adverse drug events	74
Treatment efficacy	75
Influence of isoniazid resistance on the choice of a regimen	77
Influence of isoniazid plus rifampicin resistance on the choice of a regimen	78
Strategic considerations, indications, and recommendations for the choice	
of treatment regimens in a national tuberculosis control program	78
Choice of first-line regimen.	79
8-month regimens.	80
6-month regimens.	80
12-month regimens	81
Choice of re-treatment regimen	81
Treatment of patients with organisms resistant to isoniazid and rifampicin.	82
Case holding.	84
Directly observed treatment.	84
Can emergence of drug resistance be outpaced in a national tuberculosis	0.
program?	84
The approach to management of adverse drug events	86
The patient with hepatitis	87
The patient with gastrointestinal symptoms	88
The patient with impaired vision	88
The patient with vestibulo-cochlear toxicity	89
The patient with neurologic symptoms.	89
The patient with hypersensitivity reactions or muco-cutaneous signs	07
and symptoms of toxicity	89
The patient with hematologic abnormalities.	90
The patient with acute renal toxicity	91
The patient with osteo-articular pain	91
The approach to the patient with pre-existing medical conditions	91
The patient with liver injury	91
The patient with renal failure	92
The puttern multiplication and a second se	/ 4

	The patient with impaired hearing or impaired balance	92
	The patient with impaired vision	92
	The patient with gastrointestinal malabsorption	92
	The pregnant patient	93
2.	Prophylactic treatment	95
	Rationale and experiences with prophylactic treatment	95
	Indications and recommendations for the use of prophylactic treatment	96
3.	Vaccination	97
	Early vaccine development.	97
	Vaccination with <i>Mycobacterium bovis</i>	97
	Vaccination with <i>Mycobacterium chelonae</i>	97
	Vaccination with BCG	98
	Vaccine development	98
	The BCG strain family.	101
	Safety record of BCG vaccination	101
	Management of adverse reactions due to BCG vaccination	104
	Efficacy and effectiveness of BCG vaccination	104
	Prospective and retrospective studies on BCG vaccination	107
	Protection conferred by BCG vaccination against disseminated	
	and meningeal tuberculosis, and against death from tuberculosis	108
	Protection conferred by BCG vaccination of newborns and infants	109
	Protection conferred by BCG vaccination of children over one year	
	of age	111
	Protection conferred by BCG vaccination among adolescents and adults	112
	Protection conferred by BCG vaccination across various age groups	114
	Hypotheses about the variation in the efficacy of BCG vaccination	115
	Differences in methodological stringency	116
	Differences in vaccine strains	117
	Differences in vaccine dose	117
	Differences in virulence of <i>M. tuberculosis</i> strains	117
	Differences in risk attributable to exogenous re-infection tuberculosis	118
	Differences in genetic make-up of vaccinees	118
	Differences in nutritional status of vaccinees	118
	Differences in prevalence of infection with environmental mycobacteria	119
	Other factors	121
	BCG revaccination	123
	Effects of BCG other than those directed against tuberculosis	123
	Indications and recommendations for the use of BCG vaccination.	123

4. Preventive chemotherapy	127
Prevention of disease in tuberculin skin test reactors	128
Prevention of disease in persons with risk factors	130
Recently acquired infection	130
Infection with the human immunodeficiency virus	131
Spontaneously healed tuberculosis with fibrotic residuals	134
Silicosis	135
Renal failure	135
Prevention of disease following cessation of preventive chemotherapy	136
Prevention of disease with different durations of treatment	136
Prevention of disease with alternatives to isoniazid	138
Rifampicin and rifampicin combinations in comparison to placebo	140
Rifampicin and rifampicin combinations in comparison to isoniazid	141
Effectiveness of preventive chemotherapy	143
Indications and recommendations for the use of preventive chemotherapy	145
Appendix 1 – Adjunctive treatment.	147
Adjunctive therapy with corticosteroids	147
Pulmonary tuberculosis	147
Extrapulmonary tuberculosis	148
Tuberculosis of serous membranes	148
Pleural tuberculosis.	148
Pericardial tuberculosis	148
Peritoneal tuberculosis	149
Meningeal tuberculosis	149
Corticosteroid treatment in other forms of tuberculosis	150
The role of surgery in the chemotherapy era	150
Surgical treatment in respiratory tract tuberculosis	151
Tuberculous pyopneumothorax	151
Pleural tuberculosis	152
Surgical treatment in tuberculosis of the spine	152
Amoundin 2. Action counts other them countied during and during alcours	
Appendix 2 – Active agents other than essential drugs and drug classes (second-line drugs)	153
Aminoglycosides (other than streptomycin)	153
Amikacin	153
Kanamycin	154
Capreomycin	154
Cycloserine	155

Quinolones	156 158 158 158 160 161
Drugs and drug classes with potential activity against M. tuberculosis	
under investigation and development	161
Acetamides	162
Amoxicillin plus clavulanic acid	162
	163
Fullerene derivatives	163
Nitroimidazopyrans	163
Oxazolidinones	164
Paromomycin	164
Phenothiazines	164
Tuberactinomycin	165
Appendix 3 – Current vaccine development strategies	167
Immunotherapy with <i>M. vaccae</i>	167
Vaccination with saprophytic (environmental) mycobacteria	168
Auxotrophs	168
DNA vaccines	168
Recombinants	169
Subunits	169
References	171

Acknowledgments

It would not have been possible to compile this review of currently available interventions without the help of numerous colleagues who dedicated time and effort to review chapters that touched upon their specific field of expertise.

Particular appreciation goes to Richard J O'Brien, Donald A Enarson, and Arnaud Trébucq who reviewed the entire monograph, offered critical advise, proposed corrections and, most importantly, suggested improvements in the structure and flow of argumentation. Thuridur Arnadottir, José A Caminero, Bernard Fourie, Denis A Mitchison, Mario C Raviglione, Victoria Romanus, Dean E Schraufnagel, Amalio Telenti, and Jean-Pierre Zellweger made specific comments on various chapters, pointed out errors and inaccuracies, and provided additional references to complete the picture. Anne Fanning, Giovanni-Battista Migliori, Robert Loddenkemper, and Li-Xing Zhang offered helpful comments on the manuscript. Masashi Suchi provided access to some Japanese literature and offered translation of relevant parts thereof. Uwe Molkentin unearthed literature to close gaps on the understanding of the Lübeck disaster.

Clare Pierard provided editorial assistance in improving the readability of the manuscript.

Preface

This monograph adds a module to the series on the scientific basis of tuberculosis control (figure 1).¹ The International Tuberculosis Courses of the International Union Against Tuberculosis and Lung Disease (IUATLD) follow a logical sequence with these five modules. These courses are directed principally at managers of national tuberculosis control programs, largely in low-income countries.

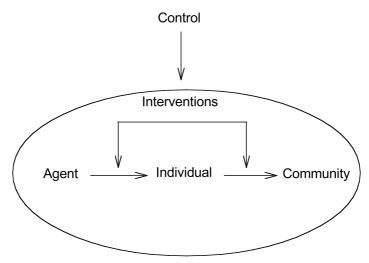


Figure 1. Modules and flow of teaching in the international tuberculosis courses of the IUATLD.¹

The courses start with the bacteriologic basis of tuberculosis control, for which several documents are available.²⁻⁶ In a second module, the effect of tubercle bacilli on the human host, the clinical presentation and diagnosis of tuberculosis is presented for which the book by Sir John Crofton and collaborators is used as background material.⁷ Following this second module, the impact of tubercle bacilli on the community is presented, i.e., the epidemiologic basis of tuberculosis control.¹ Once these three facets – the agent, the individual, and the community – are understood, the various available interventions are discussed, and finally it needs to be specifically demonstrated how this knowledge can be integrated into the practice of a national tuberculosis control program.⁸ The time allotted to each of these modules is determined by the requirements of the audience.

This monograph deals with the fourth module, interventions directed against *Mycobacterium tuberculosis* complex. There are numerous excellent reviews on the various available interventions. Often, they deal with one specific intervention. This monograph tries to assemble information about each available intervention and to weigh the role of each in current practice. Appendices provide additional information on current developments. The presentation should make it easy for the reader to select individual chapters of particular interest.

It is hoped that this module offers the review of currently available interventions that participants have been requesting since the inception of the IUATLD courses.

Paris, November 2001

Introduction

The aim of interventions in tuberculosis control or elimination strategies is to reduce or eliminate the adverse impact of epidemiological risk factors that promote the progression from one step to the next in the pathogenetically based model (figure 2).⁹

There are four principal interventions at our disposal to accomplish this task (figure 3): 10

- Treatment of tuberculosis reduces the risk of death from tuberculosis, aims at restoring health and curing patients, and reduces the risk of transmission of tubercle bacilli in the community.
- Prophylactic treatment aims at preventing infection with *Mycobacterium tuberculosis* from occurring.
- Vaccination with Bacille Calmette-Guérin (BCG) before acquisition of infection with *M. tuberculosis* aims at priming the immune system, so that the risk of progression from sub-clinical, latent tuberculous infection to clinically overt tuberculosis is reduced should such infection be acquired.
- Preventive chemotherapy is treatment of sub-clinical, latent *Mycobacterium tuberculosis* populations in the human host, given to reduce the risk of progression to clinically overt tuberculosis.

The key to improving the epidemiologic situation is linked to the specifics of the transmission (incidence) and prevalence of infection with *M. tuber-culosis.*¹ This has consequences for the role of the interventions.

The principal strategy aims at reducing the incidence infection with M. tuberculosis. A reduction in incidence of tuberculous infection is achieved by as swift as possible identification of potential transmitters of tubercle bacilli, i.e., the identification of persons with tuberculosis of the respiratory tract. Amongst these the most infectious are the patients with such a high bacillary load that the bacilli can be identified using sputum smear microscopy. While these patients account for only roughly half of all cases of pulmonary tuberculosis, they are the most potent sources of

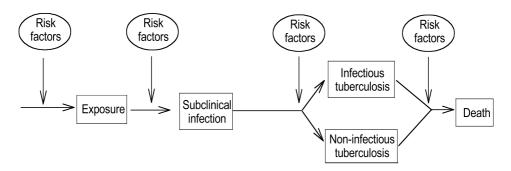


Figure 2. Pathogenetically-based model of the epidemiology of tuberculosis.¹ Reproduced from⁹ by the permission of the publisher Urban & Vogel.

transmission (figure 4).¹¹ Once identified, they should be quickly and permanently rendered non-infectious through chemotherapy. In the terminology used in this monograph, this approach is called "tuberculosis control". Thus, tuberculosis control is the strategy aimed at reducing the incidence of tuberculous infection. This strategy also includes prophylactic treatment, defined as the provision of chemotherapy to persons exposed to, but not yet infected with *M. tuberculosis*.

The second strategy aims at reducing the prevalence infection with *M. tuberculosis*. Because *M. tuberculosis* probably survives in a large proportion of persons for years following acquisition, tuberculosis will continue to emerge from the pool of persons who are already infected. A strategy to reduce the prevalence of infection in the community will be designated "tuberculosis elimination strategy" in the context of this monograph. Tuberculous infection is highly prevalent in virtually every country's population, but varies demographically in important ways.¹ To be epidemiologically effective, preventive chemotherapy to reduce the prevalence of infection must target groups that are both easily identifiable and that potentially contribute a large fraction of future morbidity.

Vaccination with BCG varies somewhat from this concept, as it aims at reducing the risk of progression from infection to disease. Consequently, its effect is expected to be similar to that of the strategy to reduce the prevalence of infection.

The options available to address the tuberculosis problem in a community will first aim at reducing the incidence of tuberculous infection (case-finding and treatment of the most infectious cases, supplemented by prophylactic treatment of special populations). Where this has been achieved and maintained over a substantial period of time, a reduction of the prevalence of tuberculous infection by preventive chemotherapy must be considered as the next logical step. Vaccination with BCG will supplement tuberculosis control efforts, particularly in high-burden countries, mainly by reducing disability and death in young children.

In this monograph, the approach chosen to discuss the various interventions follows the sequence in the epidemiological model presented elsewhere (figure 2).^{1,9} Interventions aim at reducing the impact of those risk factors recognized to promote the progression from one step to the next in the chain of events in the pathogenesis of tuberculosis (figure 3).¹⁰

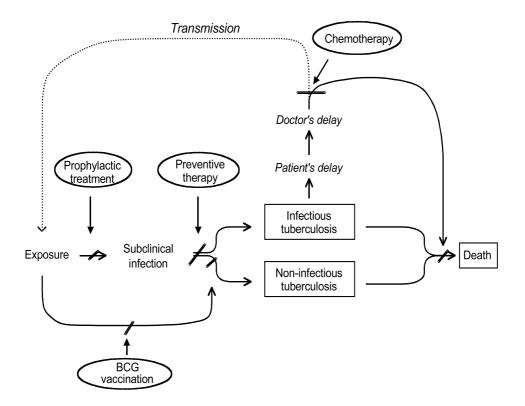


Figure 3. Model of interventions based on the epidemiology of tuberculosis. Reproduced from¹⁰ by the permission of the publisher Marcel Dekker.

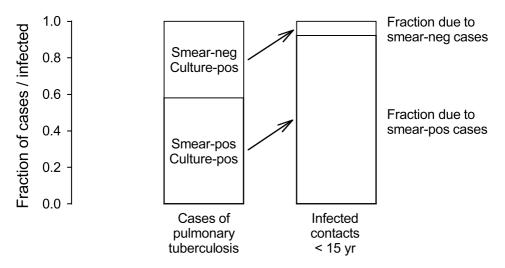


Figure 4. Sensitivity of sputum smear microscopy in identifying pulmonary tuberculosis among culture-confirmed cases and sensitivity of sputum smear microscopy in identifying transmitters of *M. tuberculosis*.¹¹

Summary – the role of specific interventions

Chemotherapy

Chemotherapy of tuberculosis is universally indicated for all newly identified tuberculosis patients. No patient with newly diagnosed tuberculosis should be denied treatment.

Chemotherapy is the most powerful weapon in tuberculosis control. It carries individual benefit by reducing morbidity and fatality. It has epidemiologic impact by cutting the chain of transmission effectively if effective treatment leading to cure of individual patients can be assured.

A national tuberculosis program must choose and recommend efficacious, standardized treatment regimens and must ensure both that they are administered carefully to prevent emergence of drug resistance and the cure of the patient.

The limited armamentarium of available anti-tuberculosis medications imposes particular constraints on the use of the most efficacious drugs. Regimens and their administration should be designed to prevent the emergence of chronic excretors with incurable, multidrug-resistant tuberculosis. The following six drugs are currently on the essential drug list:

- *Isoniazid* is the cornerstone of every first-line regimen. It has the most potent early bactericidal activity of all known drugs. It rarely causes adverse drug events, the most important of which is hepatic injury, which may result in hepatitis in a small fraction of patients. It interacts with several medications, but the single most important is its enhancement of the effect of anti-epileptics.
- *Rifampicin* has unique relapse-preventing properties that allowed the duration of chemotherapy to be shortened to nine months or fewer. It is a superbly tolerated drug that may, however, complicate isoniazid-associated hepatitis, mainly by supporting cholestasis. Immunologically-linked events might be serious and life-threatening, but are very rare. Rifampicin interacts with a multitude of other medications: the most important interactions in practice are reduction of the efficacy of oral

contraceptives and anti-retroviral medications, which preclude any such combination.

- *Pyrazinamide* also has particular relapse-preventing properties that have allowed the duration of required chemotherapy to be further reduced. In the currently recommended dosages, it is also a very well tolerated drug. Arthralgias are the most frequently reported adverse event that can be alleviated by the administration of acetylic salicylic acid or intermittent administration. There are no practically important interactions with other medications.
- *Ethambutol*'s main purpose is to reduce the risk of emergence of resistance to isoniazid. It is a very well tolerated drug, and optic neuritis, its main adverse drug event, occurs infrequently with the currently recommended dosages. It does not interact with any other drug, but its absorption might be reduced if patients take certain antacids.
- *Streptomycin* might add bactericidal activity to a regimen in the intensive phase and may add additional protection against the emergence of drug resistance, particularly in patients receiving a re-treatment regimen. It is reasonably well tolerated by young patients, but its vestibulo-cochlear toxicity and hypersensitivity reactions make its prolonged use an unpleasant experience for many patients. The only potentially important interaction in daily practice is that its toxic effects are enhanced by some diuretics.
- **Thioacetazone**'s main purpose is to reduce the risk of emergence of drug resistance to isoniazid and to reduce the risk of failure and relapse where there is resistance to the latter. More than any other drug it has the potential of multi-system adverse drug events. Among human immunodeficiency virus (HIV) infected patients, the most prominent is a muco-cutaneous syndrome that may progress to toxic epidermal necrolysis. This precludes its use in an increasing number of countries. No important interactions with other medications are known.

The treatment of previously untreated tuberculosis patients begins with the direct observation of intake of a four-drug regimen (isoniazid, rifampicin, pyrazinamide, and ethambutol, preferably in a fixed-dose combination tablet) during a two-month intensive phase. To facilitate the organization of directly observed therapy, treatment may be given thrice-weekly after a

two-week to one-month daily phase. The continuation phase cannot usually be directly observed, thus a non-rifampicin-containing continuation phase of six months is the rule in most low-income countries. The continuation phase associates isoniazid plus ethambutol (or, increasingly rarely, isoniazid plus thioacetazone). These drugs are usually given in one-month supplies for self-administration. Where resources permit a directly observed continuation phase and second-line drugs in case of treatment failure, the continuation phase can be shortened to four months by giving isoniazid and rifampicin throughout. Twelve-month regimens based on isoniazid plus ethambutol or isoniazid plus thioacetazone, supplemented by streptomycin during the first two months, were efficacious in patients not infected with HIV, and have been widely used in the treatment of bacteriologically unconfirmed tuberculosis. Evidence is accumulating that these twelve-month regimens result in a high relapse rate in HIV-infected patients and are thus being phased out in an increasing number of countries.

Patients presenting again with tuberculosis after prior treatment are known to have an increased risk of harboring bacilli resistant to at least isoniazid. Patients whose first-line treatment regimen did not include rifampicin can be successfully treated with a re-treatment regimen of eight months duration, containing rifampicin throughout. Patients who fail (remain or become again bacteriologically positive) on a first-line regimen containing rifampicin throughout cannot usually be treated successfully with the above re-treatment regimen, and recourse to second-line drugs must necessarily be taken. In most high-burden countries, however, the resources required to appropriately treat all patients who need such a regimen are not available.

The immediate prospects for the development of new, high-quality drugs that would have nation-wide availability, be well tolerated, and affordable are slim for most high-burden countries. Consequently, the preservation of the activity of the currently available medications, particularly isoniazid, rifampicin, and pyrazinamide, must be accorded the highest priority. Directly observed therapy reduces the risk of acquisition of drug resistance and relapse, and is thus of both individual and public health benefit.

Prophylactic treatment

The evidence for the efficacy of prophylactic treatment in preventing acquisition of tuberculous infection among persons exposed to an infectious case is scant. However, the limited evidence would suggest that a child born into a household with an infectious tuberculosis patient only recently placed on chemotherapy should receive prophylactic treatment with isoniazid, continued for probably at least three months following cessation of relevant exposure. Should the bacteriologic response of the index case be poor (failing to convert sputum smears), prophylactic treatment should probably be prolonged (or adjusted where feasible if the index case has a drug-resistant strain).

Prophylactic treatment is an individual intervention primarily to protect the child without separation from the mother. No great epidemiologic or public health impact of this measure is to be anticipated.

Vaccination

Vaccination with BCG provides considerable protection against death from tuberculosis, and the development of disseminated and meningeal tuberculosis, particularly in infants. Neonatal BCG vaccination (or as early in life as possible) is thus indicated where tuberculosis is frequent, childhood tuberculosis rarely diagnosed, and adequate contact examinations rarely feasible. There is insufficient evidence to recommend vaccination beyond infancy, or re-vaccination.

BCG vaccination is an individual measure that is not expected to improve the epidemiologic situation in a country. It is of public health importance to the extent that it reduces disability and death from tuberculosis in the target population.

Preventive chemotherapy

Preventive chemotherapy using nine to twelve months of isoniazid is efficacious but operationally inefficient. In adults it carries the danger of monotherapy of clinically active tuberculosis which might not be recognized if mycobacterial culture facilities and chest radiography are not routinely available. This is of particular concern in HIV infected patients who would be most likely to benefit, because such patients frequently have active tuberculosis that cannot be identified on sputum smear microscopy alone.

The drug of choice is isoniazid, although shorter regimens based on rifampicin can be used where resources permit. Logistically and adminis-

tratively easiest, and also of least concern for the development of drug resistance, is preventive chemotherapy for asymptomatic children under the age of five years who live in the household (not all of whom are necessarily infected) of a newly identified sputum smear-positive index case. It may be pragmatic to adjust the duration of isoniazid preventive chemotherapy in such cases to the length of treatment for the adult index case.

Preventive chemotherapy is an individual intervention, not shown to have as great an epidemiologic impact as chemotherapy of tuberculosis. Even if safeguards could be taken to prevent inadvertent monotherapy for patients with active tuberculosis, it remains an inefficient tool that reaches only a fraction of persons infected with *M. tuberculosis*.

1. Chemotherapy

The primary intervention must aim at reducing the incidence of infection with *M. tuberculosis*.¹² Subsequent events will reflect what happens if this primary prevention has not been properly applied. Efficacious and effective chemotherapy for patients transmitting tubercle bacilli is thus paramount to the success of a national tuberculosis program. The following major areas of concern are addressed in this chapter:

- The absolute prerequisite to effective chemotherapy is the availability of high-quality anti-tuberculosis drugs. With these drugs, optimum combination regimens have been identified. Regimens must be prescribed in a way that simultaneously prevents the emergence of resistant strains and cures the affected patient.
- Regimens suitable for use in national tuberculosis programs have been identified. The HIV epidemic has complicated tuberculosis control in general and chemotherapy in particular, and not all issues relating to treatment in the presence of HIV infection have yet been resolved.
- Administering chemotherapy through self-administered medication often gives poor results. Directly observed therapy, sometimes incorporating intermittent administration, increases the chances for a successful treatment outcome and has been shown to reduce the chance of emergence of drug resistant populations of bacilli.
- While the course of chemotherapy is uneventful in most patients and leads to complete restoration of health, some patients experience adverse drug events that need to be addressed without compromising the efficacy of treatment.

Essential drugs

There are six essential drugs that are active against *M. tuberculosis*: isoniazid, rifampicin, pyrazinamide, ethambutol, streptomycin, and thioacetazone.¹³ For each essential drug with activity against *M. tuberculosis*, a standard summary of the major aspects of interest is presented. These include, notably:

Discovery. A brief history of the discovery of the drug.

Activity, mechanism of action and resistance. Activity, mechanism of action, and mechanisms that allow *M. tuberculosis* to become resistant to anti-tuberculosis agents are intrinsically linked. In contrast to many other microorganisms, the susceptibility of virtually all wild strains of *M. tuberculosis* to the major anti-tuberculosis agents is identical. Any apparent variation in susceptibility is a misconception due to technical errors of the method used to demonstrate it. This means that an approach using the minimum inhibitory concentration (MIC) of the initial strain (in the absence of resistance) as a guide to treatment is theoretically invalid.

Pharmacokinetics. In treatment of tuberculosis (as in other diseases), concentration of the drug in the target organ determines whether the drug will have the desired effect or not. The maximum concentration that can be achieved is that which provides the highest concentration and the longest period the drug is above the MIC without being toxic.¹⁴ There are four key pharmacokinetic parameters that are of particular interest (table 1):

- C_{max} : The maximum serum concentration of the drug that can be achieved;
- T_{max} : the point in time when the maximum serum concentration is achieved following administration of the drug;
- AUC: the area under the serum concentration-versus-time curve. This is an informative parameter that summarizes the overall avail-

	Pharmacokinetic parameters in serum			
Drug	C _{max} (mg/L)	T _{max} (h)	$\begin{array}{c} \mathrm{AUC}_{0\text{-}\infty}\\ (\mathrm{mg}\times\mathrm{hr/L}) \end{array}$	
Isoniazid	4	1-2	17	2-3
Rifampicin	14	1-3	71	2-4
Pyrazinamide (1,500 mg)	25-30	1.2	420	10
Streptomycin (1 g)	25-50	1-2		2-3
Ethambutol (25 mg/kg)	5	3	30	12
Thioacetazone (150 mg)	1.8	4		13

Table 1. Pharmacokinetic parameters (rounded values) of essential anti-tuberculosis drugs.

ability of the drug in the serum of the person to whom the drug is administered;

• $\beta t_{1/2}$: the serum elimination half-life (in hours) of the drug. It indicates the time required to reduce the blood serum (or plasma) concentration to half of its maximum value.

Dosage. This is the recommended dosage in the treatment of tuberculosis in daily or thrice-weekly treatment. Because neither WHO nor the IUATLD recommend twice-weekly treatment, the dosages recommended for such an administration schedule are not presented.

Adverse drug events. No drug is without side effects or adverse drug events. Four types of adverse drug events might be distinguished:¹⁵ 1) toxic, 2) idiosyncratic, 3) hypersensitivity reactions, and 4) adverse drug events that cannot be classified into one of the three preceding groups. Toxic reactions are effects that will occur in the majority of patients at a given dose. Idiosyncrasy denotes an individual genetic defect that causes a qualitative abnormal response.¹⁶ Hypersensitivity reactions are untoward immunologic reactions to a drug.

Interactions. Some drugs interact with other medications. Such interactions are listed here, to the extent known.

Isoniazid

Discovery. Isoniazid was synthesized in 1912 at the German University of Prague by Meyer and Mally (figure 5).¹⁷ In 1952 it was independently re-discovered by the Bayer Laboratories in Germany,¹⁸ Hoffmann-La Roche

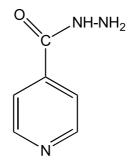


Figure 5. Chemical structure of isoniazid, synthesized by Meyer and Malley in 1912.¹⁷

Laboratories in Switzerland / United States, ¹⁹ and Squibb Laboratories in the United States, ²⁰ without the knowledge of the other groups working on the drug.

Activity, mechanism of action and resistance. Isoniazid is only active against mycobacteria. Within the genus, its effect is mainly against *M. tuberculosis* complex and to a lesser extent against a few species of environmental mycobacteria, e.g., *M. kansasii*. The MIC of *M. tuberculosis* is 0.025 to 0.05 mg/L in broth and 0.1 to 0.2 mg/L in agar plates, ²¹⁻²³ showing the uncertainty surrounding MIC determinations. Isoniazid has the most potent early bactericidal activity of all of the anti-tuberculosis drugs. ²⁴⁻²⁷ Adding other drugs will not increase this activity. ^{24,25} Thus, the rapid reduction in infectiousness observed following initiation of chemotherapy ²⁸⁻³⁰ is most likely attributable to a considerable extent to the bactericidal activity of isoniazid.

Early reports have suggested that the effect of isoniazid is on cell wall integrity. It was observed that acid-fastness was lost shortly after treatment with isoniazid.³¹ Winder and Collins demonstrated that isoniazid inhibits the synthesis of mycolic acids.³² Sacchettini and Blanchard³³ have traced the history of development in the understanding of the mechanisms of action of isoniazid. The next step in understanding was the direct correlation between isoniazid uptake, viability and mycolic acid biosynthesis.^{34,35} A specific inhibitory effect was observed on the synthesis of saturated fatty acids greater than 26 carbons, ³⁶ and the synthesis of monounsaturated longchain fatty acids in vivo.³⁷ These and subsequent observations strongly implicated enzymatic steps in the elongation of fatty acids, and the biosynthesis of the very long fatty acyl chains of mycolic acids as the site of action of isoniazid.³³ Early studies by Middlebrook *et al.* and others noted the correlation between resistance and loss of catalase-peroxidase activity.³⁸⁻⁴⁰ The molecular basis for these early observations has now been documented with the demonstration that isoniazid-resistant strains could be sensitized to the drug by transformation with the *M. tuberculosis katG*-encoded catalase peroxidase.^{41,42} Additional evidence in support of the role of catalase-peroxidase stems from the observation that deletions and missense mutations within the katG gene are common in isoniazid-resistant clinical isolates of M. tuberculosis. 43,44

The current concept classifies isoniazid as a pro-drug which requires the *katG* gene product for activation by the catalase, 33,45 targeting the last steps in mycolic acid synthesis. ⁴⁶ While details of the mode of action still

remain elusive, ⁴⁷ the general mechanism of action is fairly well understood (figure 6).⁴⁶ Several mutations have been identified which confer resistance in *M. tuberculosis*. Important mutations are located on the *katG* gene, ⁴¹ and the *inhA* gene, ⁴⁸ of which the latter is responsible for approximately 25% of clinical isolates that demonstrate resistance, generally associated with low-levels of resistance. Susceptibility to isoniazid is dependent on the presence of the catalase-peroxidase enzyme encoded by the *katG* gene.^{44,49} Mutations in catalase-peroxidase lead to high-level isoniazid resistance.^{41,50} The *ahpC* gene encodes the alkyl hydroperoxide reductase, and its absence leads to isoniazid resistance.⁵¹ Approximately 60% to 70% of isoniazid resistant strains carry mutations in one of several genes involved in its activation from pro-drug (*katG* and possibly *ahpC*) or in the

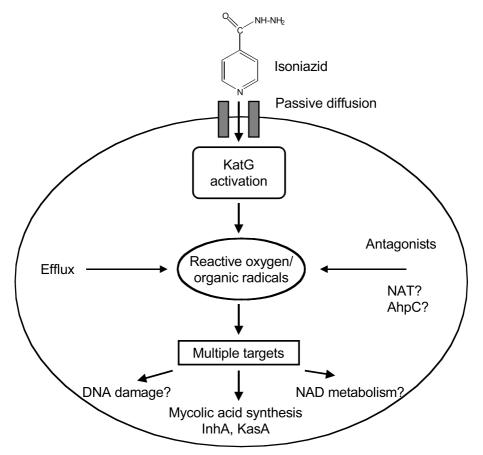


Figure 6. The proposed action of isoniazid. Reproduced from⁴⁶ by the permission of the publisher ASM Press.

drug target (*inhA*). However, the mechanism of resistance for one third of isoniazid-resistant strains remains to be elucidated.

The maximum proportion of isoniazid resistant mutants able to grow during isoniazid monotherapy of an isoniazid susceptible strain is estimated to be approximately 1 in 10^{6} . ⁵²⁻⁵⁴

Pharmacokinetics. The serum concentrations achieved by administering 300 mg isoniazid (approximately 5 mg/kg body weight) are well above the MIC (figure 7).⁵⁵⁻⁵⁷ The pharmacokinetics of isoniazid are influenced by

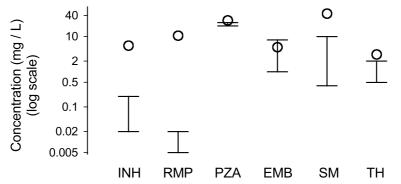


Figure 7. Maximum serum concentrations (hollow circles) and range of minimum inhibitory concentrations (lines) for isoniazid (INH), rifampicin (RMP), pyrazinamide (PZA), ethambutol (EMB), streptomycin (SM), and thioacetazone (TH).^{55,56,180,182,301,422}

acetylator type (slow versus rapid), 55 food intake, 55 and age. 57 A comparative pharmacokinetic profile of isoniazid by type of food (fasting versus high-fat meal) is shown in figure 8.55 The distribution volume of isoniazid diminishes with increasing age as shown in figure 9.⁵⁷ The elimination of isoniazid from serum is determined by the acetylator status of the individual.⁵⁸ There are three groups of acetylator types. Homozygous rapid activators are found in 40% of European and African populations and in most of those with a Mongolian ancestry. There is a heterozygous group with mutations in only one of the two alleles, and finally a homozygous group of slow inactivators with mutations in both alleles. The old "rapid inactivator group" from earlier publications consisted, in most populations, of a majority of heterozygous and a minority of homozygous rapid inactivators (Mitchison DA, personal written communication, May 22, 2001). The serum half-life, $\beta t_{1/2}$, in slow acetylators is about three hours following a dose of 5 mg/kg body weight, and about

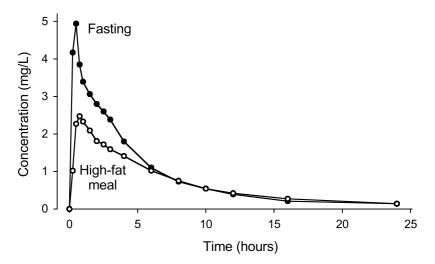


Figure 8. Pharmacokinetics of isoniazid following fasting and a high-fat meal.⁵⁵ Original data kindly provided by Charles A Peloquin.

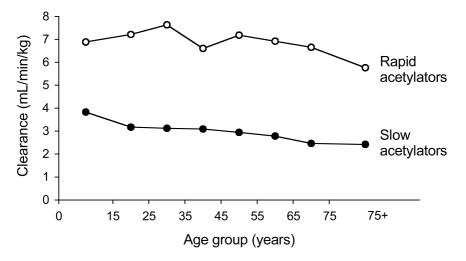


Figure 9. Clearance of isoniazid from serum, by acetylator type and age among tuberculosis patients.⁵⁷

half as long for rapid acetylators.^{59,60} The AUC is similarly affected by acetylator type which is approximately 14.2 mg/h/L in slow eliminators as compared to 2.3 mg/h/L in rapid eliminators.⁶⁰ The acetylator type is important for widely spaced intermittent therapy. It explains to a large extent why once-weekly therapy with isoniazid is particularly ineffective in rapid acetylators. For thrice-weekly treatment, the acetylator type is not impor-

tant. Isoniazid also has excellent penetration into cerebrospinal fluid, although the peak concentrations achieved are much lower in rapid than in slow acetylators. 61

Dosage. The recommended dosage of isoniazid is 5 mg/kg body weight (usually up to a maximum of 300 mg) in daily, and 10 mg/kg body weight in thrice-weekly treatment.^{8,13}

Adverse drug events (table 2). Toxic reactions include peripheral neuropathy, ⁶² seizures, ⁶²⁻⁶⁸ and other central nervous system toxic reactions, such as hallucinosis, ⁶⁹ psychosis, ⁷⁰ memory loss, ⁷¹ optic neuropathy, ⁷² and pellagra. ^{73,74} Other toxic reactions from isoniazid include pyridoxine responsive anemia, ^{15,75,76} metabolic acidosis. ⁷⁷ Pyridoxine is effective in both treatment and prevention of these reactions, ⁷⁸⁻⁸⁰ but pyridoxine non-responsive psychosis has also been reported. ⁸¹ In case of accidental or intentional overdose both charcoal treatment, if given early, ^{82,83} and hemodialysis ⁸⁴ have proved useful.

Table 2.	Summary of	f adverse read	ctions from i	soniazid with	estimated frequen-
cies of occu	urrence. No	ote that these	are estimate	es of frequenci	es which may vary
across popu	ulation group	os.			

Frequent $(\geq 5 \text{ per } 100)$	Common (\geq 1 per 100 and < 5 per 100)	Infrequent (\geq 1 per 1,000 and < 1 per 100)	Rare (< 1 per 1,000)
Liver enzyme elevations		Hepatitis Peripheral neuropathy Drug fever	Seizures Hallucinosis Psychosis Memory loss Optic neuropathy Pellagra Pyridoxine responsive anemia Metabolic acidosis Pyridoxine non-responsive psychosis Lupus erythematosus Hemolytic anemia Agranulocytosis Pure red cell aplasia Alopecia Asthma Dermatitis

Idiosyncratic reactions from isoniazid include lupus erythematosus, ^{85,86} rheumatic-like syndromes and various hematologic disorders, such as hemolytic anemia, ⁸⁷ agranulocytosis, ^{88,89} pure red cell aplasia, ⁹⁰⁻⁹² and other blood dyscrasias. ⁹³ Other rare, probably idiosyncratic reactions, include alopecia. ⁹⁴ These reactions are reported to subside promptly with with-drawal of the drug. ¹⁵

Hypersensitivity reactions from isoniazid include drug fever, ⁹⁵ asthma, ^{96,97} dermatitis, ⁹⁸⁻¹⁰⁰ and hepatitis. ^{77,101,102} Hepatotoxicity might be increased with the concomitant use of acetaminophen. ¹⁰³⁻¹⁰⁵

Clinically, the most relevant and frequent adverse drug events from isoniazid are neuropathy and liver injury.

Routine use of pyridoxine (vitamin B6) for prevention of neuropathy is not indicated.⁷⁹ Preventive treatment with small dosages of pyridoxine (6 mg/day, not to exceed 10 to 15 mg^{106,107}) is indicated for patients with increased requirements (e.g., during pregnancy), patients with nutritional deficiencies (alcoholics and aged patients), patients with a history of seizure disorder, and patients otherwise predisposed to the development of peripheral neuropathy, such as uremic patients or patients with diabetes.⁷⁹ Treatment of isoniazid-associated peripheral neuropathy (paresthesia) is with 100 to 200 mg pyridoxine per day.⁷⁹ It should be noted that there is antagonism between isoniazid and pyridoxine,¹⁰⁸ and thus the potential of inactivation of isoniazid with very high doses of pyridoxine. Therefore many clinicians prefer lower dosages (50 mg per day).

Liver enzyme elevations are frequent, but overt clinical hepatitis (with symptoms such as gastrointestinal distress, nausea, vomiting, and jaundice) occurs in less than five per cent of patients¹⁰⁹ and is age-dependent,¹¹⁰⁻¹¹⁴ and may differ in frequency in different populations,¹¹⁴ being virtually absent among, e.g., Filipinos, ¹¹⁵ and is increased in patients with pre-existent liver injury.¹¹⁰ The hepatic damage caused by isoniazid is predominantly cytolysis.¹¹⁶ The AUC for monoacetyl hydrazine, the putative precursor of the responsible agent, was greater in slow acetylators in a pharmacokinetic study, though the AUCs for acetyl isoniazid and diacetyl hydrazine were higher in rapid acetylators.¹¹⁷ The association of differences in pharamcokinetics of isoniazid and its metabolites with hepatitis risk is poorly understood,¹¹⁸ and has not been shown to be of great importance.¹¹⁹ Indeed, evidence obtained from patients in Hong Kong and Singapore showed that elevated transaminase levels during treatment with isoniazid-containing regimens were no higher in rapid than in slow acetylators.¹²⁰⁻¹²² In the IUATLD trial on preventive chemotherapy with isoniazid in patients with fibrotic lesions, ¹²³ the risk of hepatitis due to isoniazid alone could be assessed. Among patients receiving isoniazid for one year, the risk of hepatitis was 5.8 per 1,000 persons. ¹¹⁰ It increased from 2.8 per 1,000 for subjects aged less than 35 years to 7.7 per 1,000 for those aged 55 years or more, but risk was much lower in those without pre-existing liver damage (figure 10). ¹¹⁰ The hepatitis risk was highest in the first two months of treatment (figure 11). In a US Public Health Service survey, the hepati-

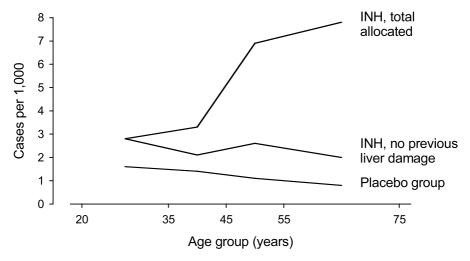


Figure 10. Hepatotoxicity from isoniazid during preventive therapy by age and pre-existing liver damage.¹¹⁰

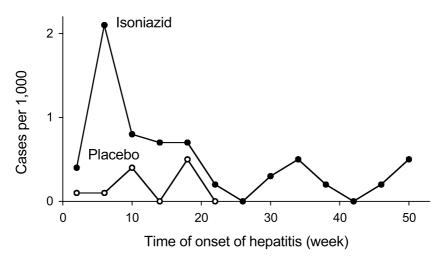


Figure 11. Hepatotoxicity from isoniazid during preventive therapy by length of exposure.¹¹⁰

tis risk per 1,000 subjects was zero for those aged less than 20 years, 2.4 for those aged 20 to 34 years, 9.2 for those aged 35 to 49 years, and 19.2 for those aged 50 to 64 years.¹¹¹ Isoniazid and rifampicin given together may potentiate the risk of hepatitis, and cases of hepatitis caused by the two drugs in combination have been reported.^{124,125}

Patients abusing alcohol may be treated with isoniazid provided they do not display signs of overt alcoholic hepatitis. Careful clinical control, limitation of alcohol consumption and (where feasible) control of liver enzymes in such patients are recommended.

Interactions. Isoniazid is an inhibitor of oxidative and demethylation metabolism as well as other cytochrome P-450 dependent microsomal pathways.^{126,127} It is also a monoamine and diamine oxidase inhibitor.^{128,129} These properties bear on the various interactions that have been reported, ¹³⁰ in that the most important interaction leads to a potentiation of the companion drug (opposite to the usual interactions seen with rifampicin).

Scombroid fishes (such as mackerel, tuna and salmon) have a high histidin content which is converted to histamine by bacteria, if improperly refrigerated. Eating such fish while taking isoniazid may lead to the typical signs of scombroid fish poisoning, with erythematous and urticarial rash, facial flushing, diarrhea, palpitations, headache, nausea, paresthesias, abdominal cramps, and dizziness.¹³¹⁻¹³⁴ It may progress to bronchospasm and hypotensison.

Certain types of cheeses rich in monoamines may also lead to hypersensitivity reactions.¹³⁵⁻¹³⁸ With wine, such reactions have also been reported.¹²⁸

Effects of isoniazid potentiated: para-aminosalicylic acid, ¹³⁹ insulin, ¹⁴⁰ carbamazepine, ¹⁴¹ valproic acid (a single report, usually the effect is the opposite), ¹⁴² theophylline. ¹⁴³

Effects of isoniazid opposed: prednisolone, ⁵⁹ ketoconazole.¹⁴⁴ After intake of ethanol, most is metabolized to acetaldehyde in the liver. Acetaldehyde appears to form acetaldehyde-adducts with isoniazid *in vitro*, and thus may lower its bioavailability, but the adduct itself may increase the toxicity of either drug.¹⁴⁵

Effect of drug potentiated by isoniazid:

• acetominophen hepatotoxicity is increased by isoniazid; ^{103,104}

- anti-coagulants such as warfarin; ¹⁴⁶
- anti-epileptics such as phenobarbital, diphenylhydantoin, ¹⁴⁷ and phenytoin, ^{148,149} carbamazepine, ¹⁵⁰⁻¹⁵³ ethosuximide, ¹⁵⁴ epanutin, ¹⁵⁵ and valproic acid; ^{142,156,157}
- anti-neoplastics such as vincristine; ¹⁵⁸
- benzodiazepines which are oxidatively metabolized (not through glucorination), ¹⁵⁹ such as diazepam¹⁶⁰ and triazolam; ¹⁶¹
- haloperidol¹⁶² and tricyclic anti-depressants;¹⁶³
- theophylline.^{143,164} The effect on theophylline pharmacokinetics might be such that even in combination with rifampicin (which has the opposite effect), theophylline clearance might be lowered, requiring a lower dose of theophylline in patients simultaneously treated with isoniazid and rifampicin.¹⁶⁵

Because of its monoamine oxidase inhibiting activity, isoniazid may potentiate the effect of monamine oxidase inhibitor anti-depressants, ^{166,167} meperidine ¹⁶⁸ and levodopa. ¹⁶⁹

Effects of drug opposed by isoniazid: enflurane, ¹⁷⁰ cyclosporine ¹⁵⁵ (disputed).

Rifampicin

Discovery. In 1957, a *Streptomyces* strain, designated strain ME / 83, later named *Streptomyces mediterranei* was isolated in the Lepetit Research Laboratories from a soil sample collected at a pine arboretum near Saint Raphaël, France.^{171,172} From this strain several rifamycins, whose structure was elucidated by Oppolzer and collaborators, ¹⁷³ were isolated. By reduction of one of these, rifamycin S and rifamycin SV were obtained. Rifamycin SV was only effective parenterally, as it was not absorbed to a significant degree when administered orally. Further chemical modification led to an orally active substance, rifampicin (figure 12).¹⁷⁴

Activity, mechanism of action and resistance. The minimum inhibitory concentration of rifampicin for *M. tuberculosis* is about 0.25 mg/L in broth

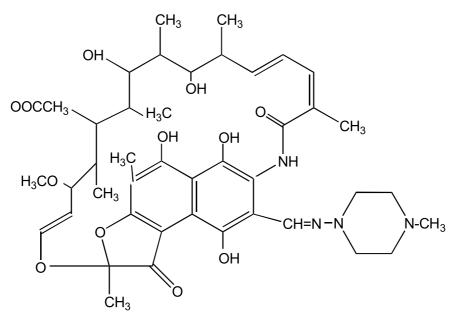


Figure 12. Chemical structure of rifampicin, isolated and semi-synthesized by Maggi and collaborators in 1966.¹⁷⁴

and 0.5 mg/L in agar.²¹⁻²³ Rifampicin is active against a wide range of micro-organisms including *M. leprae*, *S. aureus*, *N. meningitidis*, and *L. pneumophila*.

Rifampicin, like all naphthalenic ansamycins (the class to which rifampicin belongs), is a specific inhibitor of DNA-dependent RNA polymerase.¹⁷⁵

Rifampicin acts by interfering with the synthesis of mRNA by binding to the RNA polymerase.¹⁷⁶ Mycobacteria develop resistance to rifampicin by mutations in a defined region for the RNA polymerase subunit β . Mutations in the *rpoB* gene of *M. tuberculosis* are responsible for most of the resistance.¹⁷⁷ Mutations have been found in more than 97% of resistant isolates.^{178,179}

The maximum proportion of rifampicin-resistant mutants able to grow during rifampicin monotherapy of an isoniazid-susceptible strain is estimated to be approximately 1 in 10^{8} .⁵³

Pharmacokinetics. After oral administration of rifampicin on an empty stomach, the absorption is rapid and practically complete.¹⁸⁰ With a dose of 8.1 (\pm 0.7) mg/kg body weight, a peak level of 6.3 (\pm 0.5) mg/L is achieved 3.2 hours after oral administration. After oral intake of 600 mg

rifampicin, a peak level of 12 to 14 mg/L is achieved after one to three hours (figure 13).^{181,182} The AUC (between 0 and 12 hours) is 36 mg/L/hr, and the half-life is estimated to be 4.7 (\pm 1.9) hours, ¹⁸³ but has been found to be shorter in three studies (3.8 to 4.1 hours) after a single dose of 10 mg/kg body weight.¹⁷⁵ A drug-concentration – time profile is shown in figure 49.¹⁷⁵ Rifampicin is excreted unchanged in urine and bile and is also metabolized. Its major metabolite, desacetyl-rifampicin, is excreted principally in bile, but also in urine.¹⁸⁴ It appears that there are differences between men and women in the blood levels achieved, with women achieving significantly higher levels than men, a difference not explained by differences in body weight.¹⁸⁵ The pharmacokinetics of rifampicin are influenced by meals, 186,187 but depend on the type of constituents. Carbohydrates and proteins seem to have virtually no influence, while a fatty meal reduces serum concentrations considerably, as shown in four groups of 35 patients each (figure 14).¹⁸⁸ The major differences in pharmacokinetics following a meal include a reduced total amount absorbed (area under the curve) and delayed achievement of peak serum levels.¹⁷⁵

Tissue penetration of rifampicin is excellent into cavity linings, lung parenchyma and kidneys, with levels above the serum levels (figure 15).¹⁷⁵ Levels below the serum levels but well above the minimum inhibitory concentration are achieved in pyogenic bone lesions and the pleura. Critical

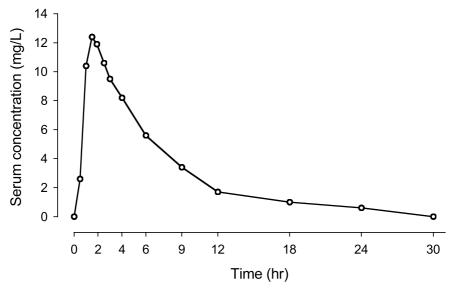


Figure 13. Pharmacokinetics of rifampicin in healthy volunteers. Reproduced from¹⁸¹ by the permission of the publisher ASM Press.

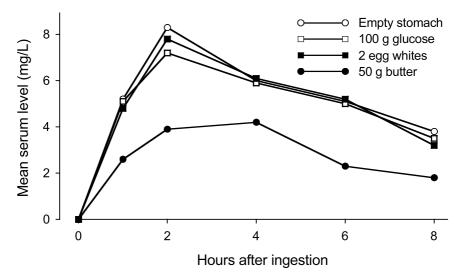


Figure 14. Pharmacokinetics of rifampicin following meals compared to fasting. Reproduced from¹⁸⁸ by the permission of the publisher Churchill Livingstone.

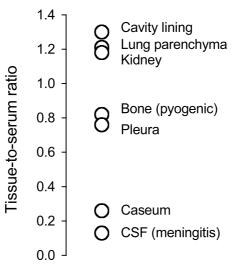


Figure 15. Tissue penetration of rifampicin with tissue-to-serum ratios.¹⁷⁵

concentrations close to the minimum inhibitory concentration were measured in caseum and cerebrospinal fluid in meningitis.

The cerebrospinal fluid to plasma concentration level ratio is between 0.52 and 1.17 over 12 hours in the experimental (healthy) rabbit model.¹⁸⁹ In comparative studies, mean peak rifampicin concentrations in the cerebrospinal fluid of patients with tuberculous meningitis of 2.4 mg/L were

obtained six hours after administration of 600 mg rifampicin, while a delayed mean peak of only 0.81 mg/L was reached nine hours after drug ingestion in normal subjects. 175

The quality of rifampicin is very susceptible to the manufacturing process. The same amount on a weight basis can lead to markedly reduced bioavailability if the particle size in the manufacturing process or the excipient are changed.¹⁹⁰ A particularly critical issue in the manufacture of rifampicin is its crystalline structure, which might be affected during the mixing process (particularly if there is a failure to properly control temperature and the grinding process), especially in fixed-dose combination preparations.¹⁹¹⁻¹⁹³

Dosage. The recommended dosage of rifampicin is 10 mg/kg body weight in daily treatment.¹³ The recommended dosage in thrice-weekly treatment is the same as the daily dosage, because an increased frequency of a flulike syndrome has been observed with intermittent treatment at higher dosages.¹⁹⁴

Adverse drug events (table 3). Serum bilirubin levels increase above normal values with the usual dosage of rifampicin on the first day of treatment, but normalize within two weeks (figure 16).^{195,196} The most frequent hepatic abnormality caused by rifampicin is cholestasis. Rifampicin induces isoniazid hydrolase, leading to increased formation of hydrazine, a finding that could explain the increased hepatotoxicity observed in patients receiving both rifampicin and isoniazid (figure 17).^{124,197,198} Hepatitis as a result of combination therapy with rifampicin and isoniazid is the most important adverse drug event in adults ^{120,194,199,200} and occurs also in children, albeit less frequently.^{109,201-203} Patients with HIV infection appear to be at particularly high risk of hepatotoxicity.²⁰⁴⁻²⁰⁶ Whether hepatitis B HBsAg carriers are at increased risk appears to be equivocal.^{207,208} In contrast, hepatitis.²⁰⁶

Rifampicin has been reported to cause acute interstitial nephritis $^{\rm 209}$ and glomerulonephritis. $^{\rm 210}$

Hypersensitivity reactions are infrequent or rare, and include pruritus,²¹¹ and rarely severe muco-cutaneous toxicity, such as toxic epidermal necrolysis,²¹²⁻²¹⁴ particularly in HIV-infected patients.^{215,216}

Rifampicin may cause menstrual disturbances such as oligomenorrhea and amenorrhea.²¹⁷ Anaphylactic shock has been reported among HIVinfected patients.^{218,219}

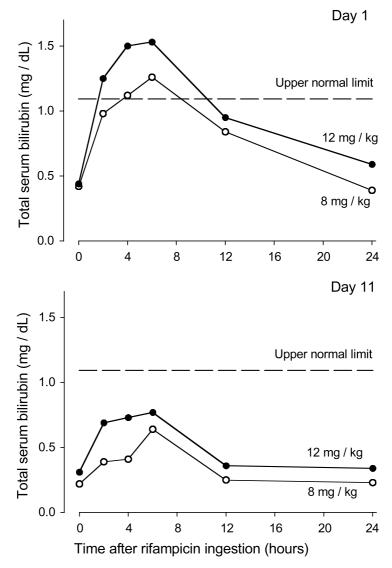


Figure 16. Total serum bilirubin levels in adults with normal liver function after ingestion of rifampicin at the beginning of treatment and after two weeks. Reproduced from¹⁹⁵ by the permission of the publisher Monaldi Archives for Chest Disease.

Among hematologic abnormalities, rifampicin has been reported to cause leukopenia, ²²⁰ hemolytic crisis, ²²¹ and thrombocytopenia, ²²² the latter being perhaps one of the more frequent adverse drug events.

Rifampicin reduces pruritus in patients with primary biliary cirrhosis, similar to the effect of phenobarbitone.²²³

Table 3. Summary of adverse reactions from rifampicin with estimated frequencies of occurrence. Note that these are estimates of frequencies which may vary across population groups.

Frequent $(\geq 5 \text{ per } 100)$	Common (\geq 1 per 100 and < 5 per 100)	Infrequent (\geq 1 per 1,000 and < 1 per 100)	Rare (< 1 per 1,000)
Bilirubin elevatio in the beginnin of treatment Orange discolorat of urine and te Liver enzyme elevations	ion	Hepatitis Pruritus Flu syndrome Drug fever	Interstitial nephritis Glomerulonephritis Renal failure Toxic epidermal necrolysis Oligomenorrhea Amenorrhea Anaphylactic shock Neutropenia Leukopenia Hemolytic anemia Pseudomembranous colitis Eosinophilic colitis Lupus erythematosus Myopathy

* Not an adverse drug event, but a normal occurrence that might cause anxiety in patients.

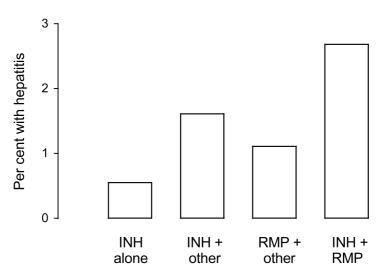


Figure 17. Hepatitis frequency following isoniazid alone or in combination with rifampicin.¹⁹⁸

Rifampicin may rarely cause pseudomembranous colitis $^{\rm 224,225}$ and eosinophilc colitis. $^{\rm 226}$

Rifampicin has also been reported to induce lupus erythematosus.²²⁷ Rifampicin has also been reported to cause myopathy.²²⁸

With intermittent therapy, when higher doses than the now-recommended daily dose equivalent have been used, a "flu" syndrome has frequently been reported.^{194,229} Also shortness of breath, hemolytic anemia, and renal failure usually occur only if rifampicin is given intermittently.¹⁹⁴

Although not an adverse drug event, it should be noted that rifampicin leads to orange discoloration of urine and tears, ²³⁰ and may permanently damage soft contact lenses. ²³¹ Intoxication leads to the "red man syndrome". ²³²⁻²³⁴

Interactions. Rifampicin is an inducer of several enzymes in the cytochrome P-450 system, ²³⁵ leading to numerous interactions with multiple drugs. This action leads most frequently to faster elimination and lower concentrations of the companion drug, an effect opposite to that seen with the most common isoniazid interactions.

No important interactions between rifampicin and other anti-tuberculosis drugs have been found, with the exception of para-aminosalicylic acid preparations¹³⁹ containing a bentonite excipient.¹⁷⁵ Rifampicin reduces the incidence of pyrazinamide-associated arthralgia, not by increasing pyrazinamide elimination, but presumably through increased excretion of uric acid.²³⁶ Numerous interactions with other medications have been described, ^{140,237-240} as detailed below.

Effect of rifampicin potentiated: Cotrimoxazole.²⁴¹ A pharmacokinetic study reported an inhibitory effect of the anti-retroviral indinavir on the metabolism of rifampicin.²⁴²

Effect of rifampicin opposed: No drug has been identified yet that opposes the action of rifampicin.

Effect of drug potentiated by rifampicin: Acetominophen hepatic failure and encephalopathy as a result of suspected potentiation by rifampicin has been reported.¹⁰⁵

Effect of drug opposed by rifampicin:

• anti-arrhythmics such as quinidine, ^{243, 244} phenytoin, ¹⁴⁸ and lorcainide; ²⁴⁵

- anti-asthmatics such as theophylline.²⁴⁶⁻²⁴⁸ The effect on theophylline pharmacokinetics might be opposed if rifampicin is given in combination with isoniazid (which has the opposite effect), so that theophylline clearance might be lowered, requiring a lower dose of theophylline in patients simultaneously treated with isoniazid and rifampicin;¹⁶⁵
- anti-coagulants such as acenocoumarol, ^{249,250} phenprocoumon, ^{251,252} and warfarin; ²⁵³⁻²⁵⁷
- anti-diabetics such as tolbutamide, ^{258,259} glidazide ²⁶⁰ or, to a lesser extent, glimeripide ²⁶¹ and glyburide; ²⁶²
- anti-fungals such as the imidazol derivatives fluconazol^{263,263} and keto-conazol;¹⁴⁴
- anti-malarials such as hydroxychloroquine²⁶⁴ and quinine²⁶⁵ and mefloquine;²⁶⁶
- antimicrobial agents such as chloramphenicol;²⁶⁷
- anti-retroviral agents such as protease inhibitors (saquinavir, ritonavir, indinavir, nelfinavir), ^{268,269} nevirapine (inconsistent), ²⁷⁰ and other antiviral agents such as zidovudine; ^{271,272}
- barbiturates such as hexobarbital; ²⁵⁹
- benzodiazepins such as diazepam; ²⁷³
- beta-blockers such as propranolol; ²⁷⁴
- calcium blockers or antagonists such as verapamil²⁷⁵⁻²⁷⁷ and nifedipine;²⁷⁸
- cardiac glycosides such as digoxin; ^{244,279,280}
- haloperidol; ¹⁶²
- hormones such as oral contraceptives, ²⁸¹ gluococorticoids, ^{282,283}, insulin, ^{284,285}, and thyroxine; ²⁸⁶
- immunosuppressants such as azathioprine, ¹⁴⁰ cyclosporin, ²⁸⁷⁻²⁹⁰ and tacrolimus; ²⁹¹

- opioids; 292-294
- vitamin K²⁹⁵, vitamin D metabolism;²⁹⁶
- sulphasalazine. 297

Pyrazinamide

Discovery. Following up on the anti-tuberculosis activity of nicotinamide (a vitamin B_3 precursor), further experimentation led to the synthesis of pyrazinamide by Kushner at the Lederle Laboratories, communicated in 1952 (figure 18)²⁹⁸ and by Solotorovski at the Merck laboratories in the same year.²⁹⁹ The synthesis of pyrazinoic acid, the active metabolite of pyrazinamide, had already been patented in 1934.³⁰⁰

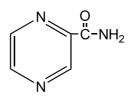


Figure 18. Chemical structure of pyrazinamide, synthesized by Kushner and collaborators in 1952.²⁹⁸

Activity, mechanism of action and resistance. Pyrazinamide is only active against mycobacteria, and among the genus, mycobacteria other than *M. tuberculosis* (including *M. bovis*) are naturally resistant.³⁰¹ It was recognized early on that pyrazinamide acts only in an acid environment.³⁰² The active derivative of pyrazinamide is pyrazinoic acid, which is preferentially accumulated in an acidic pH.³⁰³ Pyrazinamide itself is not active against intracellularly growing *M. tuberculosis*:³⁰⁴ only the accumulation of pyrazinoic acid through the action of the amidase pyrazinamidase by susceptible *M. tuberculosis* leads to its intracellular bactericidal action.³⁰⁵

The presence of both a functional pyrazinamidase and pyrazinamide transport system into *M. tuberculosis* have been postulated as prerequisites for drug susceptibility.³⁰⁶ Relatively little is known about the actual drug target, although the NAD metabolic pathway has been postulated as one of the potential targets.³⁰⁷

Mutations in *pncA*, a gene encoding pyrazinamidase, cause resistance to pyrazinamide.^{308,309} Resistance against pyrazinamide appears to develop rapidly if given as a single effective agent.³¹⁰ *M. bovis* is naturally resistant to pyrazinamide.³¹¹

Pharmacokinetics. After oral intake of 1500 mg of pyrazinamide, a peak level of 25 to 30 mg/L is achieved after one to one and a half hours (figure 19).¹⁸¹ Pyrazinamide has one of the best penetrations into cerebrospinal fluid among the anti-tuberculosis medications.^{312,313} About four per cent of pyrazinamide is excreted unchanged in urine and about 30% as pyrazinoic acid.³¹⁴ It is only slightly influenced by ingestion of antacids, but with a fatty meal T_{max} is delayed and C_{max} slightly lowered, although these effects are unlikely to bear clinical relevance.³¹⁵ Absorption of pyrazinamide is not influenced by food intake.

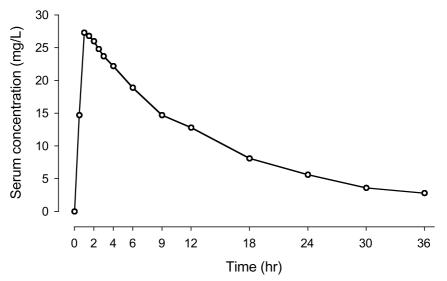


Figure 19. Pharmacokinetics of pyrazinamide in healthy volunteers. Reproduced from¹⁸¹ by the permission of the publisher ASM Press.

Dosage. The dosages of pyrazinamide have varied greatly since its introduction. In early periods, the usual dosage was around 40 to 50 mg/kg body weight^{316,317} and up to eight grams were given per day.³¹⁰ Such dosages frequently resulted in hepatotoxicity³¹⁷ and its early withdrawal from chemotherapy regimens. The current recommendations are to give 25 mg/kg body weight per day.^{8,13}

Adverse drug events (table 4). The two major adverse drug events of pyrazinamide are hepatotoxicity ^{115,120,200,310,317-324} and interference with the metabolism of purine. The latter leads to decreased excretion and accumulation of uric acid, occasionally accompanied by gout-like arthralgia. ^{310,325,326} The suppressive effect of pyrazinoic acid on uric acid excretion is maximal for **Table 4.** Summary of adverse reactions from pyrazinamide with estimated frequencies of occurrence. Note that these are estimates of frequencies which may vary across population groups.

Frequent $(\geq 5 \text{ per } 100)$	Common (\geq 1 per 100 and < 5 per 100)	Infrequent (\geq 1 per 1,000 and < 1 per 100)	Rare (< 1 per 1,000)
Arthralgias	Nausea	Hepatitis Rash Nausea	Sideroblastic anemia Lupus erythematosus Convulsions Photodermatitis

24 hours.³²⁷ Thus, uric acid retention could be reduced by intermittent administration.

Relatively frequent events include rash^{200,328,329} and nausea.²⁰⁰ Rarer adverse drug events include sideroblastic anemia,^{75,93} lupus erythematosus,³³⁰ convulsions,³³¹ and photodermatitis.³³²

Interactions

Effect of pyrazinamide potentiated: Allopurinol increases plasma concentrations of pyrazinoic acid which is directly responsible for the inhibition of renal urate secretion.³³³ Therefore, pyrazinamide-induced arthralgias are unresponsive to allopurinol.

Effect of pyrazinamide opposed: A potentially serious interaction may exist with zidovudin, with combination treatment leading to barely detectable pyrazinamide levels.³³⁴ However, these findings have not been confirmed.

Effect of drug potentiated by pyrazinamide: None identified.

Effect of drug opposed by pyrazinamide: Pyrazinamide might antagonistically affect the action of medications that have a uricosuric effect such as acetylic salicylic acid, ascorbic acid, probenecid, and iodine containing radio-contrast offering preparations.^{335,336}

Ethambutol

Discovery. The synthesis of ethambutol (figure 20) was reported in 1961.³³⁷ Its excellent activity *in vitro* and *in vivo* against *M. tuberculosis* was reported in the same year.³³⁸⁻³⁴⁰

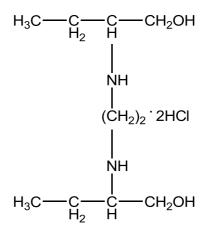


Figure 20. Chemical structure of ethambutol, reported by Thomas and collaborators in 1961.³³⁷

Activity, mechanism of action and resistance. Ethambutol is only active against mycobacteria.³⁴⁰ Ethambutol is bactericidal on both extracellular and intracellular tubercle bacilli.³⁴¹ The MIC of ethambutol for *M. tuberculosis* is about 0.95 to 7.5 mg/L in broth and 1.9 to 7.5 mg/L on agar.^{21,23}

Ethambutol specifically inhibits biosynthesis of the mycobacterial cell wall.⁴⁶ It acts on the biosynthesis of arabinogalactan, the major polysaccharide of the mycobacterial cell wall.³⁴² It inhibits the polymerization of cell wall arabinogalactan and of lipoarabinomannan.^{343,344} It indirectly inhibits mycolic acid synthesis (by limiting the availability of arabinan for the mycolic acids to attach to)³⁴⁵ and triggers a cascade of changes in the lipid metabolism of mycobacteria, leading to the disaggregation of bacteria clumps into smaller clusters.³⁴⁶ It appears to be able to break down the "exclusion barrier" located in the *M. avium* cell wall and thus significantly enables the activity of other drugs, both intracellularly and extracellularly.^{347,348}

The maximum proportion of ethambutol-resistant mutants able to grow during ethambutol monotherapy of an isoniazid-susceptible strain is estimated to be approximately 1 in 10⁸.⁵³

Pharmacokinetics. The absorption of ethambutol is rapid. Following a dosage of 25 mg/kg body weight, a peak serum concentration of 4 to 5 mg/L is achieved approximately two to four hours after administration (figure 21). ^{349,350} The drug is not extensively metabolized. Roughly 80% of ethambutol is eliminated by glomerular filtration and tubular secretion. ³⁴⁹⁻³⁵¹

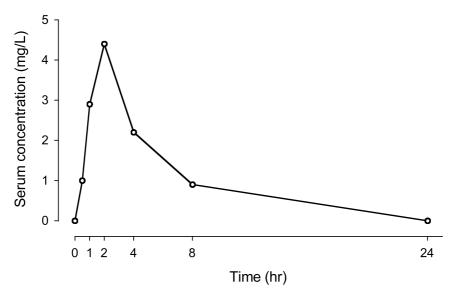


Figure 21. Pharmacokinetics of ethambutol in healthy volunteers. Reproduced from³⁴⁹ by the permission of the publisher American Thoracic Society at the American Lung Association.

Ethambutol penetrates tissues rapidly and in high concentrations, including lung, liver and kidney in experimental tuberculosis.^{352,353} It has poor penetration into cerebrospinal fluid and brain.³⁵² Renal failure decreases body clearance and increases serum half-life, and dose adjustment in such patients is mandatory.³⁵⁴ High-fat meals alter the pharmacokinetics of ethambutol somewhat, but probably not importantly.³⁵⁵

Dosage. Although 15 mg/kg body weight in the continuation phase and 25 mg/kg body weight in the intensive phase have been recommended, ³⁵⁶ international consensus recommends 15 mg/kg (range 15 to 20 mg/kg) throughout ^{8,13} to obviate operational difficulties in changing the dosage and to further reduce toxicity.

Adverse drug events (table 5). The most important adverse drug event of ethambutol is ocular toxicity, first reported in 1962, ³⁵⁷ and followed by numerous accounts.³⁵⁸⁻³⁷⁶ It is postulated that many instances of ethambutol's ocular toxicity might be explained by its binding to zinc or copper.^{152,377} Two types of ocular toxicity (optic neuropathy) from ethambutol have been described.^{372,378} The more common form is a noninflammatory axial fiber disease involving central fibers of the optic nerve.³⁷⁸ Patients with central or axial toxicity have reduced visual acuity, central scotoma,

Table 5. Summary of adverse reactions from ethambutol with estimated frequencies of occurrence. Note that these are estimates of frequencies, which may vary across population groups.

Frequent $(\geq 5 \text{ per } 100)$	Common (\geq 1 per 100 and < 5 per 100)	Infrequent (\geq 1 per 1,000 and < 1 per 100)	Rare (< 1 per 1,000)
		Retrobulbar neuritis Periaxial ocular toxicity	Aplastic anemia Eosinophilic pneumonia Thrompocytopenia Hyperuricemia

and loss of ability to see green (reported as white or gray). The ability to see red, which has been reported as pink, has occasionally been affected. Those with periaxal toxicity have a defect in the peripheral isopters of their field of vision, with little or no decrease in visual acuity and normal redgreen color discrimination. The optic discs and the fundi appeared normal in both types of toxicity. The incidence of ocular toxicity is dose dependent.^{372,379} The chance of visual recovery appears to be related to the total dose administered and the initial degree of loss of vision.³⁷⁰ It has been recommended not to use ethambutol in children too young for objective tests for visual acuity.³⁶³ There is, however, no evidence that children are particularly prone to ocular toxicity, 380 and ethambutol may thus be used in children. However, as children might be less likely to report ocular toxicity, particular caution may be warranted. Ocular toxicity is usually reversible upon cessation of ethambutol administration, but recovery might be protracted.³⁶⁸ Compared to the frequency of fatal outcome resulting from anti-tuberculosis medication, the occurrence of blindness from ethambutol is rare. 367

Ethambutol may cause aplastic anemia, ³⁶⁷ but this is exceedingly rare. Ethambutol is a rare cause of pulmonary infiltrates with eosinophilia, ³⁸¹ rash, ^{367,382} exacerbation of lupus erythematosus, ³³⁰ thrombocytopenia ³⁸³ and hyperuricemia. ³⁸⁴

Interactions

Effect of ethambutol potentiated: Although listed in some text books, ethionamide and isoniazid have not been conclusively shown to increase ethambutol ocular toxicity.

Effect of ethambutol opposed: Aluminum-magnesium antacid reduces ethambutol resorption, and lowers and delays, respectively, C_{max} and T_{max} .³⁵⁵

Effect of drug potentiated by ethambutol: None identified.

Effect of drug opposed by ethambutol: None identified.

Streptomycin

Discovery. Selman A Waksman isolated Actinomyces griseus from soil in 1916, ³⁸⁵ later termed Streptomyces griseus. ³⁸⁶ In 1939, Waksman's research group started an extensive study of substances produced by soil organisms which destroyed other soil organisms (termed antibiotics by Waksman). ³⁸⁷ The first antibiotic isolated from an Actinomyces species was actinomycin in 1940. ³⁸⁶ In 1942, streptothricin was isolated. ³⁸⁶ In September 1943 Streptomyces griseus was re-identified. ³⁸⁸ and the isolation of streptomycin was reported in January 1944 (figure 22). ³⁸⁹ It is noteworthy that the original table presenting the antimicrobial activity of streptomycin accorded a single, inconspicuous line to its effect on *M. tuberculosis* and this finding found no mention in the text (figure 23). ³⁹⁰ But in the same year Schatz and Waksman published a paper devoted particularly to the action of streptomycin on *M. tuberculosis*. ³⁹¹ In 1952, Waksman received the Nobel Prize for Physiology or Medicine. ^{386,387}

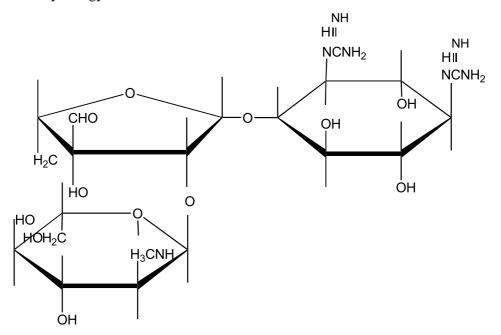


Figure 22. Chemical structure of streptomycin, isolated by Schatz, Bugie, and Waksman and reported in 1944.³⁹⁰

		Units of activity per gram ash-free dry material	
Organism	Gram stain	Streptomycin x 1000	Streptothricin x 1000
B. subtilis O	+	125	500
B. mycoides O	+	250	< 3
B. mycoides 317-911	+	20	< 3
B. mycondes 917 911 B. cereus	+	30	< 3
B. mesentericus	+	15	_
B. megatherium	+	100	150
S. aureus	+	15	200
S. lutea	+	100	150
M. phlei	+	100	50
M. tuberculosis	+	30	—
Phytomonas pruni	_	100	400
Listerella	_	10	_
Shigella gallinarum	_	_	150
E. coli	_	25	100
S. marcescens	_	25	5
A. aerogenes	_	10	50
P. vulgaris	_	10	50
S. aertycke	_	2.5	_
S. schottmülleri	_	_	50
Ps. fluorescens	_	2	< 3
Ps. aeruginosa	_	1	< 3
Cl. butylicum	_	3	< 3

TABLE III.Comparative Bacteriostatic Spectra of Streptomycin and Streptothricin.
On basis of crude, ash-free dry material.

Figure 23. Original table published when the isolation of streptomycin was reported. Reproduced from³⁸⁹ by the permission of the publisher Society for Experimental Biology and Medicine.

Activity, mechanism of action and resistance. Streptomycin has a broadspectrum activity against many gram-positive and gram-negative microorganisms and against various species of mycobacteria. Its effect on *M. tuberculosis in vitro* and in the guinea pig was reported as early as December 1944, ³⁹² and a preliminary report on its usefulness in the treatment of tuberculosis in man in September 1945 by Feldman and Hinshaw, ^{393,394} followed by a more extensive report in 1946. ³⁹⁵ The MIC of *M. tuberculosis* is 0.25 to 2.0 mg/L. ^{21,56} It had been surmised that streptomycin is active only against extracellularly growing tubercle bacilli, but this notion has not been borne out by experiments which have demonstrated its activity against bacilli residing inside macrophages as well.³⁹⁶

Streptomycin inhibits protein synthesis of *M. tuberculosis*. Streptomycin acts on ribosomes and causes misreading of the genetic code, inhibition of translation of mRNA, and aberrant proofreading.³⁹⁷

It was demonstrated a half century ago that a strain may contain different variants with different levels of susceptibility (or resistance) to streptomycin.³⁹⁸ Interestingly, problems with molecular techniques to properly identify clinically relevant resistance led some authors to conclude that the seemingly outdated use of drug-containing media described in these early reports³⁹⁸ may again become a valid procedure.³⁹⁹ Resistance results from a limited number of missense mutations in the *rrs* gene (16S rRNA) or in the *rpsL* gene (ribosomal protein S12).⁴⁰⁰

The maximum proportion of streptomycin resistant mutants able to grow during streptomycin monotherapy of an isoniazid susceptible strain is estimated to be approximately 1 in $10^{8.53}$

Pharmacokinetics. Streptomycin is not at all, or only insignificantly, absorbed from the gut and its administration is parenteral. Following intramuscular administration, resorption is rapid and maximum serum concentrations are achieved within one to two hours (figure 24).^{183,401} Streptomycin, like all aminoglycosides, is excreted by glomerular filtration. When kid-

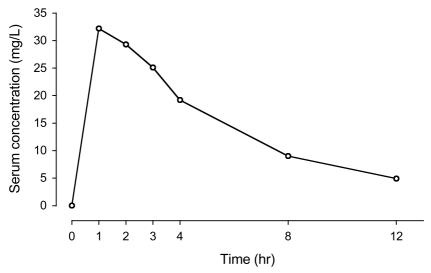


Figure 24. Pharmacokinetics of streptomycin in tuberculosis patients. Reproduced from¹⁸³ by the permission of the publisher American Thoracic Society at the American Lung Association.

ney function is impaired, the dosage must be adjusted, as excretion is exclusively renal.⁵⁶ Streptomycin has a limited ability to penetrate membranes, resulting in low concentrations of cerebrospinal fluid.⁴⁰²

Dosage. After large doses (up to three grams daily were given in the early trials ³⁹⁵) toxicity was frequent and dosage reductions were sought that would not compromise efficacy. ⁴⁰³ The current recommendation is to give 15 mg/kg body weight (range 12 to 18), ^{8, 13} with a usual maximal dose of one gram in adults. The dosage is reduced in elderly patients. It has to be administered parenterally, usually by intramuscular injection, but intravenous application is preferred by some because of higher peak but lower trough levels. ⁴⁰⁴

Adverse drug events (table 6). The main adverse effect of streptomycin is vestibulo-cochlear toxicity, which is usually ^{323,403,405} but not always, dose-dependent. ⁴⁰⁶ Hypersensitivity reactions are also relatively frequent and important, ⁵⁶ not only in patients, but also in health care personnel administering the medication. ⁴⁰⁷ Because of its penetration into amniotic fluid and its ototoxic effect on the fetus, ⁴⁰⁸ streptomycin should never be administered to pregnant women. ⁵⁶ Streptomycin may cause neuromuscular block-ade, ⁴⁰⁹ not reversed by neostigmine. ⁴¹⁰

Table 6. Summary of adverse reactions from streptomycin with estimated frequencies of occurrence. Note that these are estimates of frequencies, which may vary across population groups.

Frequent $(\ge 5 \text{ per } 100)$	Common (≥ 1 per 100 and < 5 per 100)	Infrequent (\geq 1 per 1,000 and < 1 per 100)	Rare (< 1 per 1,000)
Vestibular toxicity	Cochlear toxicity Hypersensitivity reactions	Renal damage	Neuromuscular blockade

Interactions

Effect of streptomycin potentiated: Ototoxicity of streptomycin is increased by diuretics such as furosemide⁴¹¹ and ethacrynic acid.⁴¹²

Effect of streptomycin opposed: None identified.

Effect of drug potentiated by streptomycin: Like other aminoglycosides, streptomycin has a neuromuscular blocking effect⁴¹³ and may lead to pro-

longed respiratory depression following curare-like drugs, such as pancuronium,⁴¹⁴ succinylcholine or tubocuronium⁴¹⁵ or non-depolarizing relaxants such as diallyl-nortroxiferine.⁴¹⁶

Effect of drug opposed by streptomycin: None identified.

Thioacetazone

Discovery. Freund and Schander synthesized benzaldehyde-semicarbazone in 1896 and 1902, respectively.^{417,418} From this basic compound, derivatives with anti-tuberculosis properties were later developed. After investigations on sulphonamides had revealed that thiazoles and thiodiazole derivatives exerted some activity against mycobacteria,⁴¹⁹ Domagk and collaborators at the Bayer Laboratories synthesized a new class of drugs, the thiosemicarbazones, of which thioacetazone (figure 25) was shown to be active against tubercle bacilli.⁴²⁰

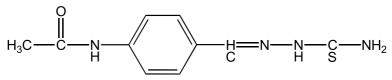


Figure 25. Chemical structure of thioacetazone, synthesized by Domagk and collaborators in 1946.⁴²⁰

Among the numerous derivatives of semicarbazones, three have found particular activity against *M. tuberculosis*: ⁴²¹

p-acetylamino-benzaldehyde-semicarbazone;

p-methoxy-benzaldehyde-semicarbazone;

p-ethylsulfone-benzaldehyde-semicarbazone.

Of these, p-acetylamino-benzaldehyde-semicarbazone, tested under the name TB I, now known as thioacetazone, became the most widely used semicarbazone.

Activity, mechanism of action and resistance. Thiosemicarbazones, including thioacetazone, are active only against mycobacteria, and favorable *in vitro* and *in vivo* results against *M. tuberculosis* were published in 1949.⁴¹⁹ The observed *in vitro* susceptibility of *M. tuberculosis* varies considerably, depending on the technique of susceptibility testing and the origin of the strain.⁴²² An observation made in a comparison of tubercle bacilli isolated from India and in the United Kingdom showed that Indian strains were considerably less susceptible to thioacetazone than strains from the United Kingdom.⁴²³ This geographic variation was subsequently confirmed.⁴²⁴⁻⁴²⁷ The susceptibility of strains may vary even within the same country.⁴²⁸ The correlation between *in vitro* and *in vivo* results is often very poor.⁴²⁹

The mode of action of thioacetazone has not been elucidated, ⁴³⁰ although it has been shown that thioacetazone forms copper complex salts and it has been postulated that these might represent the effective compound. ⁴³¹

There is partial cross-resistance between thioacetazone and ethion-amide. $^{\rm 422}$

Pharmacokinetics. Thioacetazone is rapidly absorbed and maximum serum concentrations are achieved about four hours (range two to six hours) after ingestion, ⁴³²⁻⁴³⁴ and is eliminated from serum almost completely within 24 hours (figure 26). ⁴³⁴

Dosage. The currently recommended dosage of thioacetazone is 2.5 mg/kg body weight per day.¹³ Only daily treatment is recommended.

Adverse drug events (table 7). Thioacetazone frequently causes adverse drug events, ⁴³⁵⁻⁴³⁸ which occur in up to 40% of patients. The most fre-

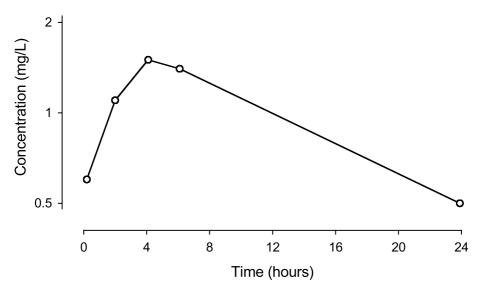


Figure 26. Pharmacokinetics of thioacetazone in healthy volunteers.434

quent adverse drug events are gastrointestinal (weight loss, nausea, vomiting),⁴²¹ central nervous system,⁴²¹ and cutaneous adverse drug events.⁴³⁵ An international investigation in 13 countries into adverse drug events due to thioacetazone was coordinated by the British Medical Research Council.⁴³⁹ The frequency of adverse drug events in that study was 21% compared to eight per cent of patients who were not receiving thioacetazone. More than half of the adverse drug events were mild. The study confirmed earlier observations that gastrointestinal and neurologic adverse drug events (headache, blurred vision, perioral numbness, mental symptoms, and peripheral nerve symptoms) were the most frequent, followed by cutaneous adverse drug events. Two out of 1,000 patients developed agranulocytosis.⁴³⁹ The frequency of adverse cutaneous reactions varied in different populations. Differences in nutrition may be a contributor to this observation as, for example, consumption of cheese and fish appear to increase the risk of cutaneous and neurologic adverse drug events.⁴⁴⁰

It was recognized relatively early that patients with HIV infection have increased susceptibility to developing toxic epidermal necrolysis when given sulfur-containing medications such as sulfadoxine⁴⁴¹ or sulfamethoxazole.⁴⁴² The causal relationship between the occurrence of cutaneous adverse reactions and the use of thioacetazone has been elegantly demonstrated (figure 27).⁴⁴³ Reactions may present as pruritus without rash, rash without epidermolysis, and most seriously as toxic epidermal necrolysis.⁴⁴⁴ The latter has a case fatality rate of 20% to 30%, depending on the selection of cases (figure 28). Both reactions and deaths occur relatively early in the course of administration, with more than half occur-

Frequent (≥ 5 per 100)	Common (\geq 1 per 100 and < 5 per 100)	Infrequent (\geq 1 per 1,000 and < 1 per 100)	Rare (< 1 per 1,000)
Weight loss Nausea Vomiting Itching Mental disturbances Headache Blurred vision Perioral numbness	Toxic epidermal necrolysis (in HIV) infected patients)	Toxic epidermal necrolysis (in non HIV) infected patients)	Agranulocytosis

Table 7. Summary of adverse reactions from thioacetazone with estimated frequencies of occurrence. Note that these are estimates of frequencies, which may vary across population groups.

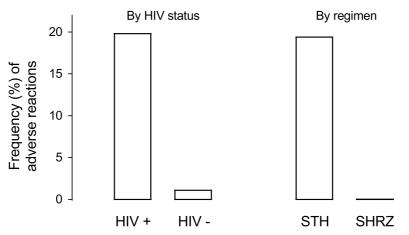


Figure 27. Demonstration of the causal relation between cutaneous adverse reaction and thioacetazone, by HIV status and regimen.⁴⁴³

ring within the first three weeks of treatment (figure 29) (Tanzania National Tuberculosis / Leprosy Programme, IUATLD, unpublished data). Numerous accounts have now been published that confirm the potential seriousness of the utilization of thioacetazone in HIV infected tuberculosis patients.⁴⁴⁵⁻⁴⁴⁸ While most adverse reactions are toxic effects, cutaneous adverse reactions appear to be largely idiosyncratic, and are not influenced by reducing the dosage.⁴²¹

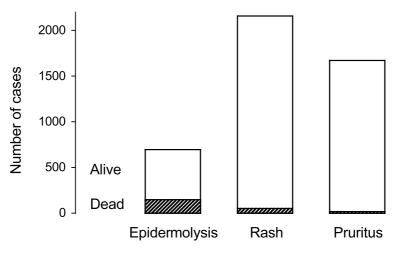


Figure 28. Adverse cutaneous reactions and deaths associated with the use of thioacetazone, by severity of reaction. Tanzania National Tuberculosis / Leprosy Programme and IUATLD, unpublished data.

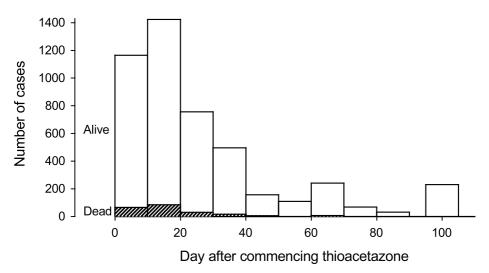


Figure 29. Adverse cutaneous reactions and deaths associated with the use of thioacetazone, by duration of thioacetazone intake. Tanzania National Tuberculosis / Leprosy Programme and IUATLD, unpublished data.

Because of this strong association, thioacetazone should never be given to patients known to be HIV infected.¹³ It is also sensible policy to relinquish its use in countries where the HIV prevalence among tuberculosis patients is known to be high.^{449,450}

Interactions. Interactions between thioacetazone and other medications are not known.

Fixed-dose combinations

Tuberculosis needs to be treated with multiple drugs. It is thus not surprising that efforts have been undertaken to develop so called fixed-dose combinations. Fixed-dose combinations simplify treatment, minimize prescription errors, and simplify supply management.^{191,451} As fixed-dose combinations containing rifampicin may be particularly prone to posing difficulties in assuring bioavailability, specific requirements have been outlined to ensure their quality.^{452,453}

The dosages of the individual components in a fixed-dose combination are of critical importance to prevent both over- and under-dosage. The WHO recommends the dosages per tablet as summarized in table 8.^{454,455}

FDC per tablet	For daily use (mg drug)	For three-times weekly use (mg drug)
{TH}	150 T + 300 H	
	50 T + 100 H *	_
{EH}	400 E + 150 H	_
RH	300 R + 150 H	_
	150 R + 75 H	150 R + 150 H
	60 R + 30 H *	60 R + 60 H
{RHZ}	150 R + 75 H + 400 Z	150 R + 150 H + 500 Z
()	60 R + 30 H + 150 Z *	_
{RHZE}	150 R + 75 H + 400 Z + 275 E	_

Table 8. Fixed-dose combinations (FDC) of antituberculosis drugs and dosages of individual drugs as recommended by WHO.⁴⁵⁴

Fixed-dose combinations will guarantee that drugs cannot be taken separately. They thus reduce the potential of acquisition of drug resistance. However, prescription errors or selective use of the number of tablets by the patient may lead to sub-inhibitory concentrations of all drugs. The need for direct observation of drug intake is thus not obviated with their introduction into national programs.

Principal prerequisites for an efficacious anti-tuberculosis drug

It is general practice to define the action of antimicrobial agents as "bacteriostatic" or "bactericidal". This terminology might not be that useful in describing the activity of anti-tuberculosis medications. Mitchison has proposed the utility of defining three prerequisites for an anti-tuberculosis drug (table 9): ⁴⁵⁶

- Early bactericidal activity;
- Sterilizing activity;
- Ability to prevent emergence of resistance to the companion drug.

Extent of activity	Prevention of resistance	Early bactericidal	Sterilizing
High	isoniazid rifampicin	isoniazid	rifampicin pyrazinamide
	ethambutol streptomycin	ethambutol rifampicin	isoniazid
Low	pyrazinamide thioacetazone	streptomycin pyrazinamide thioacetazone	streptomycin thioacetazone ethambutol

Table 9. Grading of activities of anti-tuberculosis drugs. Reproduced from ⁴⁵⁶ by the permission of the publisher Churchill Livingstone.

Early bactericidal activity

Early bactericidal activity is defined as the ability of the drug to kill tubercle bacilli in the first few days of treatment.^{24,25,456} In a study measuring sputum colony counts in newly diagnosed tuberculosis patients treated with a multitude of monotherapy and multidrug therapy regimens during the first two weeks of treatment, no other drug or drug combination was superior to isoniazid alone in the first two days of treatment (figures 30 and 31).^{24,25} This high early bactericidal activity of isoniazid was subsequently confirmed.^{26,457} It is likely that the rapid reduction in infectiousness seen in

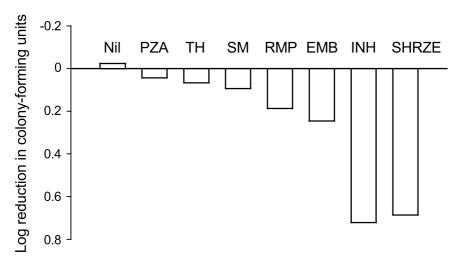


Figure 30. Early, two-day bactericidal activity of anti-tuberculosis drugs, measured as the reduction in colony-forming units in sputum.²⁴

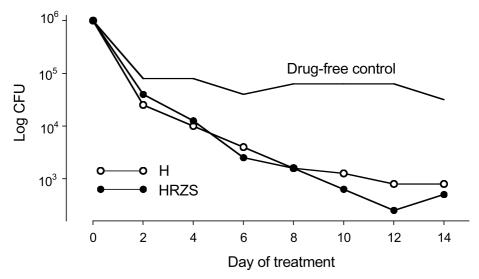
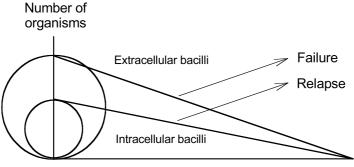


Figure 31. Bactericidal activity of isoniazid compared to a four-drug combination therapy over the first two weeks of treatment.²⁵

tuberculosis patients once placed on chemotherapy ^{29,30,458,459} is largely due to the use of isoniazid.

Sterilizing activity

Sterilizing activity is defined as the ability to remove so called "persisters", once the large bulk of rapidly growing organisms has been killed. A model presented by Grosset clarifies these two major components of chemotherapy (figure 32).⁴⁶⁰ Inability to destroy rapidly growing bacilli, located largely extracellularly, leads to treatment failure, while inability to eradicate persisters leads to relapse subsequent to treatment completion. Persisters are bacilli that have a lower metabolic activity and thus replicate much more slowly than bacilli found in cavity linings. It was postulated that the efficacy of rifampicin as a sterilizing agent was due to its activity on special populations.⁴⁶¹ This was tested in an experiment reproducing conditions appropriate for high and low metabolism of tubercle bacilli, respectively, using temperature control as the means.⁴⁶² At body temperature, there was only slightly higher activity of rifampicin over isoniazid during a seven-day period. If pulsed temperature elevation was applied for only one hour per day to increase metabolism, rifampicin was considerably more active than isoniazid (figure 33).⁴⁶²



Duration of chemotherapy

Figure 32. Schematic presentation demonstrating the mechanisms for treatment failure and disease relapse. Reproduced from⁴⁶⁰ by the permission of the publisher Excerpta Medica.

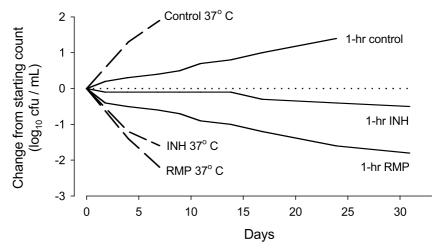


Figure 33. Comparative activity of isoniazid and rifampicin in experiments mimicking high and low metabolic activity. Reproduced from ⁴⁶² by the permission of the publisher American Thoracic Society at the American Lung Association.

Ability to prevent emergence of resistance to the companion drug

Prevention of the emergence of drug resistance is defined as the ability of a drug to prevent selection of mutants resistant to the companion drug. Not every anti-tuberculosis drug has the same ability to prevent emergence of resistance against a companion drug in clinical practice (figure 34).⁴⁶³

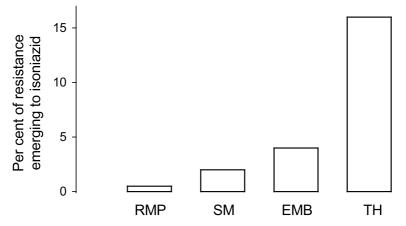


Figure 34. Ability of an anti-tuberculosis drug to prevent as a companion drug the emergence of isoniazid resistance.⁴⁶³

In summary, each anti-tuberculosis medication can be assigned a grading scale according to these three properties (table 9).⁴⁵⁶ This explains the reason for the high efficacy of chemotherapy regimens incorporating isoniazid, rifampicin, and pyrazinamide.

Emergence of anti-tuberculosis drug resistance

The most convincing evidence for the mechanism of emergence of clinically significant drug resistance is effective or functional monotherapy. This and related mechanisms are discussed in some detail.

There are several mechanisms by which tubercle bacilli may acquire resistance: ⁴⁶⁴

- Effective or functional monotherapy;
- Monotherapy during sterilization of special populations;
- Differences in bactericidal activity;
- Sub-inhibitory concentrations;
- Differences in post-antibiotic lag phase.

Effective or functional monotherapy

The first clinical trial in tuberculosis was by necessity limited to the first drug developed against tuberculosis, streptomycin. In the trial conducted by the British Medical Research Council, a total of 109 patients were admitted to the streptomycin arm.⁴⁶⁵ Serial susceptibility testing results were available among 41 of these patients, 35 of whom acquired streptomycin resistance (figure 35). As the testing interval between susceptible and resis-

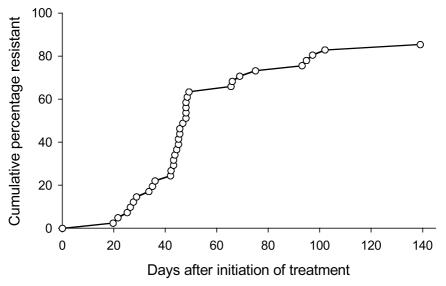


Figure 35. Emergence of streptomycin resistance during monotherapy in the British Medical Research Council trial.⁴⁶⁵

tant cultures varied to a considerable extent among individual patients, the point in time when resistance emerged cannot be known precisely. For this reason, the event was estimated to have occurred at the mid-point between the time the last susceptible and the first resistant culture was obtained. Resistance had already started to emerge after three weeks of treatment. By the time a patient had received two months of streptomycin monotherapy, the probability that drug resistance had been acquired exceeded 60%.

The explanation for this phenomenon is that among the many susceptible bacilli present in cavitary disease, spontaneous mutations occur at a given probability for each drug that convey resistance to that drug. The bacterial populations found in cavitary lesions obtained from resected lung tissue of patients were of the order of 10^7 to 10^9 bacilli, whereas those found in caseous foci did not exceed 10^2 to 10^4 bacilli.⁴⁶⁶ It has been experimentally demonstrated that it is selection of these mutants rather than adaptation to the medication.⁴⁶⁷ In a cavitary lesion containing 10^8 organisms, there will be around 10^2 isoniazid resistant mutants (i.e., one in a million) with the opportunity to replicate and become the dominant strain while the susceptible organisms are being killed off (figure 36).^{468,469}

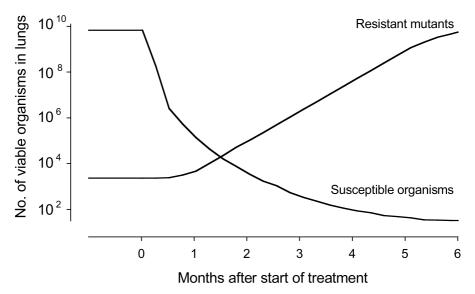


Figure 36. Diagrammatic presentation of the emergence of resistance to isoniazid during isoniazid monotherapy. Reproduced from⁴⁶⁹ by the permission of the publisher Churchill Livingstone and the author.

Monotherapy during sterilization of special populations

Not all drugs work equally well on all bacillary sub-populations. These sub-populations are exemplified in figure 37.⁴⁵⁶ None of the drugs works on the "dormant" or "latent"⁴⁷⁰ sub-population. Other specific sub-populations are the target of some drugs, such as pyrazinamide, which is active only in an acid environment. If, for instance, a patient with an initially isoniazid-resistant strain receives isoniazid, pyrazinamide, and ethambutol, the sub-population hypothesis would suggest that the patient's large bulk of rapidly metabolizing organisms is treated with ethambutol monotherapy. As there will be effective monotherapy in these special populations, resistant mutants should have a survival benefit.⁴⁶⁴

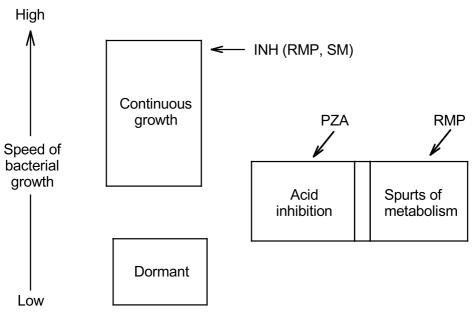


Figure 37. Special population hypothesis, indicating those bacterial populations at the start which are killed by the various drugs. Reproduced from⁴⁵⁶ by the permission of the publisher Churchill Livingstone.

Differences in bactericidal activity

Isoniazid has the highest early bactericidal activity of all of the anti-tuberculosis drugs. Thus, isoniazid-resistant mutants may have a selection advantage over a two-day period. This is not usually relevant, as this advantage is overcome over the ensuing period. However, should it happen that treatment is stopped after two days and subsequently resumed for another two-day period, the proportion of isoniazid-resistant mutants will have increased at the end of each cycle (figure 38).⁴⁶⁴

Sub-inhibitory concentrations

Whenever sub-inhibitory concentrations of a drug A are being taken, growth of bacilli susceptible to drug A will be mildly suppressed and their natural re-growth retarded if it is stopped. This does not apply to mutants resistant to drug A. They will not be affected at all by drug A but only by other drugs given simultaneously (figure 39).⁴⁶⁴ The mutants resistant to drug A will thus have a selective advantage. This might not be an uncommon scenario as the number of tablets required to be ingested (including

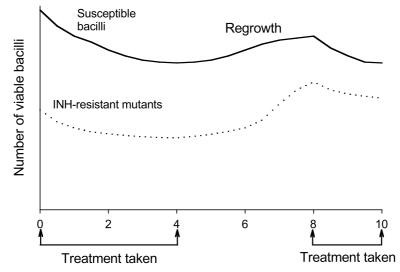


Figure 38. Bactericidal effects during two successive initial two-day phases of treatment with isoniazid and rifampicin. Reproduced from⁴⁶⁴ by the permission of the publisher International Union Against Tuberculosis and Lung Disease.

fixed-dose combination tablets) is large, and during self-administration patients might be tempted to take some but not all of the tablets. Mitchison⁴⁶⁴ points out that this mechanism would be most effective for

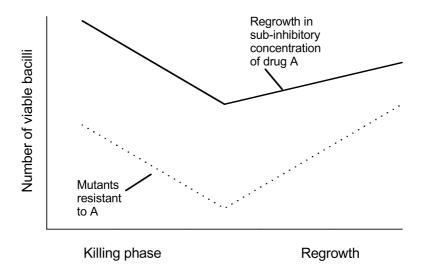


Figure 39. Sub-inhibitory concentrations of anti-tuberculosis drugs during regrowth. Reproduced from⁴⁶⁴ by the permission of the publisher International Union Against Tuberculosis and Lung Disease.

drugs with a high therapeutic margin such as isoniazid, as the effective half-life at sub-inhibitory concentrations would persist longer than that of other drugs.⁴⁷¹ The difference in pharmacokinetics of the drugs given together (in a combination tablet or in separate preparations) may be such that after several hours only one of the drugs is still active, leading to functional monotherapy. Sub-inhibitory concentrations of one or more drugs may be observed in patients with impaired gastrointestinal absorption.

Differences in post-antibiotic effect (lag phase)

When drugs are stopped, the length of time it takes bacilli to restart growth (post-antibiotic lag phase) differs for different anti-tuberculosis medications (figure 40).⁴⁷² Thus, for example, mutants resistant to drug A (with a long lag phase) are killed by drug B (with a short lag phase). Mutants resistant to drug A will thus start re-growth earlier when both drugs are stopped and obtain a selective advantage at the end of the cycle (figure 41).⁴⁶⁴

Clinical trials in the treatment of pulmonary tuberculosis

Since the discovery of streptomycin, clinical trials with anti-tuberculosis medications in various combinations have been carried out throughout the world to ascertain the shortest possible and best tolerated efficacious treatment regimens. The standard approach for studying a new drug or drug

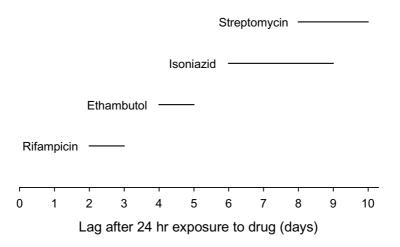


Figure 40. Post-antibiotic effects with *M. tuberculosis* lag periods before recommencement of growth after exposure in 7H10 medium.⁴⁷²

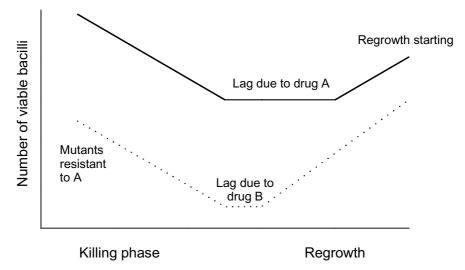


Figure 41. Bacteriopausal effects during regrowth of *M. tuberculosis*. Reproduced from⁴⁶⁴ by the permission of the publisher International Union Against Tuberculosis and Lung Disease.

combination is the randomized controlled clinical trial, whereby a group of patients is randomly assigned to the new regimen (experimental arm) or to the standard regimen (control arm). The element of randomization to reduce selection bias was actually first introduced in tuberculosis, with the first streptomycin trial of the British Medical Research Council.^{465,473,474}

Clinical trials have been conducted all over the world by different organizations and institutions. However, there can be little doubt that the leading role in the development of modern chemotherapy against tuberculosis was taken by the British Medical Research Council and its collaborators throughout the world,^{122,475} and by the United States Public Health Service and the United States Veterans Administration.⁴⁷⁶

While the efficacy of anti-tuberculosis treatment was fully appreciated, it is noteworthy that tuberculosis was for a long time not considered to be curable; temporary remission and prevention of emergence of resistance were the primary objectives for a long time. This is particularly surprising as it had been shown already in the 1950s that tuberculosis is curable using appropriate combination therapy.^{477,478}

In the following, only the highlights leading up to modern chemotherapy are summarized. For a more detailed account, the comprehensive review by Fox and collaborators from the British Medical Research Council¹²² or the individual trials conducted by the US Public Health Service^{319,479-491} might be consulted.

Streptomycin monotherapy

Shortly after the discovery of streptomycin, clinical trials with streptomycin monotherapy were conducted in Great Britain⁴⁶⁵ and the United States.⁴⁹² It was noted in these trials that case fatality from tuberculosis was considerably reduced. However, it was also seen that patients improved over the first few months and subsequently deteriorated, in many cases due to acquisition of streptomycin resistance. Among survivors, sputum conversion did not much differ between those receiving streptomycin and those not (figure 42).⁴⁹² The insoluble problem was the selection of resistant strains. While toxicity could be reduced by lowering the dosage and spacing administration more widely, the problem of bacterial resistance was not resolved.⁴⁹³ The streptomycin trials impacted considerably on research for the next 20 years, which largely concentrated on methods of preventing the emergence of drug resistance.

Streptomycin plus para-aminosalicylic acid

The introduction of para-aminosalicylic acid into the armamentarium allowed combination therapy to be used. In a study of the British Medical Research

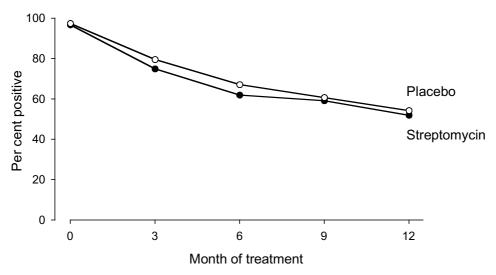


Figure 42. Sputum culture conversion among patients receiving streptomycin compared to placebo.⁴⁹²

Council, streptomycin monotherapy, para-aminosalicylic acid monotherapy and chemotherapy with both drugs combined was carried out.^{494,495} It was demonstrated unequivocally, and for the first time, that combined chemotherapy reduced the risk of acquisition of resistance. Similarly, a clinical trial in Denver, Colorado, USA showed that the combination of streptomycin and para-aminosalicylic acid overcame the emergence of resistance, in contrast to monotherapy with either one (figure 43).⁴⁹³

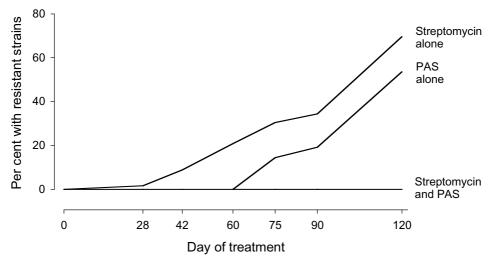


Figure 43. Emergence of resistance to streptomycin and/or para-aminosalicylic acid given alone or in combination. Reproduced from⁴⁹³ by the permission of the publisher American Thoracic Society at the American Lung Association.

Streptomycin plus para-aminosalicylic acid plus isoniazid

It appears that the logical step following the introduction of isoniazid, namely to compare the efficacy of streptomycin, para-aminosalicylic acid, and isoniazid with a control arm of streptomycin plus para-aminosalicylic acid, was never subjected to a formal randomized clinical trial, either by the US Public Health Service or by the British Medical Research Council. This is particularly astonishing, as this triple combination therapy must be considered the breakthrough in tuberculosis treatment because it introduced the certainty of consistently curing tuberculosis patients with an initially fully susceptible strain.

It is furthermore a curiosity in the history of medicine that the curative results of this combination therapy were not even accorded a full-length article. The experiences in Edinburgh of Sir John Crofton and collaborators were relegated to the correspondence section of the *American Review of Tuberculosis* (figure 44).⁴⁷⁷ The efficacy of this approach seemed convincing, although a randomized trial would surely have been indicated to remove any lingering doubts about biased selection and ascertainment. A subsequent study of the British Medical Research Council, begun in 1956, added a streptomycin supplement until susceptibility to PAS was demon-

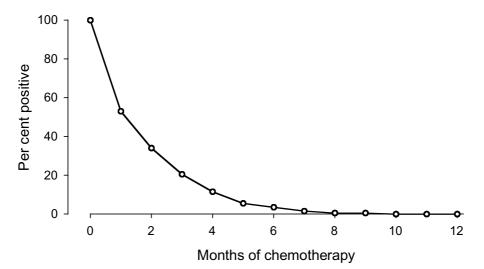


Figure 44. Sputum culture conversion in patients with pulmonary tuberculosis due to susceptible organsisms, with triple therapy consisting of streptomycin, paraaminosalicylic acid, and isoniazid. Reproduced from⁴⁷⁷ by the permission of the publisher American Thoracic Society at the American Lung Association.

strated.⁴⁹⁶ This indicates that the importance of a resistance-preventing component in the intensive phase was not yet fully appreciated. In the report on US Public Health Service trial 4, it was explicitly stated that there was no advantage of using all three drugs in cases of recent origin.⁴⁸² In US Public Health Service trial 3, a comparison of the combination streptomycin plus isoniazid with streptomycin plus PAS was made; it demonstrated the superior ability of the isoniazid-containing regimen to induce culture conversion (figure 45).⁴⁸¹ However, the difference between a regimen of streptomycin plus para-aminosalicylic acid plus isoniazid versus streptomycin plus para-aminosalicylic acid was not ascertained.

Nevertheless, common sense prevailed and by the end of the 1950s, the regimen that had been used in Edinburgh became, at least in the United

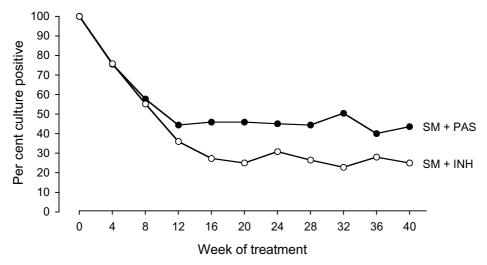


Figure 45. Sputum culture conversion among patients treated with streptomycin and para-aminosalicylic acid compared to streptomycin and isoniazid.⁴⁸¹

Kingdom, standard practice ⁴⁹⁷ following a trial by the British Medical Research Council demonstrating faster conversion, fewer bacteriologic failures and relapses. ^{496,498} The WHO considered it one of the major regimens for low-income countries. ⁴⁹⁹ It took many years for experts of other countries to be convinced of its importance, and that only after an international comparative clinical trial. ⁵⁰⁰

Isoniazid plus ethambutol

Because of the frequency of side effects associated with para-aminosalicylic acid, ethambutol appeared an attractive alternative. The US Public Health Service trial 16 showed that sputum conversion was indistinguishable in patients receiving, in addition to isoniazid, ethambutol in lieu of para-aminosalicylic acid (figure 46), although there was a marked reduction in the occurrence of adverse drug events.⁴⁸⁴

Neither the US Public Health Service nor the British Medical Research Council studied a 12-month regimen with isoniazid and ethambutol throughout, supplemented by streptomycin in the intensive phase, although this regimen has been widely used in low-income countries where thioacetazone has been abandoned.

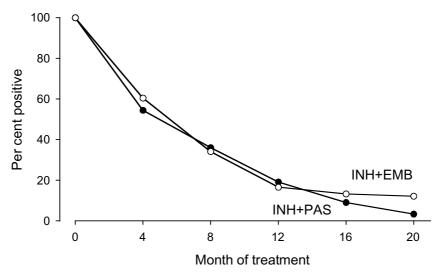


Figure 46. Sputum conversion among patients receiving isoniazid and para-amino-salicylic acid compared to isoniazid and ethambutol.⁴⁸⁴

Isoniazid plus thioacetazone

In East Africa, a comparison of 12-month regimens was carried out with isoniazid plus thioacetazone throughout, one arm containing streptomycin during the first two months and another arm without streptomycin.⁵⁰¹ It was demonstrated that the two-month supplement of streptomycin contributed to a higher cumulative conversion rate (figure 47).

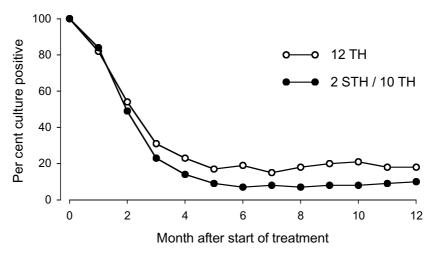


Figure 47. Effect on culture conversion of adding two months of streptomycin to a 12-month regimen of isoniazid and thioacetazone.⁵⁰¹

Thioacetazone replaced para-aminosalicylic acid very rapidly throughout English-speaking sub-Saharan Africa because of the better tolerance and important cost savings.

Isoniazid plus rifampicin

With the introduction of rifampicin, a more rapid conversion was demonstrated when it replaced streptomycin (figure 48).⁴⁸⁷ This was, however, not the main progress made with rifampicin-containing chemotherapy. In a trial in France, rifampicin-containing regimens were tested in three different dura-

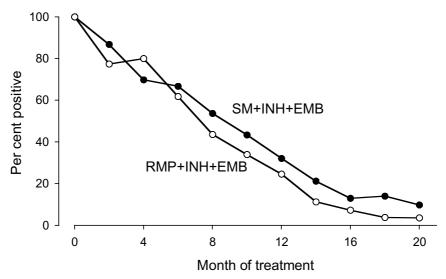


Figure 48. Effect of replacing streptomycin by rifampicin on culture conversion.487

tions of chemotherapy: six, nine, and 12 months.^{502,503} It was demonstrated that nine months of isoniazid plus rifampicin, supplemented by either ethambutol or streptomycin during the first three months, was the optimum duration,⁵⁰² and the relapse rate during a remarkable mean follow-up time of 101 months with this regimen was only two out of 85 patients.⁵⁰³

The use of rifampicin provided curative treatment of less than one year's duration, and the term "short-course chemotherapy" became the brand name of this new successful strategy.⁵⁰⁴

Isoniazid plus rifampicin plus pyrazinamide (plus a fourth drug)

Despite its remarkable efficacy in experimental models, ^{505,506} pyrazinamide was not retained in routine chemotherapy because of its hepatotoxicity.

Based on evidence that the addition of pyrazinamide hastened sputum conversion, a series of studies was designed by the British Medical Research Council. In 1970, it was demonstrated for the first time that the inclusion of rifampicin and pyrazinamide in a regimen of isoniazid and streptomycin substantially reduced the subsequent risk of relapse.¹²²

A multitude of clinical trials was designed and carried out by the British Medical Research Council with regimens containing, as a minimum, isoniazid, rifampicin, and pyrazinamide in the intensive phase, virtually always supplemented by streptomycin during this period.¹²² Two studies in East Africa were critical for future research into this combination.^{507,508} In these trials it was observed that regimens containing pyrazinamide but not rifampicin were almost as effective as those containing rifampicin. Furthermore, there was later evidence that both drugs given in the regimen were more effective than one alone.¹²² These studies laid the basis for modern treatment.

The consistent finding in these studies was that the four drugs were optimally given for a two-month intensive phase, followed either by four months of rifampicin plus isoniazid or six months of a combination of drugs not containing rifampicin (the continuation phase).

The role of the fourth drug (streptomycin or ethambutol) is unclear as few studies have evaluated it, ^{491,509} but most likely it has a minor role in patients with a strain that is fully susceptible at the outset.¹²² A recommendation to drop the fourth drug in patients with sputum smear-negative tuberculosis seems to have no evidence base. Patients with paucibacillary disease may require a shorter duration of treatment (see below); however, dropping the fourth drug in the intensive phase may not be justified as it may lead to functional monotherapy with rifampicin in lesions with a low pH among patients with a strain that is initially resistant to isoniazid (pyrazinamide not being active in such lesions).

Rifampicin-containing continuation phase

A regimen consisting of a two-month intensive phase with isoniazid, rifampicin, pyrazinamide, and streptomycin, followed by a four-month continuation phase with isoniazid plus rifampicin, all given daily, was first evaluated in Singapore.⁵¹⁰⁻⁵¹² The high efficacy of this regimen was confirmed in the United Kingdom, and was equally effective if streptomycin was replaced by ethambutol.⁵¹³⁻⁵¹⁵ It has become the standard regimen for patients with fully susceptible organisms in most industrialized countries. Shorter durations have been put on trial, ^{516,517} but the frequency of relapse makes it impossible to reduce the minimum duration of six months. In the United States, US Public Health Service trial 21 evaluated the same regimen, but without the supplement of ethambutol or streptomycin (except for those with a high probability of initial resistance) in the intensive phase and showed it to be efficacious.^{490,491} However, given the possibility of drug resistance among new cases of tuberculosis in many locations, the recommendation for a four-drug initial treatment is preferred, at least in areas where drug resistance is frequent or unknown. In the United Kingdom, a four-drug intensive phase is always recommended.⁵¹⁸ The IUATLD and WHO also recommend a four-drug intensive phase in new sputum smearpositive cases of pulmonary tuberculosis and other severe cases of tuberculosis where this regimen is being used.^{8,13}

Non-rifampicin-containing continuation phase

Current options for a non-rifampicin-containing continuation phase are isoniazid plus thioacetazone or isoniazid plus ethambutol.

A four-drug, two-month intensive phase followed by six months of isoniazid plus thioacetazone has been found to be highly efficacious in East Africa. 519,520

No critical evaluation of an ethambutol-containing continuation phase has been carried out extensively. One trial in India evaluated the effectiveness of a fully unsupervised eight-month regimen with isoniazid and ethambutol throughout, supplemented by rifampicin and pyrazinamide in the two-month intensive phase.^{521,522} The entire treatment was self-administered and compared to a six-month regimen using rifampicin throughout, given twice-weekly, and at least partially supervised. The results during chemotherapy were encouraging with the eight-month regimen. Four per cent had an unfavorable response during chemotherapy and five per cent relapsed; the relapse rate was only half that in the directly observed control arms.

Intermittent regimens

To facilitate directly observed therapy, various intermittent regimens have been studied extensively.^{122,521} In Chennai (formerly Madras), India, all parameters were superior in patients receiving twice-weekly isoniazid plus para-aminosalicylic acid, supplemented by streptomycin during the intensive phase, as compared with patients receiving once-weekly isoniazid plus para-aminosalicylic acid for self-administered treatment.⁵²³ This study represented a major advance in research aiming at improving adherence with intermittent regimens.⁴⁹⁸

The majority of the trials have evaluated six-month regimens, with rifampicin throughout, each dose given under direct observation. In Denver, Colorado, USA, for instance, a regimen with daily treatment given for just the first two weeks, followed by twice weekly adminstration for the remainder of the course, was highly successful, ⁵²⁴ and similar studies have been conducted in Poland. ^{509,525}

Intermittent regimens have been shown to be as (or almost as) efficacious as daily regimens, and greatly facilitate direct observation of drugintake. A potential problem with intermittent regimens is that errors resulting from missing one dose may have greater impact than missing a single dose in a daily regimen. This might be further compounded if the fourth drug in the intensive phase is omitted. In a controlled clinical trial in India with a twice-weekly regimen, bacteriologic sputum conversion was inferior if ethambutol was omitted (figure 49).⁵²¹ Twice-weekly regimens might also be inferior even if all drugs are being taken in populations where a large proportion of patients acetylates isoniazid rapidly, as such patients generally have inferior results in widely spaced drug administration.¹²² Thus, while regimens for both twice-weekly and thrice-weekly application have been studied, the only intermittent regimens WHO recommends are thrice-weekly regimens.¹³

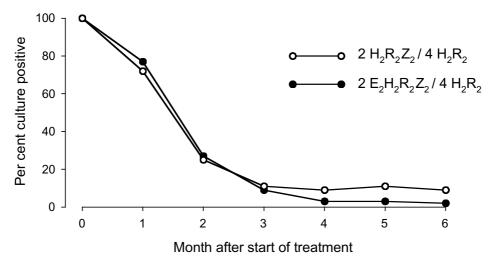


Figure 49. Effect of adding a fourth drug (ethambutol) during the first two months to a rifampicin throughout regimen on culture conversion.⁵²¹

Not all drugs are equally suitable for intermittent use. Thioacetazone, for example, is not suitable for intermittent use.⁵²⁶ Furthermore, intermittent treatment is indicated only to facilitate directly observed therapy, not for self-administered treatment. Thus, unless a rifampicin-containing continuation phase is selected, the principal issue is the efficacy of the use of intermittent therapy during the intensive phase of treatment.

Remarkably little is known about the efficacy of intermittent use during an intensive phase containing four medications, followed by a selfadministered continuation phase that does not contain rifampicin. Concerns have been raised that an eight-month regimen with an intermittent intensive phase from the outset may be inferior in HIV-infected patients.⁵²⁷ To facilitate directly observed therapy in national programs, these are critical issues that need urgent attention.

Treatment regimens of less than six months' duration

Regimens of four months' duration (containing rifampicin throughout) for bacteriologically confirmed pulmonary tuberculosis have been studied in Singapore, but yielded unacceptably high relapse rates.⁵¹⁰⁻⁵¹²

A regimen of four and a half months duration for bacteriologically (sputum smear and culture) confirmed pulmonary tuberculosis has been studied in Agra, India.^{516,517} In this trial, four drugs (isoniazid, rifampicin, pyrazinamide, and streptomycin) were given for a total of three months, followed by one and a half month of isoniazid plus rifampicin, all given daily. All but one of the 65 patients enrolled were eligible for and followed up for relapse, and only one patient relapsed during the two-year follow-up period.⁵¹⁷ Despite the seeming efficacy of this four and a half-month regimen, confirmatory studies have not become available, the regimen has never been accepted by the medical community, and the credibility of the result of the study was actually challenged.⁵²⁸

Among patients with repeatedly negative sputum smears, shorter regimens have been investigated.¹²² If the initial culture was negative (but radiologically the disease was considered to be active) or positive, relapse rates were three per cent or less with a three-month or a four-month regimen, respectively.⁵²⁹ However, in routine practice even countries involved in these trials have abandoned the practice of treating patients with newly diagnosed (but bacteriologically unconfirmed) pulmonary tuberculosis for less than six months.

Currently, regimens shorter than six months duration are not recommended by WHO for bacteriologically unconfirmed tuberculosis. A principal consideration is the prevailing uncertainty about the quality of bacteriologic examinations (sputum smear microscopy) in many national tuberculosis programs.

Clinical trials in extrapulmonary tuberculosis

Two forms of extrapulmonary tuberculosis have been studied in welldesigned clinical trials: tuberculosis of peripheral lymph nodes and tuberculosis of the spine. The treatment of tuberculosis of the central nervous system has been subject to numerous investigations, but because of the small cases in each study, the certainty about the optimum treatment is limited.

Tuberculosis of peripheral lymph nodes

In many populations, tuberculosis of peripheral lymph nodes (particularly cervical and axillary) is the most frequent extrapulmonary manifestation of tuberculosis.⁵³⁰ While in the past, in milk-consuming cultures, tuberculosis of peripheral lymph nodes may frequently have been caused by *M. bovis*, particularly in children, it is now almost universally caused by *M. tuberculosis*, ⁵³¹ it is found in all age groups, but with a predilection for the young ^{532,533} and for females.⁵³⁰

It appears that treatment of lymphatic tuberculosis was long considered to be a surgical domain, due to a misunderstanding of it as a localized disease process. This concept was demonstrated to be erroneous in a retrospective study conducted among cases diagnosed between 1965 and 1973 in the United Kingdom.⁵³⁴

A prospective study of the treatment of tuberculosis of peripheral lymph nodes was carried out in the United Kingdom, comparing two 18-month regimens, one with isoniazid plus ethambutol, the other with isoniazid plus rifampicin throughout, and both supplemented by streptomycin during the first two months.⁴⁷⁶ No difference in treatment results between the two groups was found.

In a second prospective study, conducted by the British Thoracic Society, an 18-month regimen was compared with a nine-month regimen.^{535,536} Both regimens were based on isoniazid plus rifampicin throughout and supplemented by ethambutol during a two-month intensive phase. No difference in treatment outcome was identified between the two regimens.

In a third prospective study in the United Kingdom, various regimens using isoniazid plus rifampicin throughout were compared.⁵³⁷ The control regimen was the same as the nine-month regimen in the British Thoracic Society. One of the experimental regimens had the same duration, but ethambutol was replaced by pyrazinamide. The second experimental arm received the same regimen as the first, but for only six months. No differences among the three regimens were found.

A review of published materials concludes that a six-month regimen similar to that used in pulmonary tuberculosis is also adequate for treatment of tuberculosis of peripheral lymph nodes.⁵³⁸

It was noted in the British trials that tuberculosis of peripheral lymph nodes does not always appear to respond clinically to treatment, and treatment may be declared a failure on clinical grounds. Cultures from nodes that were newly developing during treatment or from abscesses from newly draining nodes subsequent to treatment completion remained bacteriologically sterile. It has been postulated that the phenomenon is caused by an immunologic response to tuberculo-protein.⁵³⁵

Tuberculosis of the spine

Tuberculosis of the spine is one of the most important extrapulmonary forms of tuberculosis both in terms of relative frequency and the substantial potential of permanent disability. It has been estimated that more than half of all osteoarticular manifestations of tuberculosis in India affect the spine.⁵³⁹

Before the advent of anti-tuberculosis chemotherapy, treatment consisted of bed-rest, improvement of the patient's nutritional status, and, in some cases, posterior spinal fusion. 540

In the 1950s and early 1960s, two extreme positions marked the divergence of opinions about the appropriate approach to the treatment of tuberculosis of the spine. In Nigeria, successful treatment with chemotherapy alone was reported. ^{541,542} In Hong Kong, excellent results were reported with anterior spinal fusion. ⁵⁴³⁻⁵⁴⁶

It was against this background that the British Medical Research Council planned ^{539,547,548} and conducted a series of controlled clinical trials, resulting in 14 scientific reports. ⁵⁴⁹⁻⁵⁶²

The trials were conducted in Hong Kong, India, Korea, and Zimbabwe. All trials evaluated the role of chemotherapy and various operative and nonoperative surgical procedures. Chemotherapy lasted from six to 18 months at various points in time. The most recent trial established that a regimen of six months' duration with isoniazid plus rifampicin throughout was as effective as any other regimen.⁵⁶² It was concluded that outpatient chemotherapy with standard short-course chemotherapy based on isoniazid, rifampicin, and pyrazinamide should be the main management of uncomplicated spinal tuberculosis.⁵⁶¹

It is likely, based on the results of these studies, that a regimen that is effective for pulmonary tuberculosis should be equally effective for the treatment of tuberculosis of the spine.

Tuberculosis of the central nervous system

Tuberculous meningitis is the most important central nervous system manifestation of tuberculosis. The optimum treatment of tuberculous meningitis is not known, and recommendations are based on the pharmacokinetic properties of the medications and, to a large extent, on inference and common sense.

The blood-brain barrier poses particular problems for the choice of the right drug combinations as penetration into the cerebrospinal fluid and its ratio to serum concentrations varies widely among the various anti-tuber-culosis drugs.

The key issue is the extent of plasma binding of the drug, as probably only the unbound portion penetrates into the central nervous system, thus explaining the differences between isoniazid and pyrazinamide on one hand, and rifampicin on the other.

Isoniazid is recognized as a drug with excellent penetration into cerebrospinal fluid. $^{\rm 61,563}$

Rifampicin, in contrast to isoniazid, has very poor penetration into cerebrospinal fluid, ⁵⁶³ but seems to appear in higher concentrations at the beginning of treatment, in the phase where the meninges are inflamed. ^{564,565} However, because tuberculosis is not a localized disease, the use of rifampicin is beneficial for the treatment of lesions other than those in the central nervous system that may be simultaneously present.

Pyrazinamide has excellent penetration into cerebrospinal fluid. 563

Ethambutol penetrates poorly into normal or uninflamed meninges, but penetrates fairly well into inflamed meninges. 565-568

Streptomycin penetrates relatively poorly into cerebrospinal fluid. 563

Among the thioamides, ethionamide has been found to have high penetration into cerebrospinal fluid. ^{565,568-570} Based on these pharmacokinetic properties and other considerations, it has been recommended that treatment for suspected or confirmed tuberculous meningitis should begin with a two-month intensive phase incorporating isoniazid, rifampicin, and pyrazinamide plus streptomycin. ⁵⁷¹ The optimum duration of the continuation phase is not known, but based on limited information ⁵⁷² a continuation phase associating isoniazid and rifampicin for a duration of at least seven months has been advocated. ⁵⁶³ This regimen may pose problems in patients with an isoniazid-resistant strain because of the unpredictable concentrations of rifampicin. Where available, ethion-amide might provide a less well tolerated alternative in such a case. ^{570,573}

Influence of HIV infection on the choice of a regimen

Among tuberculosis patients with HIV infection, two major issues need to be addressed.

The first concerns the initial observations made by clinicians when treating HIV-infected patients with anti-tuberculosis drugs: tolerance of the medications was poorer than in patients without HIV infection.

A second issue concerns the efficacy of the regimens usually prescribed. Patients with HIV infection may suffer from diarrhea, which may, through its lowering of drug serum concentrations, adversely compromise the efficacy of the regimen, favoring the emergence of resistance and subsequent relapse.

Adverse drug events

Adverse drug events occur much more frequently among HIV-infected tuberculosis patients. In particular, cutaneous hypersensitivity reactions are frequent. These have mostly been attributable to thioacetazone, ^{443,445,447,574-577} and to a lesser extent to streptomycin, ⁵⁷⁵ rifampicin, ^{215,216,578} and isoniazid. ⁵⁷⁸

The frequent and sometimes fatal cutaneous adverse drug events among HIV-infected tuberculosis patients due to thioacetazone preclude its use in patients known to be HIV-infected.^{8,13} It is best replaced with ethambutol.

An increased frequency of non-cutaneous adverse drug events (hepatotoxicity, gastrointestinal disturbances, thrombocytopenia) to isoniazid ^{578, 579} and rifampicin has been reported. ⁵⁷⁸⁻⁵⁸⁰

Anti-retroviral therapy poses particular problems because of interactions with rifampicin that preclude simultaneous use of the two regimens.

Treatment efficacy

As enteropathy is a frequent occurrence in HIV-infected patients, anti-tuberculosis medications might be less well absorbed,⁵⁸¹ thus leading to treatment failure, relapse⁵⁸² or acquisition of drug resistance.⁵⁸³ Pharmacokinetic studies among patients with AIDS in various centers in Puerto Rico and the USA have demonstrated that serum peak concentrations, particularly of rifampicin and ethambutol, were frequently lower than expected.³³⁴ However, malabsorption of anti-tuberculosis medications does not seem to be a major issue in most HIV-infected patients.^{584,585}

Sputum conversion is rapid, and even faster among HIV-positive than HIV-negative patients (figure 50).⁵⁸⁶ However, concern has been expressed

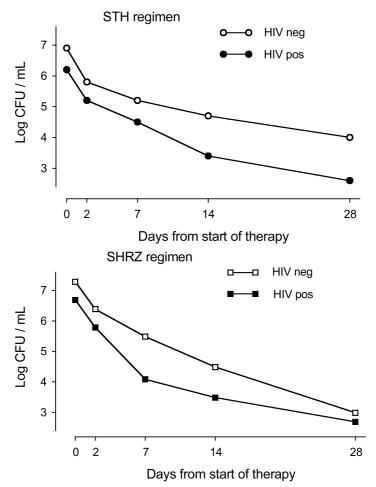


Figure 50. Bacteriologic response to chemotherapy among HIV-negative and -positive patients, by treatment regimen.⁵⁸⁶

that the sputum bacillary load may not reflect the underlying number of bacilli a patient harbors, and thus there might be a need for prolonged treatment.⁵⁸⁷

Regimens of six to nine months duration containing rifampicin throughout have been highly efficacious in terms of both low frequency of bacteriologic failure ⁵⁷⁶ and relapse. ^{579,588,589} Eight-month regimens give acceptable results in the field. ⁵⁹⁰ In contrast, 12-month regimens that do not incorporate any rifampicin have shown a high frequency of failures ^{591,592} and relapse. ^{591,593,594}

If antiretroviral therapy is given simultaneously with treatment for tuberculosis, paradoxical responses have been reported with worsening of the clinical presentation, assumed to be an immunologic response.⁵⁹⁵ Antiretroviral drugs such as protease inhibitors (saquinavir, indinavir, ritonavir, and nelfinadvir) and non-nucleoside reverse transcriptase inhibitors (nevirapine, delaviridine, and efavirenz) have substantive interactions with rifamycins.⁵⁹⁶ Rifampicin will reduce the blood concentrations of protease inhibitors. The efficacy of the latter will thus be reduced when concommitantly administered with rifampicin. The interaction with nucleoside reverse transcriptase inhibitors (zidovudine, didanosine, zalcitabine, stavudine, and lamivudine) is probably not clinically relevant.⁵⁹⁶

The US Public Health Service conducted study 22, comparing the efficacy of once-weekly isoniazid plus rifapentine with twice-weekly isoniazid plus rifampicin in a four-month continuation phase following a four-drug, two-month intensive phase.⁵⁹⁷ Among 61 patients with concomitant HIV infection, none experienced treatment failure. However, three of the 31 patients on the rifampicin-containing continuation phase relapsed, all with fully susceptible organisms, but five of the 30 patients on the rifapentine regimen relapsed, four of whom had acquired rifamycin resistance. Obviously, isoniazid as a companion drug in once-weekly treatment is inadequate, and patients effectively received rifapentine monotherapy. There is indeed cause for concern that, by analogy, HIV-infected patients with initial isoniazid resistance may acquire unnoticed (no apparent failure during treatment) rifampicin resistance if treated with this drug in the continuation phase.⁵⁹⁸ Nevertheless, the reasons for acquisition of rifamycin resistance in this study have not yet been fully elucidated, and there are indications that it is attributable to an inadequate dosage of rifapentine.

Relapse following cessation of chemotherapy appears to be more frequent among HIV-infected compared to HIV-non-infected individuals, ^{594,599} and post-treatment preventive chemotherapy with isoniazid appears to reduce that risk. ⁵⁹⁹

Influence of isoniazid resistance on the choice of a regimen

Isoniazid is a key drug in the treatment of tuberculosis and its inclusion in every first-line regimen is the standard of care. Pre-existing initial resistance to isoniazid might be conducive to the development of additional resistance, particularly if treatment organization is poor, as the data from the WHO/IUATLD global surveillance project on drug resistance seem to suggest (figure 51).⁶⁰⁰

Patients with initial isoniazid resistance who are given a four-drug intensive phase for two months, followed by isoniazid and thioacetazone in the continuation phase, fail more frequently than patients with fully susceptible organisms.^{519,601} Such patients can be re-treated effectively with a regimen containing rifampicin plus ethambutol throughout, supplemented by pyrazinamide during the first three months, and additionally by streptomycin during the first two months.^{8,13,602-604}

It is not very well known how effective such a re-treatment regimen is if there is additional ethambutol resistance. The extent to which such functional rifampicin monotherapy in the continuation phase of the re-treat-

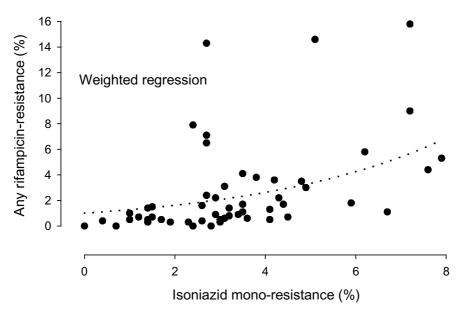


Figure 51. Correlation between isoniazid mono-resistance and any rifampicin resistance among never treated patients. Ecological analysis from the Global Project on Surveillance of Anti-tuberculosis Drug Resistance.⁶⁰⁰

ment regimen is efficacious and not causing drug resistance in HIV-infected patients remains to be seen. ⁵⁹⁸

Influence of isoniazid plus rifampicin resistance on the choice of a regimen

Patients with multidrug-resistant tuberculosis (bacilli resistant to at least isoniazid and rifampicin) are only rarely expected to be cured solely using the six essential drugs. Under program conditions treatment outcome with the standard WHO recommended re-treatment regimen is poor if there is multidrug resistance.⁶⁰⁵ Barring spontaneous remission, such patients are incurable and frequently become chronic excretors of bacilli in countries where only the essential drugs are available for use.

Drugs other than the six essential drugs are of lower efficacy, much more costly, and in the majority of cases, much less well tolerated.⁶⁰⁶⁻⁶⁰⁸ It is also not yet known which treatment strategy is best. Proposals for treating multidrug-resistant tuberculosis include the utilization of a standard regimen or an individualized approach based on susceptibility testing.⁶⁰⁹ There have been no randomized controlled clinical trials to evaluate these regimens and insufficient experience has been accumulated to make firm recommendations at this point in time.

Strategic considerations, indications, and recommendations for the choice of treatment regimens in a national tuberculosis control program

The number of possible errors can be minimized by the systematic, country-wide use of standard regimens. Recommended standard treatment regimens are based on clinical efficacy trials in terms of dosage, mode of administration, and duration of treatment. Deviations from standard treatment regimens are indicated only in the case of adverse drug events, for patients presenting with pre-existing medical conditions that require a modification of the regimen, or in the presence of suspected or confirmed resistance to one or more drugs.

Both WHO and the IUATLD recommend standard treatment regimens which vary according to the category of the patient.¹³ The three categories are:⁸

- Patients with sputum smear-positive tuberculosis or severe extrapulmonary tuberculosis never treated before for as much as one month;
- Patients with other forms of tuberculosis (sputum smear-negative and extrapulmonary) never treated before for as much as one month;
- Patients with sputum smear-positive tuberculosis treated previously for one month or more (return after treatment failure, return after default, and relapse).

No specific recommendations have been made on how to deal with patients with continued bacteriologically active disease following a full re-treatment course (chronic excretors).

A primary objective of any tuberculosis control program must be to limit to the largest possible extent the emergence of organisms resistant to the available medications. This is a guiding principle for any chemotherapy, but it is particularly crucial in tuberculosis control, because the armamentarium of drugs is limited and the prospects in the near future for new, affordable drugs with an efficacy comparable to that of isoniazid, rifampicin or pyrazinamide are slim for most low-income countries.

Choice of first-line regimen

First-line regimens of six to eight months duration are the most efficacious available. All are based on a four-drug initial intensive phase. Whether a four-month (with rifampicin) or a six-month continuation phase (without rifampicin) is selected depends on the availability of resources for drugs and personnel, and considerations about the fall-back (re-treatment) regimen, particularly in the case of treatment failure. Twelve-month regimens (without rifampicin) have been widely used for bacteriologically unconfirmed disease, but their efficacy in HIV-infected patients appears to be inferior to the shorter, but more intensive alternatives.

The continuation phase in the eight-month regimen consists of six months of isoniazid plus thioacetazone. A frequently chosen alternative to thioacetazone is ethambutol. This change potentially weakens the re-treatment regimen (functional rifampicin monotherapy in the continuation phase). This increases the risk of development of multidrug resistance. The IUATLD therefore recommends the addition of pyrazinamide throughout the re-treatment regimen⁸ when ethambutol has been used in the continuation phase of initial treatment.

Many countries have moved towards a first-line regimen which contains rifampicin throughout. Patients truly failing on such a regimen have a high probability of initial multidrug resistance (or initial isoniazid resistance and acquired rifampicin resistance). The re-treatment regimen recommended by the IUATLD and WHO is highly unlikely to cure such a patient, and additionally carries the risk of acquisition of ethambutol resistance. It is not clear whether re-treatment incorporating both ethambutol and pyrazinamide in the continuation phase will overcome this problem. Given the relative weakness of these two drugs, there is a risk of losing both. This has been termed the "amplifier effect" (a new term for an old phenomenon, successive acquisition of additional drug resistance) and has been observed to occur in an outbreak in urban Peru.^{610,611} It has not been observed in other settings where a non-rifampicin-containing continuation phase is routine in the first-line regimen.⁶¹²

8-month regimens

The eight-month regimen evaluated in East Africa (a directly observed fourdrug, two-month intensive phase followed by six months of self-administered isoniazid plus thioacetazone) has become the principal treatment regimen for previously untreated smear-positive pulmonary tuberculosis in IUATLD collaborative programs.^{8,604} Programs basing their chemotherapy on this regimen are using a highly cost-effective intervention.⁶¹³

Replacement of streptomycin by ethambutol in the intensive phase did not adversely affect adherence to directly observed therapy in a study conducted in large urban settings in Tanzania, ⁶¹⁴ and gave similar treatment outcome under routine conditions in Madagascar, although the proportion of failures was somewhat higher than in the streptomycin group. ⁶¹⁵ It also yielded good results in Benin. ⁶¹⁶

It is likely that replacement of thioacetazone by ethambutol is equally effective, as demonstrated in a clinical trial in India in a patient population with a low prevalence of HIV infection.^{521,522} When thioacetazone cannot be used because of a high prevalence of HIV infection, its replacement by ethambutol is therefore often recommended.⁸

6-month regimens

The shortest treatment regimen of proven efficacy for bacteriologically confirmed tuberculosis consists of six months of isoniazid plus rifampicin, supplemented by pyrazinamide plus either streptomycin or ethambutol during the first two months. This has been convincingly demonstrated where all medications were taken daily throughout the course of treatment.¹²² In Poland, a study with this regimen, with the continuation phase given twice weekly, led to neither failures nor relapses.⁵²⁵ Similar good results were obtained with the same regimen in Singapore, with the continuation given thrice weekly.⁶¹⁷

Most industrialized countries have adopted this regimen, given daily in the intensive phase and daily or intermittent in the continuation phase, as their regimen of choice for patients without a history of prior treatment.

12-month regimens

The best documented 12-month regimen currently used in low-income countries consists of 12 months of isoniazid plus thioacetazone, supplemented by streptomycin during the first two months.¹²² This regimen has been widely used in IUATLD collaborative programs in patients without a prior history of treatment. Amongst these, it is given for cases with positive sputum smears who cannot receive a directly observed rifampicin-containing intensive phase and for the majority of patients whose sputum smears are negative or who have extrapulmonary tuberculosis which is not lifethreatening.

In Uganda, the frequency of adverse drug events and survival as the main outcomes of interest were compared for the above 12-month regimen and a nine-month, rifampicin-throughout regimen (supplemented by pyrazinamide during the first two months) among HIV-infected patients.⁶¹⁸ As expected, adverse drug events were much more common in the former than the latter regimen, but survival over a two-year follow-up period was identical.

In Malawi, HIV-infected patients with sputum smear-negative tuberculosis who were treated with a 12-month regimen (12 months of isoniazid plus thioacetazone or ethambutol, supplemented with streptomycin during the first month), had a very high relapse rate approaching 20% (compared to seven per cent among HIV-negative patients).⁵⁹⁰ These findings critically challenge the continued use of such a regimen in countries where the prevalence of HIV infection among tuberculosis patients is high.

Choice of re-treatment regimen

Treatment regimens for a national tuberculosis control program should be designed to allow curative treatment of patients requiring a re-treatment regimen, because it is the patient's last chance to get cured. The need for a re-treatment regimen is based on the increased probability of resistance to the medications used in patients who have received prior treatment. That this is the case has been amply demonstrated.⁶¹⁹ An efficacious re-treatment regimen must encompass at all times, throughout treatment, at least two drugs to which the organism is still likely to be susceptible. Countries which do not have access to medications other than the six essential drugs for patients who might require them must choose a re-treatment regimen based on these six drugs.

Because isoniazid is always given in the first-line regimen, a patient failing to respond to the treatment regimen will have a high probability of already having isoniazid resistance at the outset of treatment. To adhere to the principle of a re-treatment regimen incorporating at least two efficacious drugs, neither rifampicin nor ethambutol should have been used as the sole companion drug with isoniazid at the point of failure (defined as sputum smear-positive at five months or later), if either of these drugs is to be effective in a re-treatment regimen. Their use as a sole companion drug with isoniazid constitutes functional monotherapy in such a patient and presents a risk that resistance will have developed to the companion drug (in this case, either rifampicin or ethambutol). This has been the rationale behind the recommendation of the IUATLD to utilize isoniazid plus thioacetazone in the continuation phase. Should bacilli resistant to thioacetazone emerge, re-treatment is still likely to be successful.

This re-treatment regimen proposed by WHO and the IUATLD consists of eight months of isoniazid, rifampicin, and ethambutol, supplemented by pyrazinamide during the first three, and streptomycin during the first two months. This regimen uses the full range of available drugs except thioacetazone. Such a regimen has a high probability of curing any patient who does not commence treatment with organisms already resistant to both isoniazid and rifampicin. Patients with multidrug-resistant strains have, after taking the re-treatment regimen, an outcome that is not appreciably better than reported outcomes in the pre-chemotherapy era.⁶²⁰

Treatment of patients with organisms resistant to isoniazid and rifampicin

Patients who fail on directly observed treatment containing isoniazid and rifampicin throughout, i.e., patients failing on a six-month first-line regimen or the above-mentioned eight-month re-treatment regimen, are more likely to harbor organisms resistant to both isoniazid and rifampicin (multidrug-resistant organisms). In most low-income countries such patients are designated "chronic excretors" whose fate has to be left to the natural course of the disease, as alternative drugs (other than the six essential drugs) are not usually available in sufficient quantity.

The emergence of multidrug-resistant tuberculosis has been documented in an increasing number of countries and has, in some countries, reached levels that seriously threaten tuberculosis control.^{621,622}

The WHO has addressed this issue in both a formal publication 606 and workshop proceedings. 608,609

Curative treatment of multidrug-resistant tuberculosis poses a multitude of problems. Amongst these are:

- the high cost of the necessary drugs (currently up to 100 times as expensive per course as a first-line regimen);⁶⁰⁹
- the relative weak activity of most of these drugs against *M. tuberculosis*;
- the high frequency of adverse reactions requiring specialist expertise;
- the prolonged duration (21 months has been proposed as a minimum);⁶⁰⁶
- the logistic difficulties anticipated in implementing such regimens in a national tuberculosis program;
- difficulties in implementing standardized laboratory facilities to correctly identify susceptibility patterns; ⁶²³
- gaps in knowledge as to what approach to treatment (individualized or standardized) is most appropriate. ⁶²⁴

As there is an increasing demand to utilize such alternative medications, and technical knowledge is generally poor about their proper usage in most countries where the problem has emerged or is emerging, the danger of uncontrolled usage is great. Resistance to these drugs is likely to emerge quickly in unprepared settings.⁶²⁵ It is hoped that the agenda set forth by WHO⁶⁰⁸ will generate sufficient information in an ordered and timely fashion and appropriate technical expertise will accompany implementation of any such project to ensure continued curability of tuberculosis in such settings. Unfortunately, multidrug-resistant tuberculosis has emerged precisely in areas of the world that have demonstrated poor tuberculosis control in the first place, and whether a deterioration of the situation in such settings can be prevented with the introduction of drugs potentially able to cure multidrug-resistant tuberculosis remains to be seen.

Case holding

Prescription of an adequate course of treatment is not sufficient; it must be ensured that the prescribed medications are also actually taken until the successful, curative completion of therapy.

Directly observed therapy

Ensuring regularity of treatment is the key to timely completion of therapy and the prevention of acquisition of drug resistance. The problems with self-administered chemotherapy in ensuring regular adherence have long been recognized,²⁷⁹ and to ascertain the efficacy of regimens in clinical trials, direct observation of drug intake during part or the entire course of treatment has thus been standard in many investigations.¹²²

Directly observed therapy refers to treatment where a qualified person (usually, but not always, ⁶²⁶ a health care worker) ensures that the prescribed medications are taken by observing the patient ingesting them. ⁶²⁷ Directly observed ambulatory therapy has its evidence base in studies in Chennai (formerly Madras) and Hong Kong, ⁶²⁸ and the recognition of the need for alternatives to costly hospitalization.

Directly observed therapy might be conceived of as a coercive procedure, but it may also help to strengthen the relationship between patient and health care worker.⁶²⁹ If this does not occur, then directly observed therapy may not achieve an increase in the proportion of patients completing therapy.⁶³⁰

The major effects of directly observed therapy that might be expected are a reduction in the risk of acquiring drug resistance and in the frequency of relapse following completion of chemotherapy, as convincingly demonstrated in a study in the United States (figure 52).⁶³¹

Can emergence of drug resistance be outpaced in a national tuberculosis program?

Strains resistant to isoniazid should have a comparative advantage, as patients harboring such a strain will, on average, be transmitters for a longer period of time than patients with a fully susceptible strain. Thus, one would expect an increase in the prevalence of primary resistance to isoniazid. This is, however, not the case in well-managed programs.^{632,633} Some studies sug-

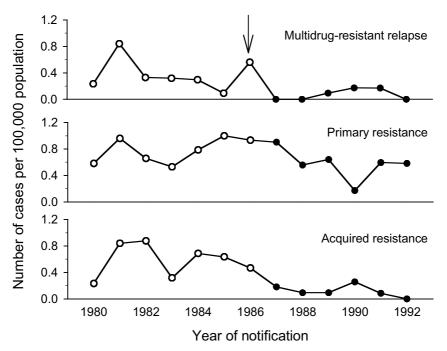


Figure 52. Effect of directly observed therapy on relapse, primary resistance, and acquired resistance, in Tarrant County, Texas, United States. Arrow indicates point in time of introduction of universal directly observed therapy. Reproduced from⁶³¹ by the permission of the publisher Massachusetts Medical Society.

gest that the transmissibility is the same for isoniazid-susceptible and isoniazid-resistant strains, 634,635 while others indicate that transmissibility of isoniazid-resistant strains is reduced;¹²² thus the question of transmissibility has not been fully resolved. However, strains which are resistant because of katG gene deletion have lower virulence in experimental animal models, 636,637 while mutation of the inhA gene has no effect on virulence. 636 Thus, a fraction of isoniazid-resistant strains may have a comparative selection disadvantage. In an effective tuberculosis program with a directly observed intensive phase utilizing the four most potent drugs, followed by a self-administered, non-rifampicin-containing continuation phase, no significant multidrug resistance (resistance to at least isoniazid and rifampicin) has emerged over 12 years of usage.⁶³² This could indicate that a qualitatively good program may outpace the rate of emergence of drug resistance. However, a study from The Netherlands indicates that some specific mutations of the *katG* gene lead to high-level resistance and as great a probability of producing secondary cases as isoniazid-susceptible strains. 638

Strains resistant to isoniazid alone can virtually always be killed when regimens containing both rifampicin and pyrazinamide are used and rifampicin is given throughout.⁶⁰³ The introduction of the short-course regimens in countries such as Algeria and Korea was accompanied by a clear decline in resistance to isoniazid and chronic excretors for this reason.^{639,640} However, in the case of Algeria, the introduction of short-course regimens was associated with the appearance of and slow increase in cases with multidrug resistance among previously treated patients, possibly related to the fact that directly-observed treatment was not the policy. The rate of decline in cases of tuberculosis (and particularly of re-treatment cases) in that community was greater than the rate of appearance of multidrug resistance, thus outpacing the drug resistance. However, if this community had experienced a rise in the numbers of cases, rather than a decline (as would have occurred if the community was affected heavily by HIV infection), this might not have been the case.

This is one of the main reasons why the IUATLD has preserved a very conservative policy with respect to treatment regimens, in order to preserve the usefulness of rifampicin as an efficacious agent in the overall scheme of treatment policy.

The approach to management of adverse drug events

The major clinical presentations of adverse drug events that may occur in a patient treated with the essential drugs and the approach to managing them will be discussed here. Adverse drug events from second-line drugs should always be dealt with by a specialist in the field. The discussion is limited to the major clinical syndromes occurring in the routine management of tuberculosis in clinical practice.

In any patient who takes prolonged treatment, episodes of ill health may occur which may be ascribed by the patient or the health care provider to adverse effects of the treatment given. This is not necessarily the case. In the large clinical trials of preventive chemotherapy carried out by the US Public Health Service among household contacts of tuberculosis patients, one group of patients was assigned to the treatment arm and another to a placebo in which identical tablets were given which contained no active medication.⁶⁴¹ Neither the patient nor the care provider knew the type of pills that individual patients were taking. The events that occurred during treatment were thus observed without knowledge of the treatment. In a number of cases, the health care provider, based on the assumption that the treatment was causing adverse drug events, discontinued the treatment.

When the code indicating what the patient was taking was broken and the results analyzed, it became apparent that 20% of all episodes considered to have been adverse drug events to the "medication" were, in fact, "placebo" effects.⁶⁴¹ This indicates that the "adverse events" were, indeed, intercurrent illnesses or events unrelated to the treatment itself, although they had every appearance of having been due to the medications.

This has important implications for the evaluation of "adverse events" in patients on treatment for tuberculosis. If one or other of the essential medications used in the treatment of tuberculosis (such as isoniazid or rifampicin) is stopped due to what is (incorrectly) perceived as an adverse drug event, the outcome of the treatment can be seriously affected. Discontinuation of an essential medication in the treatment of a tuberculosis patient for what is perceived as an adverse drug event must be carefully considered and correctly undertaken if the patient's chances of successful treatment are not to be seriously affected.

The patient with hepatitis

Clinical hepatitis is to be suspected in a patient presenting with a syndrome of malaise, nausea, vomiting, anorexia, fever, abdominal pain, hepatomegaly, jaundice or dark urine.⁶⁴²

Hepatic disease during anti-tuberculosis chemotherapy is not necessarily caused by the drugs, but may be attributable to other causes, such as alcohol abuse, cirrhosis, infectious hepatitis or indeed the tuberculosis itself. Nevertheless, appropriate management of the patient requires an approach as if one or more of the drugs were responsible.

The key suspect drugs are isoniazid, pyrazinamide, and rifampicin, if the patient is on any of these. In that case, such as in the intensive phase of chemotherapy, all three drugs should be stopped immediately if the symptoms are severe and/or if there is jaundice. The patient should temporarily be placed on ethambutol plus streptomycin in such a case. This combination is unlikely to be hepatotoxic and, while a relatively weak combination, still ensures temporary adequate treatment without a high risk of emerging drug resistance. In the presence of malaise and nausea only (without jaundice), rifampicin might in addition be kept in the regimen as it is rarely a cause of hepatitis. The patient is maintained on these two drugs until the acute symptoms subside, which usually occurs within one or two weeks.

Isoniazid might then be added in a dosage of 50 mg per day. If there is no clinical deterioration, the dose of isoniazid may then be increased to 100 mg on day 4, to 200 mg on day 7 and to the full dose on day 14.⁶⁴² Following the patient for another seven days, rifampicin might then be reintroduced, and if well tolerated, pyrazinamide might finally be added if rifampicin plus isoniazid has been well tolerated for seven days, and if it has been given for less than two months prior to the onset of hepatitis. This schedule can be expected to be successful in over 90% of cases.⁶⁴²

Some clinicians prefer to re-introduce isoniazid at the full dose when liver enzymes (where available) have normalized, or if liver enzyme tests are not available after two weeks (Schraufnagel DE, personal written communication, April 3, 2001; O'Brien RJ, personal written communication, April 19, 2001).

The patient with gastrointestinal symptoms

Gastrointestinal symptoms such as nausea, pain and vomiting might be prodromal symptoms of hepatitis, and close clinical observation is mandatory. In addition to isoniazid, rifampicin, and pyrazinamide, thioacetazone frequently causes gastrointestinal symptoms. In a patient on isoniazid and thioacetazone, the latter is probably the cause. Such reactions can often be dealt with easily by taking the medications with a meal or before going to bed. Monitoring of the response is important. If the symptoms do not subside, the isoniazid plus thioacetazone combination should be replaced by isoniazid plus ethambutol. Should the symptoms persist despite the change, isoniazid and the possibility of liver toxicity must be suspected and the patient be placed on streptomycin plus ethambutol until symptoms subside. Isoniazid might be re-introduced subsequently, as described above.

The patient with impaired vision

The most frequent drug-related cause of impaired vision among the medications used for treating tuberculosis is ethambutol. Optic toxicity is not detectable fundoscopically. If ethambutol is suspected, it must be withdrawn immediately and never be given again. If the event occurs in the intensive phase where ethambutol is given as a fourth companion drug, no replacement is necessary (although streptomycin might be used if deemed necessary). If the event occurs in the continuation phase when the patient is on isoniazid plus ethambutol, the latter should be replaced by thioacetazone or rifampicin.

The patient with vestibulo-cochlear toxicity

Vestibulo-cochlear toxicity is virtually always due to streptomycin. It is often, but not always, dose-dependent. Thus, it should first be checked whether the dosage given is appropriate to weight and age (toxicity increases with both). If the dose cannot be reduced or if dose reduction fails to improve the symptomatology, streptomycin should be stopped and not be given again (unless the drug resistance pattern makes its use imperative). As streptomycin is usually given only in the intensive phase as a fourth companion drug, it can be stopped without replacement. Streptomycin should never be given to pregnant women because of the potential risk of causing deafness in the unborn child.

The patient with neurologic symptoms

A distinction should be made between peripheral and central nervous system toxicity from anti-tuberculosis medications.

Peripheral neuropathy, presenting as paresthesia, such as tingling and numbness, starting at the feet with proximal spread is the usual manifestation.⁴⁰⁸ Myalgias, weakness, and ataxia may accompany these symptoms. Peripheral neuropathy is usually due to isoniazid, is rare and occurs usually only in malnourished or alcohol-dependent patients. Pyridoxine is effective in treating this condition, but the dosage for treatment should not exceed 50 mg per day, as there might be antagonism with isoniazid, ¹⁰⁸ although the clinical relevance of this antagonism is not clear.

Infrequently, toxic psychosis and epileptic convulsions may occur with isoniazid, and very rarely, in patients with signs of malnutrition or malabsorption, a pellagroid syndrome (with dermatitis, diarrhea, and dementia) has been reported. Pyridoxine is usually effective for treating such cases.

The patient with hypersensitivity reactions or muco-cutaneous signs and symptoms of toxicity

Cutaneous adverse drug events, ranging from pruritus, to rashes, and most severely to toxic epidermal necrolysis, sometimes accompanied by fever, may be caused by thioacetazone, isoniazid, rifampicin, streptomycin, or pyrazinamide. Cutaneous adverse drug events are much more frequent among patients with HIV infection than among non-HIV-infected patients. If the patient is on thioacetazone, it is by far the most likely cause. It should be stopped immediately, and never be given again.

In all instances of rash with or without fever, all drugs should be stopped. When the symptoms subside, usually within a day or two, the drug least likely to be the cause should be re-introduced in a test dose. This drug is usually isoniazid and is given at a dose of 150 mg. If the patient was hypersensitive to isoniazid, a rise in temperature, pruritus, or rash will develop within two to three hours.⁴⁹⁹ If there is no reaction to the test dose, the next test dose might be tried. Over the following days, the full dose is gradually introduced. Subsequently, rifampicin might be similarly re-introduced, starting with a test dose of 75 mg (or less), and so on. Under strict observation, it might be possible to desensitize with rifampicin much more rapidly, i.e., within two days.⁶⁴³ If there is pruritus or rash only, desensitization to isoniazid might not be necessary, as symptoms often subside spontaneously.

Very often desensitization is successful, and the full range of medications can be reintroduced within one to two weeks. It should be reiterated that such desensitization should never be attempted with thioacetazone.

The patient with hematologic abnormalities

Blood dyscrasias comprise only 10% of the total number of drug-induced adverse events but account for approximately 40% of fatal reactions related to drug administration.⁹³ They occur with all six essential anti-tuberculosis medications. In symptomatic patients, the offending drug should be withdrawn and never be given again.

Relative leukopenia and hemolytic anemia due to isoniazid require permanent withdrawal of the drug and often treatment with corticosteroids to reverse hemolysis. Sideroblastic anemia due to isoniazid is usually responsive to treatment with pyridoxine. Rarely, other neutropenia, eosinophilia, and thrombocytopenia may occur, which will respond to withdrawal of isoniazid. Similarly, the rare pure red cell aplasia responds to withdrawal of isoniazid. Complete recovery from agranulocytosis usually occurs following withdrawal of isoniazid.

With the exception of thioacetazone, blood dyscrasias due to anti-tuberculosis drugs are rare events. It is probably exceedingly difficult to identify the offending drug in the field.

The patient with acute renal toxicity

Acute renal toxicity may be the result of a hemolytic anemia, glomerulonephritis and interstitial nephritis. The most likely cause of this rare adverse drug event is rifampicin. The drug should be withdrawn and never be given again. If renal insufficiency has developed, the dosages of ethambutol and streptomycin must be reduced according to the remaining function as these drugs are almost entirely excreted through the kidneys.

The patient with osteo-articular pain

Arthralgia is a frequent adverse drug event resulting from accumulation of uric acid due to pyrazinamide. In many instances, the dosage of pyrazinamide is higher than that recommended in patients who have such reactions and, if so, should be reduced to within the recommended limits. It often occurs towards the end of the intensive phase, when pyrazinamide can be withdrawn without replacement. Alternatively, acetyl salicylic acid commonly alleviates the symptoms. Intermittent administration of pyrazinamide will also reduce the effect of uric acid retention. Allopurinol is ineffective.

The approach to the patient with pre-existing medical conditions

Patients may present not only with tuberculosis but also other medical conditions that require modifications of the standard treatment. In this chapter, some of the major medical conditions that require such adjustments are discussed.

The patient with liver injury

Patients with mild and clinically unrecognizable liver injury, including those who abuse alcohol, may be treated with the standard treatment, which needs to be adjusted only if clinical signs of hepatitis occur as discussed in the previous chapter.

Patients presenting with clinical signs of hepatitis should not be given the drugs with the greatest potential for hepatotoxic reactions. These include isoniazid, rifampicin, and pyrazinamide. Such a patient might be treated with ethambutol plus streptomycin until the acute signs of hepatitis subside. Subsequently, isoniazid and / or rifampicin might be re-introduced under close observation. Depending on the feasibility of introducing the latter, treatment duration will need to be adjusted. If neither rifampicin nor isoniazid can be given, treatment should probably be given for 18 months. The continuation phase with streptomycin and ethambutol should not be given more frequently than three times per week to reduce the cumulative toxicity of streptomycin.

The patient with renal failure

Streptomycin and ethambutol are excreted mainly through the kidneys and are thus safe only if appropriate dose adjustments can be made in patients with renal insufficiency. This is not usually possible without access to monitoring of blood levels or measurement of creatinine clearance, a service not usually available in low-income countries. Such a patient is thus best treated with isoniazid, rifampicin, and pyrazinamide in the intensive phase. In the continuation phase, isoniazid plus thioacetazone or isoniazid plus rifampicin can be given. Treatment duration is not affected.

The patient with impaired hearing or impaired balance

Patients with pre-existing vestibulo-cochlear impairment should not be given streptomycin. Streptomycin may be replaced by ethambutol.

The patient with impaired vision

Patients with impaired vision other than due to myopia, hyperopia or presbyopia, should not be given ethambutol. Ethambutol may be replaced by streptomycin in such cases.

The patient with gastrointestinal malabsorption

Patients recognized or suspected to have gastrointestinal malabsorption may pose serious problems for adequate chemotherapy, as shown in a study on risk factors for acquisition of rifampicin monoresistance.⁶⁴⁴ On the other hand, a study among HIV-infected patients in Nairobi has not demonstrated important differences in pharmacokinetic profiles of isoniazid, rifampicin, and ethambutol between patients with and patients without HIV infection,

and no association with diarrhea.⁵⁸⁴ Similarly, studies in South Africa have shown that malabsorption in asymptomatic HIV-infected patients is not a major issue and no important pharmacokinetic differences have been seen in a series of AIDS patients.⁵⁸⁵ Thus, malabsorption of anti-tuberculosis medications in HIV-infected patients may not be that serious a problem. Nevertheless, it is probably reasonable to always include the parenteral streptomycin in patients suspected of having malabsorption.

The pregnant patient

Pregnant women with tuberculosis do not pose particular problems for treatment. Dose adjustment is probably indicated with increasing body weight as the volume of distribution increases. Because of the potential of vestibulo-cochlear toxicity to the fetus, streptomycin should not be given in pregnancy. Isoniazid, rifampicin, ethambutol, pyrazinamide, and thioacetazone are safe in pregnancy, and are not reported to have teratogenic or other adverse effects on the fetus.

Second-line drugs that should be avoided in pregnancy include other aminoglycosides, polypeptides, thioamides, and quinolones.

2. Prophylactic treatment

In this monograph, prophylactic treatment is defined as treatment to prevent acquisition of infection with M. *tuberculosis* in a person exposed to tubercle bacilli. Its aim is to minimize the risk of acquiring latent infection.

Little evidence is available to document the efficacy of such prophylactic treatment. The little that is known is summarized here.

Rationale and experiences with prophylactic treatment

In the early 1950s, Zorini reported experiments with prophylactic treatment in guinea pigs, using various dosages of isoniazid. 645,646 Briefly, guinea pigs were given isoniazid or placebo in their drinking water for one month and then challenged with an endoperitoneal injection of *M. tuberculosis*. The results were unequivocal in that a considerably larger proportion of placebo-treated animals developed tuberculosis in comparison to those receiving isoniazid.

Among humans, the effect of isoniazid compared to placebo in preventing tuberculin skin test conversion has been ascertained within the context of clinical trials on preventive chemotherapy.⁶⁴¹ A tuberculin skin test was given before random allocation to either isoniazid or placebo for one year. At the end of treatment, the rate of conversion among persons who were initially tuberculin skin test negative was compared in the two groups. These four US Public Health Service studies were conducted among various groups of patients (patients in a mental institution, contacts of known cases, school children, and contacts of newly diagnosed tuberculosis patients). The protection afforded against conversion from a negative to a positive tuberculin skin test after one year of treatment with isoniazid in these studies is summarized in figure 53.⁶⁴¹ It shows that the confidence intervals are wide (small numbers eligible for assessment), and thus that the extent of protection is uncertain.

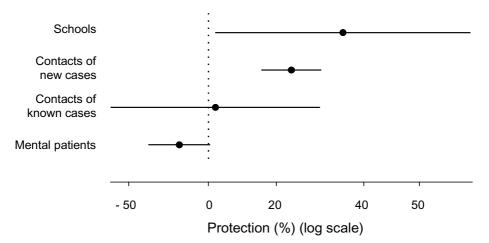


Figure 53. Protection from prophylactic treatment in the prevention of acquisition of tuberculous infection in four clinical trials conducted by the US Public Health Service.⁶⁴¹

Indications and recommendations for the use of prophylactic treatment

Prophylactic treatment is, for all practical purposes, rarely indicated. Even if the evidence is scant, however, it makes sense to provide it to a newborn child with a potentially infectious parent, especially the mother. This is recommended in industrialized countries, ³⁵⁶ but should most likely be a universal indication.

It is not clear what the appropriate duration of prophylactic treatment should be. It is probably indicated, however, to continue it for perhaps up to three months after relevant exposure has ended.

Children under the age of five years are also at high risk of acquiring tuberculous infection from a person with sputum smear-positive tuberculosis living in the same household and, if they become infected, are at high risk of progression to clinically manifest tuberculosis. The IUATLD has thus recommended systematic treatment with isoniazid of asymptomatic children in such a situation.⁸ Some of these children will not yet have been infected (the infected being the primary target group) and will thus receive true prophylactic treatment.

Early vaccine development

Vaccination with Mycobacterium tuberculosis

Early in the twentieth century, von Behring attempted vaccination (or as he called it, "Jennerization") of cattle by utilizing increasing doses of living *M. tuberculosis*.^{647,648} Similar to these attempts, Webb in the United States tried to make experimental animals resistant to re-challenge with increasing doses of virulent *M. tuberculosis*, and a few children were also "vaccinated" with this approach, apparently with no adverse outcome.⁶⁴⁹ While this approach seemed indeed to provide some protection against a subsequent challenge in cattle and other experimental animals compared to controls, protection was incomplete in the case of von Behring's "bovovaccination" and in the guinea pig. Furthermore, with "Jennerization" in cattle there was the potential that the microorganism would appear in milk.⁶⁵⁰ Theobald Smith also pointed out that the unknown duration of the incubation period carried great dangers, even if the immediate effect seemed to be innocuous.⁶⁵⁰ This approach was therefore only short-lived.

However, more recently, the idea of attenuating *M. tuberculosis* and using such an attenuated strain as a vaccine has been picked up again, and it is expected that the vaccine properties of such mutants will be tested at least experimentally in the near future.⁶⁵¹

Vaccination with Mycobacterium chelonae

Early in the twentieth century, Friedmann proposed vaccination with *M. chelonae*, a mycobacterium recovered from the turtle.⁶⁵² Because the argument of vaccination was largely based on the hypothesis that persons ill with tuberculosis could develop increased resistance in suppressing progression from morbidity to death, this method was mainly used in the treatment of clinically manifest tuberculosis.⁶⁵³ There was probably no effect at all if judged by current standards. Only a very small study was published reporting the results of *M. chelonae* vaccination in children exposed to tuberculosis but without clinical signs of the disease.⁶⁵⁴ The study was

too small to allow a meaningful interpretation of the efficacy of this vaccination. The method never gained much attention beyond Germany, and fell into oblivion as vaccination with BCG drew increasing attention in the immediately succeeding years.

Vaccination with BCG

Vaccine development

A virulent strain of *M. bovis*, isolated by Nocard in 1902, from milk obtained from a cow with tuberculous mastitis⁶⁵⁵ was inoculated for the first time on January 8, 1908, by Albert Calmette (1863-1933) and Camille Guérin (1872-1961)⁶⁵⁶ at the Pasteur Institute in Lille, France⁶⁵⁷ onto a medium consisting of cooked potato and glycerinated bile.

The strain, to become known as Bacille Calmette-Guérin (BCG), was sub-cultured in 230 passages on bile potato medium until 1921 when it no longer changed its characteristics.

After thirty passages the strain ceased to kill guinea pigs; after sixty it was still slightly virulent for rabbits and horses, but avirulent for guineapigs, monkeys, and calves.⁶⁵⁵ From 1912 onwards, experiments were conducted among calves, demonstrating their resistance to subsequent infection with virulent bacilli.⁶⁵⁵ It may be noted that the main objective in the development of this vaccine was to obtain an effective vaccine against tuberculosis in goats ⁶⁵⁸ and cattle.^{658,659} It is now clear that it was not the glycerinated bile medium that was the reason for the loss of virulence.^{660,661} By sub-culturing four bovine strains on Calmette's bile-potato medium over six years, Griffith failed to reproduce Calmette's finding and to induce stable attenuation.⁶⁶² The reasons for the loss of virulence of *M. bovis* BCG remain unclear until today.

On July 1, 1921, Weill-Hallé, a pediatrician, requested the vaccine for use in an infant born to a mother who had died of tuberculosis shortly after delivery. The child was to be brought up by a grandmother who was herself suffering from tuberculosis.⁶⁶³ The child was given 6 mg of BCG orally and developed normally over the next six months without any sign of illness, either from the vaccine or from tuberculosis.^{655,663} Over the next three years, 317 infants (67 of whom were born into, and brought up by families with tuberculosis patients) were vaccinated with 30 mg oral BCG vaccine, given in three portions at 48-hour intervals.

Following these early experiments in humans, BCG was distributed to a large number of laboratories, largely in Europe, and given to hundreds of thousands of children within a decade after its introduction. ⁶⁶⁴⁻⁶⁶⁷ Trials to evaluate its impact began in Europe ⁶⁶⁸⁻⁶⁷⁰ and North America. ^{671,672}

Controlled assessment of the vaccine's efficacy was conspicuously absent, and one of its most violent opponents was Petroff in the USA, who doubted both the vaccine's innocuousness and efficacy.^{673,674} Despite the justified concerns about the quality of the data on efficacy given all the methodological problems (such as selection bias), it seemed apparent that BCG reduced case fatality from tuberculosis among exposed children in a variety of settings (figure 54).⁶⁶⁶ It also seemed to protect adult student nurses heavily exposed to tuberculosis both from death and disease (figure 55).^{668-670,675}

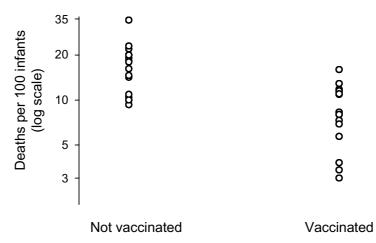


Figure 54. Early, non-controlled comparisons in crude infant mortality before and after introduction of BCG vaccination in 16 countries, reported up to 1932.⁶⁶⁶

The assumption of the safety of BCG vaccination was severely challenged when 72 of 251 children who were presumably vaccinated with BCG between December 10, 1929, and April 30, 1930, died from tuberculosis in Lübeck, Germany.⁶⁷⁶⁻⁶⁷⁸ While not all circumstances surrounding this disaster have ever become public, ⁶⁷⁹ it soon became apparent that BCG was not the cause. The preliminary epidemiologic analysis in July 1930 already showed large differences in case fatality by week of vaccination (figure 56), indicating that strains with different virulence had been mixed. ⁶⁸⁰ This was bacteriologically confirmed by demonstrating that virulent tubercle bacilli, but not BCG, were consistently isolated on autopsy.⁶⁷⁶ The epidemiologic

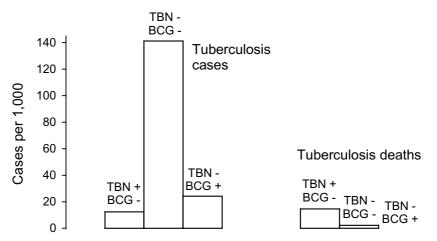
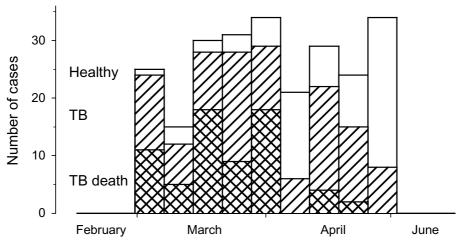


Figure 55. Results from a non-randomized, self-selection evaluation of the effect of BCG vaccination on tuberculosis cases and deaths among student nurses in Norway.⁶⁷⁰



Weekly vaccinations with critical strain

Figure 56. Curve of the tuberculosis epidemic following an accidental mix of BCG vaccine strain with a virulent strain of *Mycobacterium tuberculosis* in Lübeck, Germany, 1930.⁶⁷⁶

and bacteriologic investigations demonstrated conclusively that batches containing both BCG and *M. tuberculosis* in varying proportions had been fed to the infants during the epidemic.^{676,678,681,682} Among the 53 fatal cases ascertained by mid-July 1930, the interval between vaccination and death ranged from 34 to 129 days with a median of 79 days. Petroff's concerns about a reversion to virulence of BCG have never been confirmed, and his observation of different colony morphology with virulent and avirulent colonies⁶⁷³ have not been confirmed elsewhere.⁶⁷⁶

The BCG strain family

Until the introduction of freeze-drying in Japan in 1943,⁶⁸³ the only means of maintaining a viable strain was through sub-culturing. With the distribution of the vaccine strain to multiple laboratories in the world, each using slightly different techniques for strain maintenance, it is not surprising that the BCG family shows large diversity.⁶⁶⁰ The first freeze-dried French strain (1949) from the Pasteur Institute in Paris was strain 1173-P₂, from which the Glaxo and Danish strains descended.⁶⁸⁴

Recent work based on molecular characterization of the various substrains points to various mutations that have occurred at different points in time (figure 57),⁶⁸⁵⁻⁶⁸⁷ and indicates that the various BCG sub-strains are morphologically and genetically different from each other.

Safety record of BCG vaccination

A large review has shown BCG to be one of the safest vaccines.^{688,689} The demarcation between a normal reaction and an adverse reaction is not always clear.⁶⁹⁰ The normal reaction is a red indurated area measuring five to 15 mm. A crust is formed around this induration, which is soft at the center for three to four weeks. At six to ten weeks, the crust falls off, leaving a flat scar measuring three to seven millimeters.⁶⁹⁰ Regional lymphadenopathy in the absence of erythema or vesicle formation should also be considered a normal reaction to the vaccine.⁶⁹¹ Complications include cutaneous lesions and regional suppurative lymphadenitis; more severe localized or multiple lesions (such as musculo-skeletal lesions); 692-694 and nonfatal and fatal complications resulting from hypersensitivity reactions or mycobacterial dissemination. 688,689,695-702 The risk of complications varies with the type of vaccine and with the age at vaccination. The risk of osteomyelitis ranged from 0.01 to 50 per 1 million vaccinations, that of multiple or generalized lesions from 0.01 to 2 and that of fatal cases from 0.01 to 1 per million vaccinated individuals.^{688,689} The lowest complication rates were reported with the Tokyo strain, and the highest with the Gothenburg strain produced in Denmark. 692,703

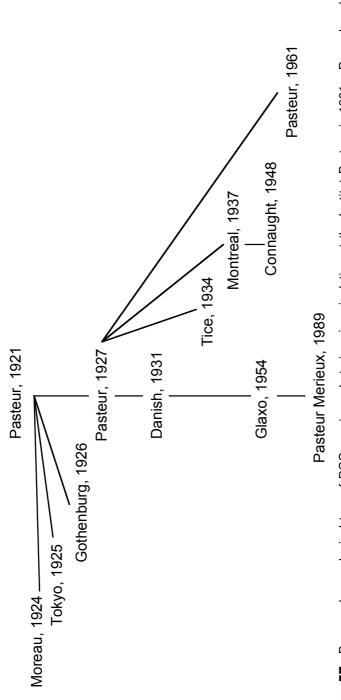


Figure 57. Proposed genealogical tree of BCG vaccine substrains since isolation at the Institut Pasteur in 1921. Reproduced from686 by the permission of the publisher Churchill Livingstone.

In a prospective study in South Africa among 10,000 neonates receiving the Copenhagen strain intradermally at birth, at six weeks post vaccination the vaccination scar had healed in more than 95% of children, 1.5% had no vaccination scar, and in 3% adverse events were noted.⁷⁰⁴ All adverse events were local (oozing, abscesses, rarely combined with lymphadenopathy).

Because BCG is a live vaccine, concerns were raised early on about the safety of its use in persons infected with HIV, ⁷⁰⁵⁻⁷⁰⁷ and several case reports about disseminated mycobacteriosis ⁷⁰⁸⁻⁷¹⁴ and mycobacterial meningitis due to BCG ^{708,710,715} have been published. A study among motherchild pairs with and without HIV infection has shown that children of mothers with HIV infection who also had HIV infection themselves had a slightly increased risk of suppurative lymphadenitis, but the manifestations were mild and easily manageable (figure 58). ⁷¹⁶ Apparently, living BCG can persist for decades and cause localized ⁷¹⁷ or disseminated ⁷¹⁸ complications after acquisition of immunosuppression. Nevertheless, most of these case reports appear to be isolated events, although it has been argued that disseminated disease attributable to BCG vaccination in HIV-infected children might be exceedingly difficult to diagnose. ⁷¹⁹ However, a study in Zambia among HIV-symptomatic children with a median age of 15 months, showed that mycobacteremia due to BCG must be exceedingly rare. ⁷²⁰ A recom-

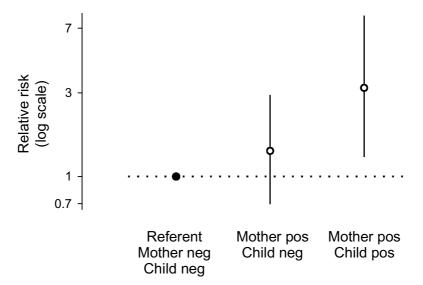


Figure 58. Relative risk of a complication following BCG vaccination among children born to an HIV-infected mother.⁷¹⁶

mendation by WHO states that no principal changes in BCG vaccine policy are warranted unless children present with symptomatic HIV infection,⁷²¹ a statement that has not been challenged.^{722,723}

Management of adverse reactions due to BCG vaccination

Children with lymphadenitis due to BCG were randomly allocated to receive either isoniazid or no treatment.⁷²⁴ There was no difference in the duration of lymphadenitis between the two groups, nor did isoniazid prevent the occurrence of suppuration. Similarly, children with abscess formation were randomly assigned to receive either isoniazid or erythromycin (serving as placebo).⁷²⁵ The response in each treatment group was the same. In another study, comparing excision, excision plus isoniazid, and isoniazid alone compared to a control group without intervention, no significant differences were observed between the various interventions, and in particular, isoniazid offered no advantage.⁷²⁶ Non-suppurative lymphadenitis is a normal reaction, and is best left without antibiotic treatment.^{690,727}

Patients with suppurative lymphadenitis following BCG vaccination were randomly assigned to treatment with simple needle aspiration, introducing the needle subcutaneously two to three centimeters distant from the node, versus no treatment.⁷²⁸ Regression was significantly faster in the treated than in the non-treated group, and spontaneous drainage was less frequent.

For osteoarticular mycobacteriosis due to BCG, combination therapy is indicated, but results were not always favorable (both in terms of sequelae and relapses) in a case series from Sweden.⁷²⁹

A standard course of treatment (as for clinically manifest tuberculosis) is also indicated in disseminated mycobacteriosis due to BCG. As this is a rare complication, however, treatment regimens have not been amenable to formal study. In treatment, it should be kept in mind that BCG is, like its parent organism, *M. bovis*, naturally resistant to pyrazinamide.

Efficacy and effectiveness of BCG vaccination

Efficacy is the extent to which an intervention produces a beneficial result under ideal conditions. The best setting to address efficacy is thus prospectively, in a controlled clinical trial. In contrast, effectiveness takes the various constraints that are found in the field into account in the actual routine delivery of the intervention.⁷³⁰ Effectiveness is often ascertained retrospectively, such as in case-control studies. Efficacy (in clinical trials) and effectiveness (in case-control studies) have been ascertained in various settings. The principle underlying the design of prospective and retrospective studies is summarized in table 10. These trials were supplemented by community trials and contact studies. The variation in estimates of protection ranged widely, from harm (more cases among the vaccinated than among controls) to a high level of protection.

The efficacy of BCG vaccination is best ascertained in a prospective clinical trial, while an estimate of its effectiveness in routine application might be obtained through retrospective studies, such as case-control, contact, or case-population studies, although possible confounding effects cannot be controlled so easily.

Briefly, clinical trials are a prospective ascertainment of cases occurring among the exposed. Clinical trials thus start with looking at the exposure (BCG vaccination given or not) and then ascertain the outcome (tuber-culosis) in a group of individuals, preferably randomly assigned to exposure (table 10).⁷³¹ These are population-based studies and the denominator is

Table 10. Study design of clinical trials and case-control studies.

tic Characteristic Person-time absent of observation – E – F
_ F+F
– E+F

Design of a clinical trial

Design of a case-control study

	Outcome		
Exposure	Case	Control	Total
Exposure present	а	b	a+b
Exposure absent	С	d	c+d
Total	a+c	b+d	N=a+b+c+d
Odds among the exposed: a / b Odds among the unexposed: c / d Relative odds: (a / b) / (c / d).			

the number of person-years of observation. The measures are incidence rates among the exposed and unexposed and the summary measure is the relative risk (the risk among the exposed divided by the risk among the unexposed). Vaccine efficacy (in per cent) is calculated as $(1 - \text{relative risk}) \times 100$.⁷³² The 95% confidence intervals were calculated (or recalculated, where appropriate) using the formula proposed by Orenstein in his review on assessment of vaccine efficacy,⁷³² unless adjusted or stratified summary estimates were provided by the authors.

To defray the costs incurred in clinical trials and to obtain results more quickly, it was proposed to ascertain the effectiveness of BCG vaccination by means of (retrospective) case-control studies.⁷³³ Briefly, case-control studies start with looking at the outcome (tuberculosis) and then ascertain exposure (BCG vaccination given or not) in a group of patients with the outcome, compared to an appropriately selected control group of persons without the outcome (table 10).⁷³⁴ A relative risk cannot be calculated as this measurement is confined to population-based studies. The measurement of risk in a case-control study is the odds ratio (or relative odds). For rare diseases the odds ratio approximates the relative risk in a clinical trial.

The advantages and disadvantages in the use of the case-control approach are linked to its being observational, having subjects selected on the basis of disease status, and using controls from the population from which the cases emanated.⁷³⁵ The advantages of case-control studies include avoidance of ethical problems arising in situations where there is already evidence that the vaccine is better than placebo; allowing much faster conduct than randomized trials; and requiring a much smaller number of subjects. They are thus substantially cheaper to conduct than randomized clinical trials.⁷³⁵

The most challenging difficulty in the design of case-control studies is the selection of appropriate controls in that they have to be selected in such a way that they are comparable to cases in every respect except for the outcome. Selection bias resulting from a failure to ensure this comparability may thus invalidate any findings.

The results of some of these case-control studies are summarized below. Vaccine effectiveness (in per cent) from a case-control study is estimated as $(1 - \text{odds ratio}) \times 100.^{732}$ For unmatched case-control studies, the 95% confidence intervals were calculated (or recalculated where appropriate) using Woolf's method.⁷³⁴ For matched and adjusted analyses, the confidence interval published by the authors of the study was chosen. If not stated for matched studies, the confidence interval around the crude odds ratio was calculated as above.

Prospective and retrospective studies on BCG vaccination

In one of the first clinical trials with a methodologically fairly acceptable design (systematic alternate allocation), BCG was given to children exposed to a parent with tuberculosis and compared to a similar group who did not receive the vaccine.⁷³⁶ The impact on fatality was dramatic, with an 82% reduction in the risk (figure 59). Nevertheless, suspicion about the efficacy of BCG vaccination persisted, particularly in the United States,⁷³⁷ but also in the United Kingdom,⁶⁷⁹ largely because the design of many studies was dubious at best.

One of the most conspicuous differences observed in the protection afforded by BCG reveals that age at vaccination is important. Of further crucial importance is the type of tuberculosis that is targeted for protection by vaccination.

In the following summary of the best-known studies in the English literature, the studies are identified as being prospective or retrospective. For each of these two study types five classes were examined:

- protection against disseminated and meningeal tuberculosis, and against death from tuberculosis;
- protection afforded to children by vaccination of newborns or infants;
- protection afforded by vaccinating children beyond the age of one year;

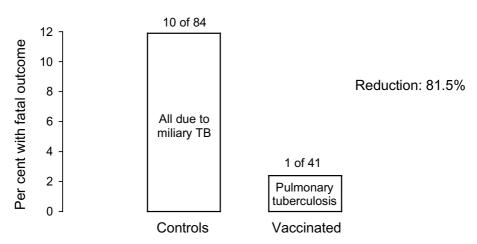


Figure 59. Comparative case fatality from tuberculosis among newborns vaccinated and not vaccinated with BCG in a clinical trial with systematic assignment to the experimental or control arm.⁷³⁶

- protection afforded by vaccinating adolescents or adults;
- protection afforded by vaccinating people of various ages.

Protection conferred by BCG vaccination against disseminated and meningeal tuberculosis, and against death from tuberculosis

Five major prospective studies have looked into the protection afforded by BCG vaccination against death from tuberculosis (figure 60). ^{736,738-744} All of these studies were conducted before the advent of curative chemotherapy. Four of the studies showed a point estimate of the protective efficacy of 80% and above, and one afforded no protection. The confidence interval was wide in all studies, because the number of events was small.

Several retrospective studies (including two using two different control groups) examined the protection against disseminated and meningeal tuberculosis (figure 61).^{745-747,747-753} The protective effectiveness was usually in excess of 80% and in no case did the 95% confidence interval include zero.

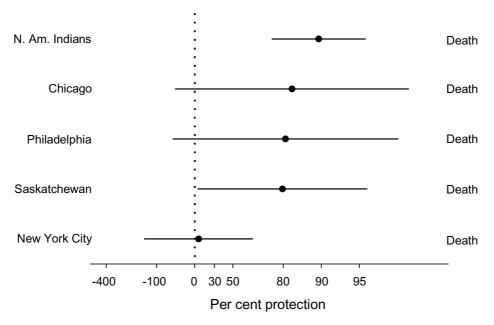


Figure 60. Results from five controlled clinical trials to evaluate the efficacy of BCG vaccination against death from tuberculosis.^{736,741-744}

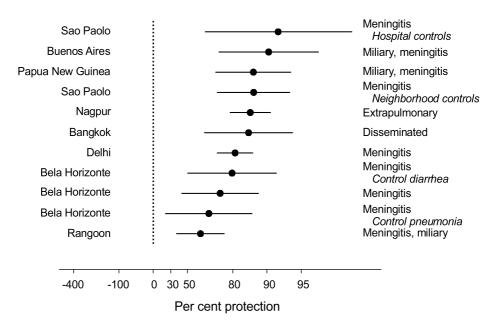


Figure 61. Results from retrospective studies on the effectiveness of BCG vaccination against death from meningeal, other extrapulmonary, or disseminated tuberculosis.^{745-747,747-753}

It may be concluded from these studies that BCG affords very good protection against death from tuberculosis, and against disseminated and meningeal tuberculosis.

Protection conferred by BCG vaccination of newborns and infants

Three prospective studies looked into the protective efficacy of BCG given to newborns or infants against all forms of tuberculosis or morbidity (figure 62).^{742,743,754} The point estimate of the efficacy was between 50% and 80%.

Several retrospective studies examined the effectiveness of newborn or infant vaccination (figure 63).^{747,748,753,755-762} The level of protection in these studies varies widely, but frequently above 50%. Noteworthy is the study from Zambia, which stratified effectiveness estimates by HIV status,⁷⁶⁰ showing that HIV-infected children had no protection as compared to 60% protection among HIV-negative children.

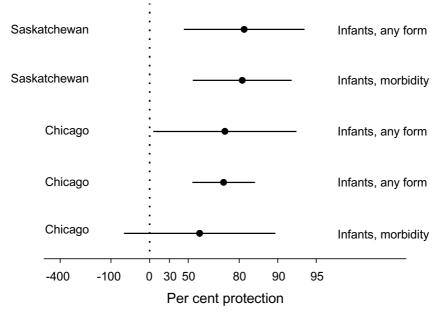


Figure 62. Results from prospective studies on the efficacy of BCG vaccination against tuberculosis in newborns and infants.^{742,743,754}

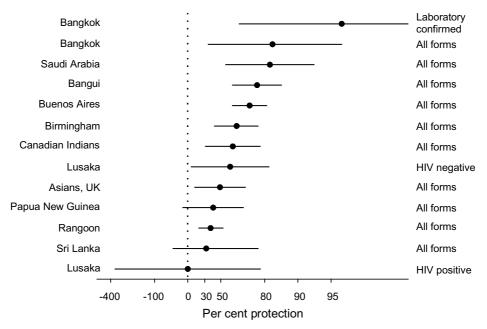


Figure 63. Results from retrospective studies on the effectiveness of BCG vaccination against tuberculosis in newborns and infants.^{747,748,753,755-762}

Protection conferred by BCG vaccination of children over one year of age

Only three protective studies of BCG vaccination of older children are available (figure 64).⁷⁶³⁻⁷⁶⁸ All three showed a very low level of protection, of less than 30%. In Chingleput, south India, where BCG gave little or no protection, there was a tendency to provide some protection in children below the age of 15 years, but a similar tendency towards harm (more cases in the vaccinated than the non-vaccinated) in older persons (figure 65).⁷⁶⁵

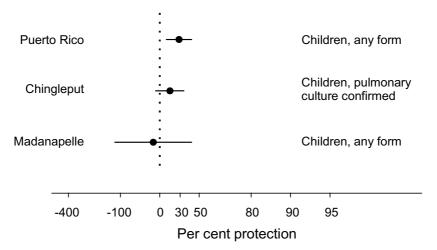


Figure 64. Results from prospective studies on the efficacy of BCG vaccination against tuberculosis in children other than infants.⁷⁶³⁻⁷⁶⁸



Figure 65. Protection from BCG vaccination by age, Chingleput, India.⁷⁶⁵

Three retrospective studies among children also showed very variable levels of protection, from 16% to 74% (figure 66).^{750,769,770}

These studies seem to show that vaccination of older children does not offer protection against tuberculosis that is as reliable as vaccination at an earlier age.

Protection conferred by BCG vaccination among adolescents and adults

Six prospective studies have examined the protection of BCG vaccination against tuberculosis among adolescents or adults (figure 67). ^{668,670,763-765,767, 768,771-778} The study in Ulleval, Norway, was the first ever conducted prospective study. ^{668,670} It does, however, not live up to current requirements for a controlled trial, as student nurses with a negative tuberculin skin test at entry could choose whether to be vaccinated or not. In this context, the study conducted in England (where *M. microti*, not BCG was used) remains the only study of high standard that has shown a very high level of protection, of close to 80%, in this age group. ⁷⁷¹⁻⁷⁷⁶ The other studies show little or no protection, with a tendency to reveal a potentially harmful effect in India. ^{763-765,777} In England, protection appeared to last for about 10 years before dropping rapidly (figure 68). ⁷⁷⁶ In contrast, in Chingleput, where there was no overall protection, vaccination appeared to confer harm (more cases than in the control group) in the first five years and minimal protection subsequently (figure 69). ⁷⁶⁵

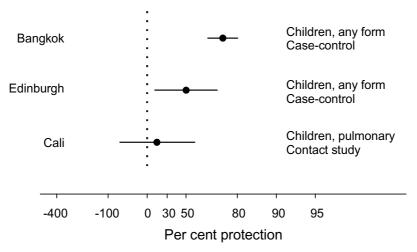


Figure 66. Results from retrospective studies on the effectiveness of BCG vaccination against tuberculosis in children other than infants.^{750,769,770}

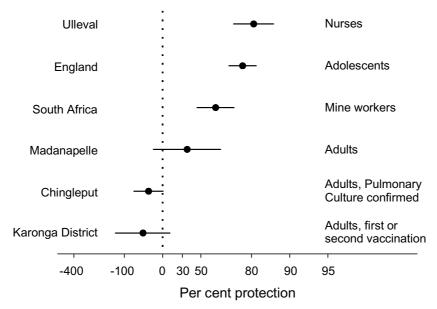


Figure 67. Results from prospective studies on the efficacy of BCG vaccination against tuberculosis in adults.^{668,670,763-765,767,768,771-778}

The two retrospective studies show a protective effectiveness of 10% ⁷⁷⁹ and close to 60%, ⁷⁸⁰ respectively (figure 70).

These studies seem to indicate that vaccination of adolescents or adults is rarely a useful intervention.

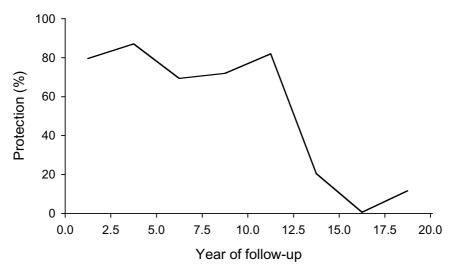


Figure 68. Protection from BCG vaccination among British school children during follow-up.⁷⁷⁶

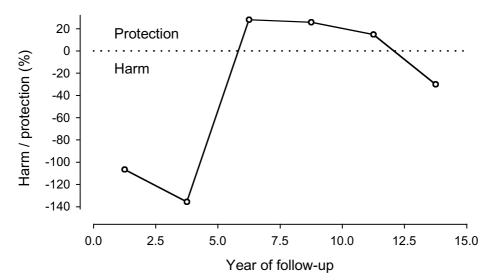


Figure 69. Protection from BCG vaccination in Chingleput, India during followup.⁷⁶⁵

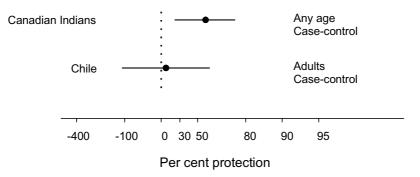


Figure 70. Results from retrospective studies on the efficacy of BCG vaccination against tuberculosis in adults.^{779,780}

Protection conferred by BCG vaccination across various age groups

Of the seven clinical trials studying protective efficacy across a wide range of age groups, with a preponderance of persons other than infants, two showed a high level of protection, of around 80%, while all of the others showed little or no protection (figure 71).^{738,763-765,767,768,781-787}

These observations reconfirm that utilization of BCG vaccination in age groups other than infants is rarely an effective intervention.

One retrospective study from the Gambia reported that 35 patients among 200 without a BCG scar died during chemotherapy, while none of

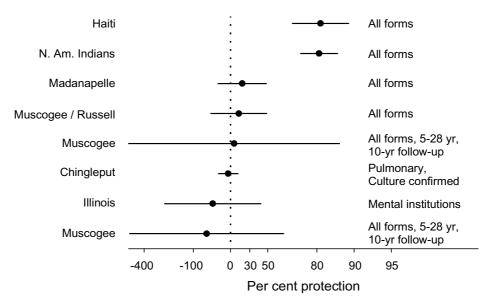


Figure 71. Results from prospective studies on the efficacy of BCG vaccination against tuberculosis in all ages.^{738,763-765,767,768,781-787}

85 with a BCG scar did so.⁷⁸⁸ While considerable attention was paid to adjustment for potential confounding factors (yet the effect remained), the authors were still cautious in concluding that BCG vaccination reduces case fatality from pulmonary tuberculosis.

Hypotheses about the variation in the efficacy of BCG vaccination

While the overall evidence is quite clearly in favor of a protective effect of BCG vaccination, the observed variations are large in both prospective and retrospective studies. A number of hypotheses have been formulated to address these discrepancies. Smith⁷⁸⁹ and Smith and Fine⁷⁹⁰ have comprehensively reviewed the evidence, and the following outline is guided by, and draws heavily on, their assessment.

The principal hypotheses to explain the variations observed in the protection offered by BCG include:

- Differences in methodological stringency;
- Differences in vaccine strains;
- Differences in vaccine dose;

- Differences in virulence of *M. tuberculosis* strains;
- Differences in risk attributable to exogenous reinfection tuberculosis;
- Differences in genetic make-up of vaccinees;
- Differences in nutritional status of vaccinees;
- Differences in prevalence of infection with environmental mycobacteria;
- Other factors.

Differences in methodological stringency

Quite obviously, not every study can be methodologically as rigorously conducted as ideal standards of study design and conduct call for.^{731,734} Among the clinical trials, several have been excluded from major reviews and metaanalyses such as those conducted by Colditz and collaborators.^{791,792} These authors found that study validity score explained 66% of the variation in prospective clinical trials and 36% in retrospective case-control studies,⁷⁹¹ and only 15% in case-control studies on BCG protection against infant tuberculosis.⁷⁹² Nevertheless, perhaps the most relevant trial showing no protection against bacteriologically confirmed tuberculosis, conducted in Chingleput, India, was judged to be of high scientific quality by a WHO expert committee specifically charged to ascertain the trial's validity.⁷⁹³

It must be kept in mind that the range of protection cannot be taken at face value, but must also be seen in the context of what the study in question sought to address. BCG trials (be they prospective or retrospective) ascertained protection against various outcomes such as morbid state (tuberculosis or death from tuberculosis) and site of disease, e.g., pulmonary, extrapulmonary single site, and disseminated tuberculosis, taking into account such things as bacteriologic certainty of the case, age of the patients, and time elapsed since vaccination. What seems apparent from the studies is the tendency of BCG to provide its greatest protection within the few years following vaccination, against death from tuberculosis, disseminated disease manifestations, and bacteriologically unconfirmed tuberculosis. In summarizing these effects, BCG is generally most effective against serious forms of tuberculosis occurring shortly after infection acquired at an early age. Thus, any evaluation of the protective efficacy of BCG vaccination should be stratified according to these variables.

Differences in vaccine strains

The available BCG vaccine strains differ widely in phenotype and genotype. 660,661,685,686 It has been proposed 794 that differences in vaccine strains may account for observed variations in vaccine efficacy. In the rabbit model, not all BCG (and *M. microti*) strains provided the same level of protection. 795 However, the most powerful argument against this hypothesis arises from the Chingleput study, where two vaccine strains were used $^{763-765}$ that had documented high efficacy in other settings but were not shown to be efficacious in Chingleput. Furthermore, one of the studies (a case-control study from Indonesia) cited for evidence of differential effectiveness of strains, examined successive vaccination policies, and was thus by necessity a non-concurrent study which additionally failed to adjust for time elapsed since vaccination. 796

Differences in vaccine dose

BCG has been administered through various routes, initially orally, then parenterally. The latter administration may have been given intradermally or transdermally via multipuncture devices. The dosage reaching the target thus may well have varied. Nevertheless, the following observations seem to contradict the argument of an influence of differential dosage effect. Three controlled clinical trials with low efficacy used multipuncture administration, ^{768,785,786} and one with high efficacy did so too. ⁷⁴² Furthermore, the trial in Chingleput specifically considered in its design the possibility of deterioration of vaccine potency in the field, and allocated vaccinees also to two arms receiving a ten-fold difference in dose, with no difference in effect. ⁷⁶³⁻⁷⁶⁵

Differences in virulence of M. tuberculosis strains

That not all tubercle bacilli are equally virulent has been demonstrated repeatedly both for *M. bovis* BCG⁷⁹⁷ and *M. tuberculosis* in general, ^{798,799} and for isoniazid-resistant strains in particular. ^{38,800-802}

The hypothesis that the relative frequency of more or less virulent tubercle bacilli affects the observed protective efficacy of BCG vaccination is based on the argument that tubercle bacilli of lower virulence might also cause tuberculin skin test reactions of smaller size. Such persons then might be classified as "non-reactors", i.e., persons not infected with tubercle bacilli, thus becoming eligible for vaccination. Vaccination of actually infected persons may thus mask any protective effect of BCG vaccination, as vaccination is not expected to provide protection against those who are already infected. 803

The argument fails to account for the fact that BCG provided no protection at all in some trials. Depending on the proportion of individuals who had escaped infection with environmental mycobacteria at the point of BCG vaccination, masking of protection by BCG vaccination would be expected to be incomplete.

Differences in risk attributable to exogenous re-infection tuberculosis

BCG vaccination is expected to provide protection against tuberculosis resulting from infection acquired subsequent to vaccination. It is not expected to provide greater protection than a naturally acquired primary infection. Protection conferred by a primary infection against disease from re-infection is incomplete.⁸⁰⁴⁻⁸¹¹ Thus, the protective efficacy of BCG might be increasingly masked as the contributory fraction of cases attributable to re-infection increases.^{812,813} Thus, following this argument, the protection afforded by BCG is expected to be lower where the risk of infection with *M. tuberculosis* (and thus re-infection) is high.

This is not borne out by observations. The annual risk of infection in the United Kingdom decreased considerably over time,⁸¹⁴ yet the level of protection afforded by BCG remained high and virtually unchanged.⁷⁶¹

Differences in genetic make-up of vaccinees

Because differences in protection from BCG among males and females were observed in at least one study, ⁷⁶⁶ other genetic factors may also play a role in the differential protection conferred by BCG. Nevertheless, the finding that BCG gave virtually no protection to children in Chingleput, ⁷⁶⁵ but high protection in children from the Indian sub-continent living in the United Kingdom ^{758,761} would tend to disfavor this hypothesis.

Differences in nutritional status of vaccinees

As nutritional status affects the functioning of the cellular immune system, it might be expected that poor nutritional status would adversely affect the protective efficacy of BCG vaccination. However, BCG provided very high protection against tuberculosis death among poorly nourished North American Indian children, even somewhat higher than among well-nourished British adolescents, ⁷⁷⁶ a finding that would tend to contradict this hypothesis.

Differences in prevalence of infection with environmental mycobacteria

BCG vaccination has been used not only for protection against tuberculosis, but also against leprosy, $^{815-821}$ often with more success than in the prevention of tuberculosis. 777,822,823 It is thus apparent that different mycobacterial species (in this case *M. tuberculosis*, *M. bovis* BCG, *M. microti*, and *M. leprae*) exert a modification of the immunologic response to infection with another mycobacterial species. 824 It is thus postulated that infection with one species of mycobacterium triggers a cellular immune response prepared to act more swiftly in the killing of mycobacteria of another species acquired during a subsequent infection. This is most apparent from the (limited) protection provided by infection with *M. tuberculosis* against superinfection with tubercle bacilli, 810 and the apparently similar effect of *M. bovis* BCG under certain circumstances. That BCG can also afford protection against leprosy would indicate that cross-protection is not limited to closely related mycobacterial species.

It has been postulated that different mycobacterial species induce different immunologic responses, some beneficially increasing protection against super-infection with another mycobacterial infection, while others may increase susceptibility to progression to clinically overt disease.⁸²⁵ In experimental models, protection afforded by vaccination with M. bovis BCG, M. fortuitum, M. avium, M. kansasii, and M. scrofulaceum (then called Gause strain) against *M. tuberculosis* was examined in the guinea pig.⁸²⁶ All environmental mycobacteria used in this study provided some protection, but with a wide variation, yet none provided as high a level of protection as BCG vaccination. It has therefore been postulated that the low protection afforded by BCG in Georgia as compared to the high protection observed in Britain may be attributable to a differential prevalence of infection with environmental mycobacteria.⁸²⁶ Edwards and colleagues demonstrated similar protection by vaccinating with M. avium complex against M. tuberculosis isolated in Chingleput as with the Danish BCG strain.⁸²⁷ Orme and Collins demonstrated that airborne infection with *M. avium* in mice was as effective as intravenous BCG in protection against a challenge with virulent tubercle bacilli.⁸²⁸ Brown and colleagues administered *M. vaccae* in drinking water to mice, subsequently challenged them with BCG and measured the proliferative response of spleen cells.⁸²⁹ The results showed that, depending on the timing of the exposure of the mice to M. vaccae before BCG vaccination, M. vaccae could enhance, mask or interfere with the expression of sensitization by BCG.

If environmental mycobacteria do indeed provide protection against *M. tuberculosis*, and infection with them occurs before the administration of BCG, then the effect of the latter will be at least partially masked.⁸¹³ This may explain the larger protection conferred by BCG given earlier in life than if given later as demonstrated in Chingleput.⁷⁶⁵

Furthermore, the risk of tuberculosis would be expected to be greater in initially tuberculin negative persons than in individuals with small tuberculin skin test reaction sizes (more likely attributable to infection with environmental than tubercle bacilli).

In Puerto Rico, protection from BCG was lower in rural areas, where non-specific sensitivity was higher than in urban areas, where protection from BCG was higher.⁸³⁰ However, in Chingleput, the rate of tuberculosis among persons with a reaction size of more than nine millimeters to a sensitin produced from *M. avium* complex (PPD-B) was identical to that among those with zero to nine millimeters reaction sizes.⁷⁶⁵

In the United Kingdom, the risk of tuberculosis was higher among initially tuberculin skin test negative adolescents than among those reacting to 100 tuberculin units only, but the risk decreased over time (figure 72).⁷⁷⁶ The protection afforded against tuberculosis by a tuberculin skin test reaction that can be elicited only by this large dose of tuberculin is remarkably similar (but smaller) to that imparted by BCG vaccination (figure 73).

In the Karonga, Malawi, trial the risk of tuberculosis during followup was lowest among those with an initial tuberculin skin test reaction size of six to 10 mm (figure 74).⁸³¹ After adjustment for age and sex, the risk was also lower among those with reactions of one to five millimeters than among non-reactors.⁸³²

That different species of mycobacteria seem to act on the immune system has also been demonstrated by observations from Sweden. After the cessation of mass BCG vaccination, there was a large increase in peripheral lymphadenitis due to environmental mycobacteria (figure 75) (703,833,834 and Romanus V, personal written communication, Feb 18, 2000). Similarly, in the Czech Republic, the incidence of lymphadenitis among children due to *M. avium* following cessation of BCG vaccination was 3.6, compared to 0.2 per 100,000 person-years among children vaccinated on the insistence of their parents, 835 suggesting a protection of 95% (95% confidence interval 88% to 98%) from BCG against lymphadenits due to *M. avium*.

While not all findings are consistent with the hypothesis that environmental mycobacteria may mask the protection that BCG can confer in their absence,⁸³² it may explain to a considerable extent certain variations in observed efficacy.

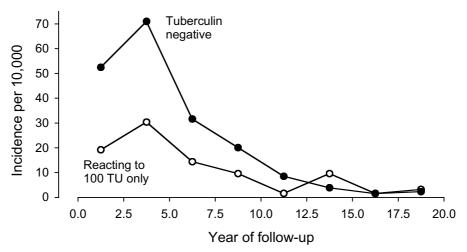


Figure 72. Risk of tuberculosis during follow-up of British school children, by initial tuberculin skin test reaction size, among placebo recipients, BCG trial, Great Britain.⁷⁷⁶

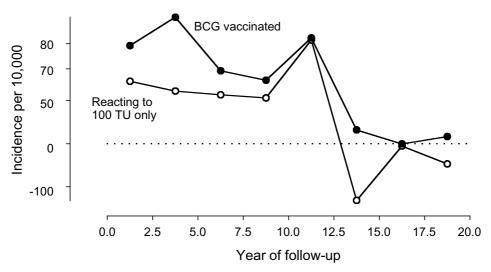


Figure 73. Comparative protection from BCG vaccination and presumed infection with environmental mycobacteria among British school children during follow-up.⁷⁷⁶

Other factors

It has been suggested that infestation with parasites, in particular with helminths, may affect the human T cell immune responses to mycobacterial antigens.⁸³⁶ Treatment of helminths resulted in significant improvement

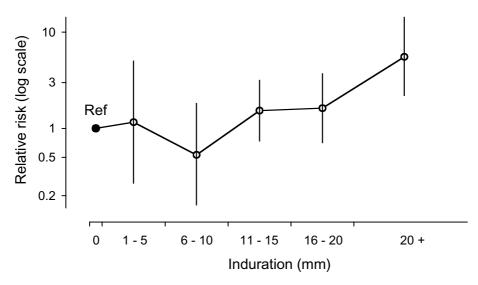


Figure 74. Risk of tuberculosis during follow-up by size of initial tuberculin skin test reaction, Karonga District, Malawi. Reproduced from⁸³¹ by the permission of the publisher Elsevier Science.

of T cell proliferation and interferon-gamma production. This could explain to some extent the reduced efficacy of BCG in countries in the world where helminthic infestation is common.⁸³⁶

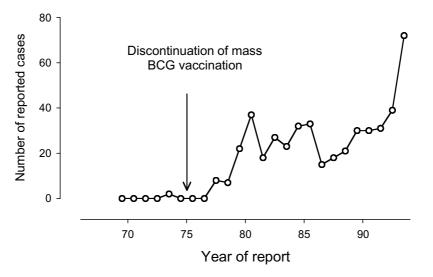


Figure 75. Reported cases of mycobacteriosis due to *M. avium* complex, Sweden, 1969-1993. Data courtesy Victoria Romanus, Swedish Institute for Infectious Diseases.

BCG re-vaccination

It is or has been the policy in many countries to re-vaccinate with BCG at school entry or later in life. There is no evidence that this increases protection against tuberculosis, ^{723,837,838} but in northern Malawi it has been shown to considerably increase protection against leprosy. ⁷⁷⁷ Re-vaccination schemes often fall into the lowest tuberculosis risk period in life (age five to 14 years) and target a population where protection from BCG vaccination is dubious or variable at best.

Effects of BCG other than those directed against tuberculosis

BCG has been shown to be protective against leprosy in some situations $^{815-818,839}$ while not in others. 819 It has also shown to be effective against *M. ulcerans*, albeit with an apparently very short-lived protection. 840

The best known indications for BCG against other than mycobacterial diseases are its use as an immunotherapeutic agent in the treatment of superficial bladder cancer⁸⁴¹⁻⁸⁵⁰ and, to a lesser extent, malignant melanoma.⁸⁵¹ It has also been suggested that BCG reduces the risk of atopy and asthma, ⁸⁵²⁻⁸⁵⁴ and reductions in the risk of intestinal nematodes in children⁸⁵⁵ and HIV-infected patients have been reported.^{856,857}

Indications and recommendations for the use of BCG vaccination

Approximately 100 million children now receive BCG every year.⁷²³ The number of doses produced in the year 2000, in descending order, were the Copenhagen 1331 strain, D2PB302, Tokyo 172, Sofia SL 222, Pasteur 1173, Glaxo 1077, and the Russian strain.⁷²³

While there have been wide variations in the protection afforded by BCG vaccination in different trials, the evidence is overwhelming that BCG provides protection against tuberculosis, especially against tuberculous meningitis and death from disseminated tuberculosis in children. Where it worked, its protective effect waned over time, to disappear after 15 to 20 years. The evidence for protection against bacteriologically confirmed tuberculosis in adults has been less consistent.

Because BCG vaccination is given early in life, the protection afforded is limited in time, and its effect on bacteriologically confirmed tuberculosis in adults is inconsistent, it cannot be expected to have a great impact on the epidemiology of tuberculosis.^{858,859}

It seems inappropriate to conclude from meta-analyses that BCG provides some average protection.^{791,792} The observed range in protection is real and remains largely unexplained.

In light of the evidence, WHO recommends its use in newborn children or as early in life as possible.^{793,860} This is still sound policy for those countries in the world where tuberculosis is highly prevalent, and tuberculous meningitis is a frequent, disabling or fatal occurrence. It fails to address the role of BCG where tuberculosis in children has become a rare occurrence.

The IUATLD has developed recommendations on criteria for the discontinuation of mass BCG vaccination.⁸⁶¹ Three key issues enter into the decision making process on the discontinuation of BCG vaccination.

The first is the extent of protection BCG actually imparts in a given location. In the USA, the low efficacy of BCG vaccination in Georgia, Georgia-Alabama, and Puerto Rico had an important impact on the decision not to routinely utilize BCG vaccination. As such prospective studies are usually beyond the realm of resource availability, effectiveness might alternatively be studied utilizing the case-control or contact study approach.

The second is the frequency of serious forms of tuberculosis in children (meningitis, disseminated forms) weighted against the frequency of adverse reactions from the vaccine itself. This has been best studied in Sweden where the frequency of serious adverse reactions from BCG vaccination (osteoarticular and disseminated mycobacteriosis due to BCG) outweighed the incidence of cases that the vaccine was intended to prevent (figure 76).⁷⁰³ Similarly, BCG vaccination may become non-cost-effective as the frequency of childhood tuberculosis decreases, so that an increasing number of children need to be vaccinated to prevent one case.

The third consideration is the value attached to the preservation of the utility of the interpretation of tuberculin skin test results. BCG vaccination induces tuberculin sensitivity and complicates the interpretation of tuberculin skin testing results. In industrialized countries with an elimination strategy in mind, the tuberculin skin test is an important means of identifying persons with tuberculous infection at a high risk of progression to tuberculosis who would benefit from preventive chemotherapy.

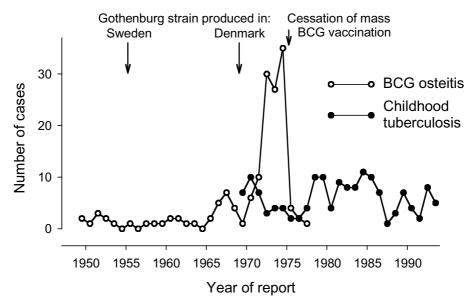


Figure 76. Osteitis due to BCG vaccination and incidence of pulmonary tuberculosis among Swedish-born children, Sweden 1949-1993.⁷⁰³

WHO discourages re-vaccination because there is no evidence of its usefulness.⁸⁶² Lack of evidence is, however, not synonymous with lack of efficacy. Re-vaccination at school entry is likely to be inefficient (even if it were efficacious), because it falls into the period in life when the risk of tuberculosis is lowest.

Finally, concerning HIV infection, the WHO has concluded after careful review of available data, that BCG vaccination schemes do not need to be altered unless HIV infection is symptomatic (AIDS).⁷²¹ This, too, seems to be a reasonable recommendation given the lack of evidence of an increased frequency of serious adverse events in BCG-vaccinated children who also have acquired HIV infection from their mother. However, it appears that HIV infection lowers the protective effect against extrapulmonary tuberculosis.⁸⁶³ In industrialized countries, where the need for BCG vaccination is generally lower, it is usually recommended not to give BCG vaccination to individuals known to have HIV infection.⁸⁶⁴

The freeze-dried vaccine should be kept refrigerated and protected from light, and diluted only immediately before vaccination. In most countries, BCG vaccine is given by the intradermal route, generally by injection with a 25 or 26 gauge needle, in the deltoid insertion region of the upper arm.⁸⁶⁵ Most manufacturers (including all those who provide vaccine for UNICEF,

the largest purchaser in the world) recommend a 0.05 mL dose for infants, and the double dose for children.

Difficulties have arisen for decision makers about the value of vaccinating health care workers at increased risk of infection with M. tuberculosis, particularly in settings where multidrug-resistant tuberculosis is common. The uncertainty stems from the scarcity of data on protection against tuberculosis among adults, and the generally low level of protection (or none at all) among adults in clinical trials. While decision analyses appear to favor the use of BCG vaccination in such settings, 866 such a conclusion has been disputed, largely based on the argument that it deprives those vaccinated from ever learning whether they have acquired tuberculous infection or not (loss of specificity of the tuberculin test).⁸⁶⁷ Nevertheless, in areas where BCG has been demonstrated to provide appreciable protection against tuberculosis among adults, where there is a high risk for health care workers of becoming infected, and where multidrug-resistant tuberculosis is common, a BCG vaccination policy for health care workers might deserve consideration. Where these conditions are not met, non-vaccination of health care workers might be more appropriate.

In summary, barring a better alternative, BCG vaccination remains a useful adjunct for the individual protection against disabling and lethal forms of childhood tuberculosis in most parts of the world where tuberculosis remains highly prevalent. It cannot be expected, however, to have great impact on the epidemiologic situation of tuberculosis.^{858,859}

4. Preventive chemotherapy

In the early 1950s Lincoln described her observations with chemotherapy and its effect on case fatality from primary tuberculosis.⁸⁶⁸ In particular, fatality from tuberculous meningitis fell from 100% to 17% with streptomycin and para-aminosalicylic acid, and to 12% with the introduction of isoniazid. None of the patients with miliary tuberculosis treated with isoniazid alone or in combination with other drugs developed tuberculous meningitis. She concluded that:

"...the use of isoniazid will have to be considered for every child with active primary tuberculosis and probably also for children with known recent conversion of tuberculin tests even if chest roentgenograms are normal. The duration of therapy would most likely be for a year in order to cover the period during which meningitis would be most likely to develop..." ⁸⁶⁸

While credit for the idea of preventive chemotherapy might be shared by various investigators, ⁶⁴¹ this is almost certainly one of the earliest accounts to spell out so clearly the research agenda on which the US Public Health Service, and subsequently other bodies, would engage.

It is noteworthy that two issues are raised here. The first is the prevention of complications from clinically manifest tuberculosis; the second is the notion of prevention of disease from recently acquired asymptomatic infection. In this monograph the term preventive chemotherapy is defined as treatment of latent, asymptomatic tuberculous infection with the intent to reduce the risk of progression to clinically manifest disease, and not what is essentially chemotherapy of active tuberculosis. Nevertheless, the first US Public Health Service controlled trial dealt precisely with that, and provided evidence of a 70% protection with isoniazid monotherapy against development of complications from primary tuberculosis. ^{641,869,870} Similarly, a controlled trial conducted in India among patients with minimal radiographic lesions, not certain to be active or inactive, offered evidence for a 68% protection against bacteriologically confirmed pulmonary tuberculosis.⁸⁷¹ This type of investigation will not be dealt with further at this point.

Numerous clinical trials of preventive chemotherapy (in the stricter sense of the definition used here) have been conducted. They encompassed a range of variables, including persons at varying risk of tuberculosis, choice of anti-tuberculosis agent, and duration of therapy. While no attempt is made here to provide a comprehensive list of all of the trials, the best known have been selected to review the efficacy that preventive chemotherapy has offered in randomized controlled clinical trials in various settings.

In order to provide comparable results, Wald 95% confidence intervals⁷³¹ were calculated for all trial efficacy estimates, unless (in some recent publications) the authors provided adjusted risk ratios or provided insufficient data to recalculate confidence intervals. In this case, the confidence intervals provided by the authors were used.

Prevention of disease in tuberculin skin test reactors

Persons who react to tuberculin, but whose acquisition of tuberculous infection lies in the remote past, have a relatively small annual risk of progression to clinically active tuberculosis compared with persons who have recently acquired infection.¹ While the exact point of acquisition of infection is rarely known for an individual, several studies have been conducted among patients whose tuberculous infection has been unlikely, to have been of recent origin on average (wide spread of acquisition points in time). The first point of interest is the efficacy of preventive chemotherapy in providing protection against progression to tuberculosis during, and only during, the length of treatment. Three of the four trials for which such information is available used isoniazid for twelve months^{641,872-874}, and in one it was given for nine months.⁸⁷⁵ The results obtained by the end of the treatment period are summarized in figure 77.

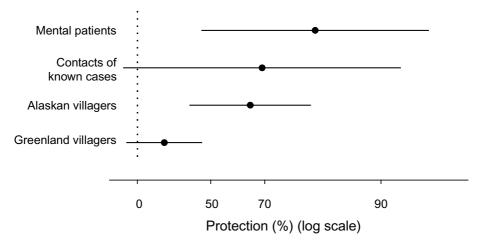


Figure 77. Protection from isoniazid preventive therapy against tuberculosis among tuberculin skin test reactors during the year of treatment.^{641,872-875}

Tuberculosis was much more frequent among mentally ill patients in institutions than in the general population in the United States, and it was natural to consider this group for preventive chemotherapy to reduce the risk of endogenous reactivation disease.⁸⁷² People of all ages were included, but older patients made up the bulk of participants, with an average age of around 50 years. After exclusion of those participants known to have had a negative tuberculin skin test at intake, the protection afforded by isoniazid during the treatment year was 81%.

Persons in contact with notified tuberculosis patients for various lengths of time were included in another US Public Health Service clinical trial.⁸⁷⁴ Some of the index cases had long since been cured of active disease, while others were still on treatment. Contacts who had already developed tuberculosis at the time of enrolment were excluded from the trial. Over half of the enrolled were initially tuberculin negative and stratification by initial tuberculin skin test result was not provided. The risk of tuberculosis during the treatment year was very low and the confidence interval around the observed point estimate of protection of 69% was thus very wide. However, eight of the nine observed cases in the placebo group had occurred among those with an initially positive tuberculin skin test.

In Alaska, a community preventive chemotherapy trial was started when the annual infection rate was almost 100 times greater than in the continental United States.^{641,873} In the Bethel Hospital service area where the trial was conducted, the annual risk of infection was 25%, a rate far exceeding any reported risk elsewhere in the world.⁸⁷⁶ By the time of starting the trial the risk of infection had already substantially decreased. Because of the adverse climatic and transport conditions it was not feasible to test all persons with tuberculin. Approximately one third of those tested had a tuberculin skin test diameter of less than five millimeters. During the treatment year the protection from isoniazid was 66%. Protection was demonstrated at all levels of adherence (amount of isoniazid taken), with an indication that six months of isoniazid might have sufficed.⁸⁷⁶

In Greenland it had been realized, by the mid-1950s, that the majority of tuberculosis cases developed in the years immediately after primary infection. A trial was thus undertaken to study the efficacy of isoniazid preventive chemotherapy at the community level.^{875,877} Children below the age of 15 years were excluded from the trial. Placebo or isoniazid were given weekly for a total duration of nine months. Half of the participants received all dosages and over 80% received at least three quarters of the intended dosages. The crude protection afforded by isoniazid during the year following commencement of treatment was 22%. For reasons that are not understood, there was no protection observed in those aged less than 25 years, in contrast to 56% protection in those aged 25 to 34 years, and intermediate protection in the other age groups.

Prevention of disease in persons with risk factors

The risk of tuberculosis among persons with long-standing tuberculous infection (those studied in the above presented trials) is fairly small, thus the effectiveness of a preventive chemotherapy scheme is relatively modest. Conversely, it is of special interest to study the efficacy of isoniazid preventive chemotherapy in persons with a recognized increased risk of tuberculosis, because the attributable fraction of cases that can be prevented is relatively large if the prevalence of the risk factor is also high.

Recently acquired infection

Recently acquired tuberculous infection is not only associated with an increased risk of progression to tuberculosis, but it is also a frequent event. Four studies of preventive chemotherapy among contacts of newly diagnosed index cases of tuberculosis are selected here to illustrate the point (figure 78). ⁸⁷⁸⁻⁸⁸¹

In April 1960, a patient with pulmonary tuberculosis in a marine camp of the Royal Netherlands Navy infected a large number of his mates in a

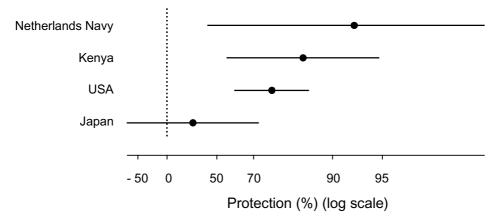


Figure 78. Protection from isoniazid preventive therapy against tuberculosis among contacts of tuberculosis patients.⁸⁷⁸⁻⁸⁸¹

barracks.⁸⁷⁸ On entering service in January, 59 of the men sharing the barracks had a negative tuberculin skin test, while by the beginning of April, 56 of them had become converters. In the entire camp 305 conversions were registered among 1,105 initially negative men. A double-blind controlled trial was carried out among the 261 converters who were not excluded due to departure or because they had already contracted tuberculosis at the time of starting the trial. After one year, nine cases had developed tuberculosis in the placebo group compared to only one in the isoniazid group. During follow-up for a total observation period of four years, three additional cases developed, all in the placebo group, indicating an overall protection of 93%.

In Nairobi, Kenya, contacts of newly diagnosed tuberculosis cases were randomly assigned to receive isoniazid or placebo for one year.⁸⁸⁰ During the treatment year and the two-year follow-up period, preventive chemotherapy provided 85% protection against culture-confirmed pulmonary tuberculosis.

A large trial was conducted by the US Public Health Service among contacts of newly identified cases of tuberculosis.⁸⁷⁹ Contacts found to have tuberculosis during the initial examination were excluded from the trial. Over 25,000 contacts were eligible for enrolment. Of these, 48% had tuberculin skin test reaction sizes of five or more millimeters of induration. Approximately two thirds of contacts were less than 20 years old. About two thirds of study participants were estimated to have taken all of their medications, and about 80% three quarters or more. Among those who could be reexamined at the end of the 12-month period of medication, isoniazid gave 77% protection against tuberculosis.

In Japan, contacts of new cases of tuberculosis were randomly assigned to receive placebo or isoniazid.⁸⁸¹ The protection afforded was only 30%, with confidence intervals including zero.

Infection with the human immunodeficiency virus

Infection with HIV is the strongest yet identified risk factor for progression from tuberculous infection to tuberculosis. Because HIV alters the biological response to *M. tuberculosis* so fundamentally, it could not be taken for granted that isoniazid preventive chemotherapy would work as well as in immunocompetent patients.

Tuberculosis is accompanied by an increase in tumor necrosis factor alpha (TNF- α). TNF- α also increases *in vitro* replication of HIV.⁸⁸² It

might be expected, therefore, that prevention of development of tuberculosis would also delay onset of AIDS among HIV infected patients (figure 79). There is, however, as yet little epidemiological evidence that this is the case.⁸⁸³

A series of controlled clinical trials has been undertaken in various settings to evaluate the efficacy of isoniazid (and other compounds) compared to placebo in protecting HIV-infected individuals against tuberculosis (figure 80).

The first study of this kind was conducted in Port-au-Prince, Haiti.⁸⁸⁴ The efficacy of 12 months of isoniazid compared to placebo was ascertained. The protection among persons with five or more millimeters of induration to a tuberculin skin test was 83%. The population was small, however, and the confidence intervals were consequently wide. An additional finding was a significant delay in onset of HIV disease in the isoniazid group compared to those receiving placebo. Survival analysis also demonstrated significant protection against AIDS-defining illnesses and AIDS-attributable death among tuberculin-positive, but not among tuberculin-negative, patients.

In Lusaka, Zambia, HIV-positive individuals were randomly assigned to receive twice-weekly isoniazid for six months or placebo and a third arm

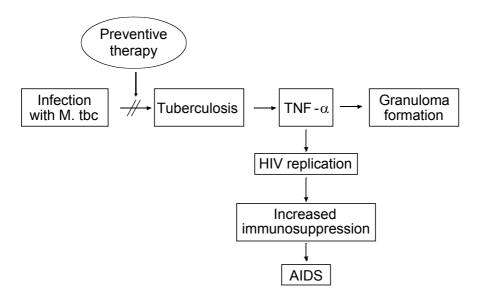


Figure 79. Schematic presentation of the impact of tuberculosis on TNF- α production and HIV replication, and prevention of the chain of events with preventive therapy.

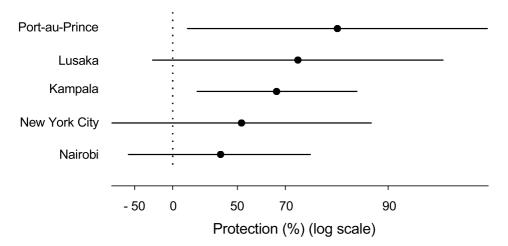


Figure 80. Protection from isoniazid preventive therapy against tuberculosis among HIV-infected persons.⁸⁸⁴⁻⁸⁸⁸

with a rifampicin plus pyrazinamide containing regimen.⁸⁸⁵ They were followed up for a median of 1.8 years. The main outcome measures were incidence of tuberculosis and death. Among those with a tuberculin skin test reaction of five or more millimeters of induration, the point estimate of protection from isoniazid was 74%, yet because of the small number, the confidence intervals were wide and included zero. There was no difference in mortality between preventive chemotherapy and placebo groups. An important observation was that the effect of preventive chemotherapy declined following cessation of treatment so that by 18 months after the completion of therapy incidence rates in treated and non-treated groups were the same.

In Kampala, Uganda, HIV-infected patients were enrolled in a randomized trial to receive one of four arms: placebo, isoniazid for six months, or two rifampicin-containing regimens (one with, the other without pyrazinamide). Among patients with a tuberculin skin test reaction of five or more millimeters of induration, isoniazid reduced the risk of tuberculosis over a mean follow-up time of 15 months by 67%.⁸⁸⁶ Survival did not differ between the groups.

Collaborating centers in New York City and elsewhere conducted a randomized trial to assess the efficacy of isoniazid in patients with HIV infection who were anergic.⁸⁸⁷ Anergy was defined as reacting with more than five millimeters of induration to tuberculin and less than two millimeters to both mumps antigen and tetanus toxoid. Patients were addi-

tionally considered to belong to groups at risk of tuberculous infection. Only nine cases of tuberculosis occurred in the entire cohort of more than 500 patients during a follow-up period of 30 months following cessation of treatment with six months of either placebo or isoniazid: three in the isoniazid group and six in the placebo group. This corresponds to an over-all protection of 52%, yet with 95% cent confidence intervals including zero.

In Nairobi, Kenya, HIV-positive patients were randomly assigned to receive either daily isoniazid for six months or placebo (irrespective of the tuberculin skin test result).⁸⁸⁸ Outcome measures were incidence of tuberculosis and death. The follow-up period from enrolment onwards was a median of 1.8 years. The protection among persons with positive tuberculin skin test reactions (not further defined) was 40%, yet with confidence intervals overlapping zero. There was a slight, statistically significant reduction in risk of death among tuberculin-positive isoniazid recipients compared to the controls.

Spontaneously healed tuberculosis with fibrotic residuals

Patients with tuberculosis that has healed spontaneously with fibrotic lesions are frequently found, and remain an important source of reactivation tuberculosis, particularly in countries where the tuberculosis risk has been rapidly declining and most cases are the result of endogenous reactivation. Three such studies are shown here (figure 81).

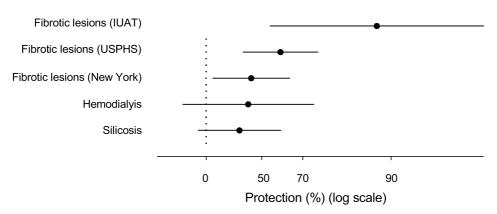


Figure 81. Protection from isoniazid preventive therapy against tuberculosis among patients with fibrotic lesions, hemodialysis, or silicosis.^{123,641,891-893}

A large trial was conducted in Europe by the International Union Against Tuberculosis Committee on Prophylaxis among patients with fibrotic lesions.^{123,889} Patients were randomly assigned to four groups, each consisting of close to 7,000 patients. A control group received placebo, and three groups received three, six or twelve months, respectively, of isoniazid. They were followed up to five years following intake. Among persons completing twelve months of, and adhering to, the prescribed course of chemotherapy the protection afforded by isoniazid was 93%. The effect was greater among those with larger than among those with smaller radiographic lesions.

Similarly, the US Public Health Service conducted a trial among patients with inactive lesions and followed them up for five years following enrolment into a randomized trial of twelve months isoniazid versus placebo.⁶⁴¹ The protection afforded by isoniazid was 60%.

In New York City, another study with two years of isoniazid was conducted among patients with inactive lesions.^{890,891} The number of patients enrolled was small, and the protection afforded was 43% over a period of six years from enrolment.

Silicosis

Silicosis is a well-recognized risk factor for tuberculosis and is highly prevalent in countries where mining industries and other environments (granite quarry workers) offer poor protection against silica dust inhalation. In a study jointly organized by the Hong Kong Chest Service, the Tuberculosis Research Centre, Madras, and the British Medical Research Council, patients in Hong Kong were enrolled into a double-blind randomized trial with six months of isoniazid (and two rifampicin-containing arms) compared to a placebo group.⁸⁹² During the five-year follow-up period, isoniazid offered a protection of 34%, but the 95% interval included zero (figure 81).

Renal failure

A relatively small study with 184 patients on renal dialysis or after renal transplant were randomly assigned to receive either one year isoniazid or placebo. They were followed up for one year following cessation of therapy. Among those who completed therapy, the protection afforded by isoniazid was 41%, but the confidence interval overlapped zero (figure 81).⁸⁹³

Prevention of disease following cessation of preventive chemotherapy

One consideration has been the duration of efficacy of isoniazid preventive chemotherapy. In the three studies shown here, the protection remained unaltered over four to five years following cessation of preventive chemotherapy (figure 82).^{123,875,876} Similar maintenance of efficacy over even longer periods was shown in other studies.⁶⁴¹ In the longest follow-up reported, from the Bethel, Alaska area, protection persisted for more than 19 years.⁸⁹⁴

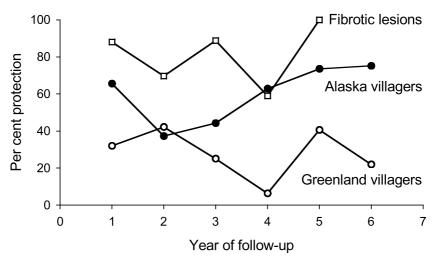


Figure 82. Long-term efficacy of preventive chemotherapy with isoniazid.123,641,876

Nevertheless, in areas where the risk of infection is high and a large proportion of cases is emanating from recently infected persons, protection might be expected to decline over time. However, once the tubercle bacilli have been eliminated from the body, one might expect some protection to be afforded against super-infection leading to disease, similar to that expected from BCG vaccination.

Prevention of disease with different durations of treatment

The duration necessary to provide optimum protection from isoniazid has not been satisfactorily determined. In fact, the only study seeking direct evidence was the trial of the International Union Against Tuberculosis Committee on Prophylaxis (figure 83).¹²³ Among "completer-compliers" the answer is quite clear-cut. Most benefit was obtained with twelve months chemotherapy with 93% protection, while six months offered 69% protection, and three months 32%.

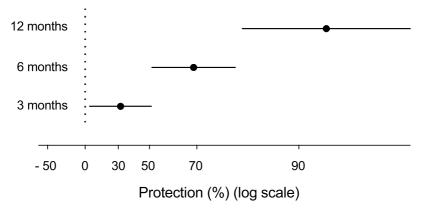


Figure 83. Impact of duration of intake of isoniazid preventive therapy on protective efficacy.¹²³

However, if all patients were analyzed (and not only "completer-compliers"), the differences between six months and twelve months became much smaller, as adherence dropped with increasing length of treatment. For this reason and considering the cumulative risk of adverse drug events and personnel costs, it has been suggested that six months of preventive chemotherapy with isoniazid was more cost-effective than twelve months.⁸⁹⁵

However, the primary decision that has to be taken in the selection of a regimen (curative or preventive) is efficacy; the second is effectiveness.

In consideration of these findings, recommendations have been made for preventive chemotherapy to be given for six to twelve months, with every effort made to ensure adherence for six months.³⁵⁶ The isoniazid preventive chemotherapy trials in the United States showed that the optimum duration might lie somewhere around nine months (figure 84).⁸⁹⁶ The American Thoracic Society and the US Centers for Disease Control now recommend nine months of isoniazid treatment.⁸⁹⁷ The British Thoracic Society recommends six to twelve months for preventive chemotherapy, the longer duration recommended for HIV-positive patients.^{898,899}

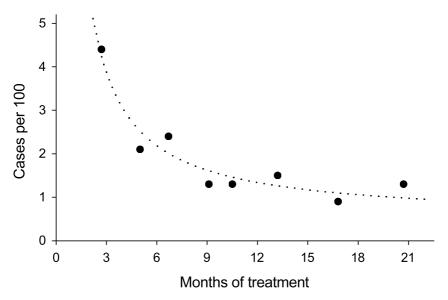


Figure 84. Tuberculosis risk and duration of intake of isoniazid preventive therapy in the Bethel, Alaska, preventive chemotherapy studies. Reproduced from⁸⁹⁶ by the permission of the publisher International Union Against Tuberculosis and Lung Disease.

Prevention of disease with alternatives to isoniazid

Rifampicin has been remarkably effective in shortening the required duration of chemotherapy of tuberculosis.^{122,504} It is postulated to act particularly well on mycobacterial sub-populations with only short bursts of metabolic activity.⁴⁵⁶ Such a situation probably exists in the case of latent tuberculous infection and it is thus appealing to hypothesize that rifampicin might be effective in preventive chemotherapy and may also reduce the duration of the required treatment period compared to isoniazid.

In a mouse model, Lecoeur and collaborators tested the efficacy of rifampicin with or without other drugs in combination as a preventive chemotherapy tool in comparison with isoniazid.⁹⁰⁰

Latent, sub-clinical infection was produced by vaccination with BCG and subsequent challenge with *M. tuberculosis*. After an initial increase in viable tubercle bacilli, this produced a stable count of bacilli in the spleen, indicating that the relatively limited population was no longer actively multiplying in the spleen when drug treatment was given. In a first experi-

ment, mice were assigned to five groups: 1) no treatment, 2) isoniazid for six months, 3) rifampicin for two months, 4) rifampicin plus isoniazid for two months, and 5) rifampicin plus isoniazid plus pyrazinamide for two months (figure 85). From this experiment, it was shown that two months of rifampicin-containing preventive chemotherapy was as effective as six months of isoniazid.⁹⁰⁰

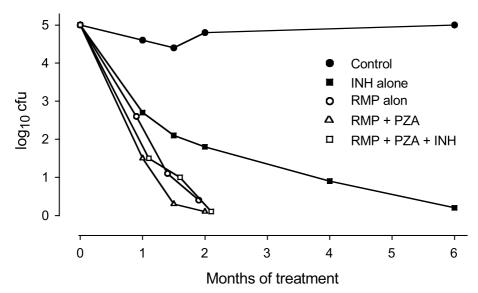


Figure 85. Mouse model of latent tuberculous infection and efficacy of various durations and combinations of preventive therapy on spleen bacillary count. Reproduced from⁹⁰⁰ by the permission of the publisher American Thoracic Society at the American Lung Association.

In a second experiment, in comparison to isoniazid the relative efficacy of various combinations with rifampicin of different durations was evaluated. Mice received 1) six months of isoniazid, 2) three months of rifampicin plus isoniazid plus pyrazinamide, 3) three months of rifampicin, or 4) two months of rifampicin plus pyrazinamide. The experiment was calibrated in such a way as to ensure that viable bacilli remained at cessation of therapy to allow their culture from spleen at cessation and after a follow-up of a six-month period without treatment. All rifampicin combinations proved superior to isoniazid treatment for six months (figure 86).⁹⁰⁰ The best combination was rifampicin plus pyrazinamide (without isoniazid). Rifampicin alone for three months was also very effective.

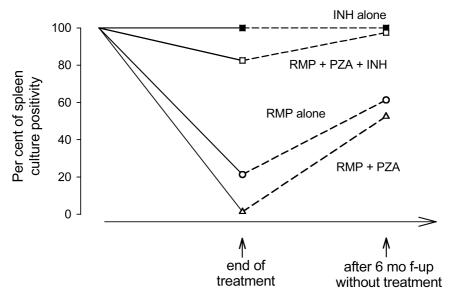


Figure 86. Mouse model of latent tuberculous infection and efficacy of various rifampicin combinations of preventive therapy on spleen bacillary count. Reproduced from⁹⁰⁰ by the permission of the publisher American Thoracic Society at the American Lung Association.

Two types of studies are available to test the hypothesis in human subjects that rifampicin with or without isoniazid is first, efficacious, and second, equivalent or better than isoniazid alone, even if given for shorter durations. The first type consists of comparisons of rifampicin and rifampicin combinations with placebo, the second comparisons of rifampicin and rifampicin combinations with isoniazid (equivalence studies). The hypothesis and sample size requirements differ in the two approaches.

Rifampicin and rifampicin combinations in comparison to placebo

Studies comparing rifampicin (and combinations) with placebo have been carried out among patients with silicosis and patients with HIV infection (figure 87). ^{885,886,892}

In Kampala, Uganda, two arms had rifampicin-containing regimens. Compared to placebo, daily rifampicin plus isoniazid for three months gave 60% protection among tuberculin-positive patients with HIV infection. Rifampicin plus isoniazid plus pyrazinamide given daily for three months offered 49% protection.⁸⁸⁶

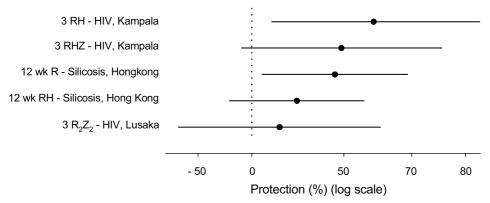


Figure 87. Protection against tuberculosis with rifampicin containing preventive therapy among persons with HIV infection or silicosis.^{885,886,892}

In the study on patients with silicosis in Hong Kong presented above, in addition to the isoniazid (for six months) and placebo arms, two additional arms contained rifampicin. One of these consisted of twelve weeks of rifampicin alone and the second of twelve weeks of rifampicin plus isoniazid.⁸⁹² All drugs were given daily. Rifampicin alone for twelve weeks gave 46% protection. Rifampicin plus isoniazid for the same duration offered 29% protection, with the confidence interval including zero.

In Lusaka, Zambia, a twice-weekly regimen of rifampicin plus pyrazinamide given for three months gave 19% protection (the confidence intervals overlapping zero) against confirmed tuberculosis in HIV-infected patients.⁸⁸⁵

Rifampicin and rifampicin combinations in comparison to isoniazid

A few studies have provided information on the equivalence of rifampicincontaining preventive chemotherapy with isoniazid preventive chemotherapy (figure 88). ^{885,892,901,902} Again, these studies were carried out among patients with risk factors (silicosis and HIV infection).

In Hong Kong, twelve weeks of rifampicin provided 44% protection compared to six months of isoniazid, a statistically significant superiority, while the 25% comparative effect of twelve weeks with rifampicin plus isoniazid was not statistically different from the protection offered by isoniazid.⁸⁹² Thus, the overall protection against tuberculosis with preventive chemotherapy among silicosis patients was relatively poor, and rifampicin alone appeared to be superior to rifampicin plus isoniazid.

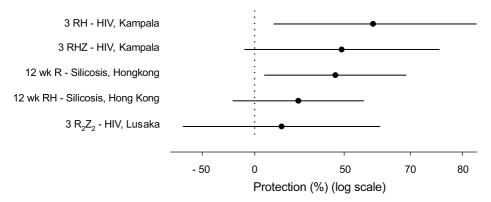


Figure 88. Protection against tuberculosis with rifampicin containing preventive therapy compared to isoniazid preventive therapy (equivalence studies) among persons with HIV infection or silicosis.^{885,892,901,902}

In Cité Soleil and Petit Place Cazeau, Haiti, patients with HIV infection and a tuberculin skin test induration of five or more millimeters were randomly assigned to receive either isoniazid for 24 weeks or rifampicin plus pyrazinamide for eight weeks.⁹⁰¹ All drugs were given twice weekly, the first weekly dose directly observed, the second self-administered. The overall protection afforded by the rifampicin-containing regimen was minus 30%, with confidence intervals overlapping zero. During the first ten months after entry, the risk among isoniazid recipients was significantly lower than among rifampicin recipients.

Similarly, in Lusaka, Zambia, isoniazid for six months gave better protection than rifampicin plus pyrazinamide for three months, but the confidence intervals were wide, the difference was not statistically significant, and the protective effect from both arms was lost after two to three years.⁸⁸⁵ The long-term evaluation showed that protection lasted for about two and a half years and none of the regimens appeared to have an effect on HIV progression or mortality.⁹⁰³

In a multi-center study involving 53 treatment units in Brazil, Haiti, Mexico, and the United States, a total of 1,583 HIV-infected patients were randomized to receive either isoniazid for twelve months (control arm) or rifampicin plus pyrazinamide for two months (experimental arm).⁹⁰² Among the inclusion criteria were the presence of a tuberculin skin test reaction of five or more millimeters of induration. For bacteriologically confirmed cases, the relative protection of the two-month regimen was 33% for bacteriologically confirmed, and five per cent for confirmed and probable cases. The 95% confidence interval was reasonably narrow, overlapped zero, and

thus suggested equivalence (the hypothesis of the study) between the two regimens. Completion of therapy was superior in the experimental compared to the control arm.

In the United States, preventive chemotherapy regimens using rifampicin (plus pyrazinamide) of two to four months' duration have been recommended.⁸⁹⁷ However, recent reports on fatal and severe hepatitis associated with preventive therapy using rifampicin plus pyrazinamide⁹⁰⁴ have led to a change of the recommendation and advising great caution in the use of this combination.⁹⁰⁴

Effectiveness of preventive chemotherapy

There can be little doubt about the efficacy of preventive chemotherapy, at least with isoniazid if given for twelve months to persons with tuberculous infection without additional risk factors. There are indications that a regimen of nine months' duration might still be similarly efficacious in reducing the risk of tuberculosis. The efficacy of isoniazid in patients with risk factors is much less well established, and many studies dealing with HIV-infected patients suffer from inadequate sample sizes. It also seems that rifampicin-containing regimens of shorter duration can afford similar protection, but the optimal duration and the role of companion drugs have not been sufficiently well established. A short-coming of most preventive chemotherapy trials has been the self-administration of medications, thus portraying more the effectiveness than the potential efficacy of the regimen in question.

All studies that have evaluated that component have clearly demonstrated the adverse effect of non-adherence on the regimen's efficacy, as would be expected. This has been the case even in the setting of clinical trials where adherence might be better than under daily operations within the context of a national program.

Several studies have also demonstrated that the type of patients who are selected for preventive chemotherapy is important, and that large numbers may have to be treated to prevent a single case if the persons selected have a low risk of tuberculosis.

In a simplified form, operational effectiveness can thus be summarized as the product of tuberculosis risk given the presence of tuberculous infection, the efficacy of the regimen, and adherence to the prescribed medications. In a few examples, table 11 summarizes different situations and the **Table 11.** Preventive therapy – effectiveness. Effectiveness of preventive therapy in dependence of risk of tuberculosis, efficacy of treatment regimen, and adherence to the regimen. All parameters are shown as fractions. Risk of tuberculosis is allowed to vary from 0.05 (estimated cumulative risk subsequent to the first five years following infection) to 0.30 (estimated cumulative risk of a person dually infected with *M. tuberculosis* and HIV).

Risk of tuberculosis	Efficacy of regimen	Adherence to treatment	Overall effectiveness	Number to treat to prevent 1 case
0.05	0.60	0.30	0.009	111
0.10	0.60	0.30	0.018	56
0.30	0.60	0.30	0.054	19
0.30	0.90	0.30	0.081	12
0.30	0.90	0.50	0.135	7
0.30	0.90	0.80	0.216	5

impact on overall operational effectiveness. The risks of tuberculosis shown here are for persons with long-standing tuberculous infection, recently acquired tuberculous infection and concomitant HIV infection (0.05, 0.10, and 0.30 for the respective risks of tuberculosis). Efficacy examples have been taken from isoniazid-preventive chemotherapy ranges, and adherence has been made up to vary as might be expected among different patients with a condition that is not symptomatic. The overall effectiveness is the product of these three variables and the number of patients that must be treated to prevent one case is the reciprocal value of effectiveness. The example shows that effectiveness will greatly vary depending on the selection of patients, the type of regimen and the extent to which patients adhere to treatment. Although reality is not quite as straightforward as in this example (it assumes that each component proportionally reduces effectiveness), it may help in deciding under which circumstances preventive chemotherapy is to be recommended. The specific indication will depend on the availability of resources, as the overall effectiveness is, under any circumstance, relatively modest. Not accounted for in this model is the probability of tuberculous infection actually being present when a "positive" tuberculin skin test is recorded.

A study in Kampala, Uganda, ascertained the operational feasibility and effectiveness of preventive chemotherapy in a high-risk population, apparently motivated to attend voluntary testing sites for HIV.⁹⁰⁵ Among patients who were found to be HIV-positive, only about 60% returned to obtain their result and to receive counseling (figure 89) and of these only

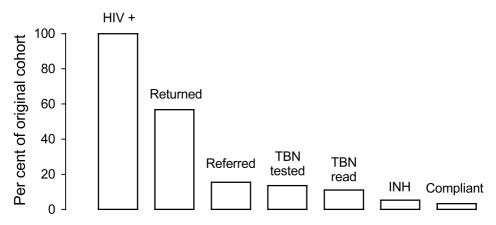


Figure 89. Operational feasibility of preventive therapy usage in a voluntary counseling and testing site for HIV infection.⁹⁰⁵

a small fraction was actually referred for evaluation of eligibility for preventive chemotherapy. Quite obviously, the collaboration was poor despite the study setting. Additional patients were lost for tuberculin testing and reading, only a fraction of these were actually eligible for preventive chemotherapy, and not all of those who were eligible were actually adherent. Only three per cent of the initial cohort completed preventive chemotherapy, and the efficacy was never assessed.

Indications and recommendations for the use of preventive chemotherapy

It is apparent that the place preventive chemotherapy will have within the context of a national tuberculosis control program will depend foremost on the epidemiologic situation and on the availability of resources. Rapid improvement in the epidemiologic situation and sufficient resources often go together, while the reverse is also the case.⁹⁰⁶

In industrialized countries embarking on strategies to eliminate tuberculosis, preventive chemotherapy will play an important role, yet the situation is rather different in countries with a high or even increasing tuberculosis burden where resources are very tight even to secure the treatment of all known cases of bacteriologically confirmed tuberculosis.

Within the context of resource availability, it should be considered that the costs for isoniazid are probably of least concern in most settings. Logistical problems may, however, impose substantial impediments in some settings in low-income countries. Of critical importance is the capability to exclude the presence of active tuberculosis. This is particularly the case in adults, where the bacterial load of unrecognized tuberculosis might be sufficiently high to favor selection of isoniazid-resistant mutants if monotherapy is being given. In the Uganda study,⁹⁰⁵ a sizeable portion of HIV-infected patients who were examined had active pulmonary tuberculosis, and not all had positive sputum smears on direct microscopic examination. For good reasons, WHO has thus recommended that both sputum smear microscopy and chest radiography are mandatory in HIV-infected patients before commencing preventive chemotherapy.⁹⁰⁷

The IUATLD limits the recommendations for preventive chemotherapy in low-income countries to asymptomatic children under the age of five years who live in the same household as a newly discovered sputum smearpositive case.⁸ This is a group of persons with a high risk of becoming infected because of the closeness of contact. Preventive chemotherapy (or prophylactic treatment in the portion of children who have escaped infection) can be administered without prior investigation except for a clinical assessment of health. Even in the presence of an asymptomatic primary complex, the bacillary load will be too small in such children to pose the problem of selecting isoniazid-resistant bacilli. The drug of choice is isoniazid (5 mg/kg body weight), as it is the least expensive and the drug with which there is most experience. The duration of treatment might be pragmatically adjusted to the length of the tuberculosis treatment regimen prescribed for the index person, i.e., between six and twelve months.

For national programs wishing to expand their preventive chemotherapy program to other risk groups, the above measures to exclude active tuberculosis before initiating preventive chemotherapy should be strictly enforced.

Appendix 1 Adjunctive treatment

Adjunctive therapy with corticosteroids

The role of corticosteroids in the treatment of tuberculosis is not precisely known and the opinion concerning their use in different clinical situations is often somewhat controversial. The available evidence for and against their use is reviewed here, following the extensive review by Dooley *et al.*, ⁹⁰⁸ supplemented by additional reports.

Pulmonary tuberculosis

The value of corticosteroids in the treatment of pulmonary tuberculosis has been evaluated in several controlled trials.⁹⁰⁹⁻⁹²⁰

Sputum conversion was not affected by corticosteroid therapy in any of these studies; early sputum conversion was faster in the control group in one study and faster in the corticosteroid group in another.

On the other hand, clinical and radiologic improvement was generally more rapid in the corticosteroid treated group, particularly among the more seriously ill patients. In the United States Public Health Service trial, prednisolone produced more frequent and more rapid radiologic clearing of the infiltrate in black patients, but was of no benefit in white patients.⁹¹⁸

In the five-year follow up of the United States Veterans Administration study, patients treated with corticosteroids were less likely to have died due to relapse of their tuberculosis, or due to respiratory illnesses such as bronchitis, respiratory insufficiency, or pneumonia.⁹¹⁷

A controlled clinical trial from India⁹²⁰ is significant in two ways. First, it is the only study using corticosteroids as adjunct therapy together with rifampicin-containing regimens. Second, it revealed that patients treated with corticosteroids who had strains initially resistant to isoniazid and streptomycin had a poorer bacteriologic response than those not treated with steroids. The deleterious effect of steroids in patients with sub-optimal chemotherapy had been observed earlier.⁹²¹ This had previously been shown in animal models as well,^{922,923} and is not unexpected. In an uncontrolled study in Zambia, HIV-infected patients treated for tuberculosis who had adjunct therapy with corticosteroids developed herpes zoster and Kaposi's sarcoma significantly more often, while generalized lymphadenopathy improved.⁹²⁴

The routine use of corticosteroids as adjunct therapy for pulmonary tuberculosis cannot, therefore, be recommended. In addition to the multitude of known adverse effects directly related to steroid use, in tropical countries parasitic infestation is common and corticosteroids in such patients may precipitate dissemination ⁹²⁵ and abscesses. ⁹²⁶

The indication for the use of corticosteroids in pulmonary tuberculosis should be restricted to patients so seriously ill that their prognosis is judged to be very poor and thus the steroids are potentially life-saving.^{7,927}

Extrapulmonary tuberculosis

Tuberculosis of serous membranes

Pleural tuberculosis

A number of studies have been reported evaluating the use of corticosteroids in pleural tuberculosis.⁹²⁸⁻⁹³⁷ Not all of the studies were conducted with the same rigor and only eight provide sufficient information for an adequate evaluation.^{928-933,936,937}

Most studies showed a more rapid resolution of effusions in those given corticosteroids. In the studies that evaluated residual pleural thickening as the crucial endpoint, three found less thickening in the steroid treated group ⁹³⁰⁻⁹³² and two found no difference between placebo and steroid treated groups. ^{936,937}

From these studies it would appear that the value of corticosteroids in the treatment of pleural effusion is doubtful. Weighted against the possible adverse effects, steroids should probably not be routinely used in tuberculous pleural effusion.

Pericardial tuberculosis

The efficacy of corticosteroid therapy for tuberculous pericarditis may differ for the different physiological stages of the disease (effusive, effusiveconstrictive, and constrictive).⁹⁰⁸ Several retrospective studies fail to address these points and lack well-defined endpoints.^{938,939} In a retrospective study addressing the critical issue of stage of disease, patients on corticosteroids had a more rapid decrease in pericardial effusion as compared to those not given corticosteroids. $^{\rm 940}$

Prospective studies evaluating the use of corticosteroids in the treatment of tuberculous pericarditis were conducted in the Transkei area of South Africa.⁹⁴¹⁻⁹⁴⁴ These studies showed that fewer patients with acute effusive pericarditis on corticosteroids required repeated drainage,⁹⁴¹ and patients with effusive-constrictive pericarditis had faster improvement.⁹⁴² In neither study were there differences in constriction, but among patients with acute effusive pericarditis, corticosteroid recipients had a lower risk of death. Among HIV-infected patients in Harare, Zimbabwe, corticosteroids significantly reduced case fatality.⁹⁴⁵

From these studies it would seem that adjuvant treatment with corticosteroids is indicated in patients with pericardial tuberculosis.

Peritoneal tuberculosis

Because of the similarity in the pathogenesis of peritoneal and pericardial tuberculosis, both involving serous membranes, a beneficial effect of corticosteroid use in peritoneal tuberculosis similar to that in pericardial tuberculosis might be expected.⁹⁴⁶ Alternatively one may argue that tuberculous peritonitis is more similar to tuberculous pleurisy. In a retrospective study from Saudi Arabia, the frequency of recurrent abdominal pain and emergency department visits were used as endpoints to compare the usefulness of corticosteroid adjunctive therapy.⁹⁴⁷ Corticosteroids appeared to have an effect in reducing the frequency of these events. However, as the study was a retrospective evaluation of a case series comparing patients who were given corticosteroid therapy with patients who had not received corticosteroids, it is not possible to draw firm conclusions. A prospective trial, allocating patients to a three-month course with either prednisone or placebo, was carried out with death and intestinal obstruction evaluated as endpoints.⁹⁴⁸ Corticosteroid-treated patients fared better, but the small sample size did not detect a significant difference in the frequency of these events.

The evidence for the usefulness of corticosteroid treatment as adjunctive treatment in peritoneal tuberculosis is sufficiently convincing to recommend its routine use.

Meningeal tuberculosis

Adjunctive treatment with corticosteroids in meningeal tuberculosis has been widely reported.⁹⁴⁹⁻⁹⁶⁴ However, not all of the published reports were prospective, controlled trials.

In none of the nine prospective trials was treatment outcome worse in the group treated with corticosteroids, as compared to the control group. Survival in the corticosteroid treated group was improved in four, ^{957,960,961,964} tended to be better in one, ⁹⁶³ and was not better in four. ^{955,956,959,964} Fewer sequelae in the corticosteroid-treated group were found in four studies. ^{959,960,963,964}

Studies that looked at the stage of disease and the effects of corticosteroids found a lack of effect in mild and terminal disease stages (assessed by the degree of neurologic impairment), but a significant benefit for patients with intermediate disease stage.^{954,957,960} One of these studies⁹⁵⁷ determined that there was no difference in treatment outcome between doses of 1 mg and 10 mg of dexamethasone. The duration of corticosteroid treatment in this study was one month.

There is sufficient evidence to recommend the use of corticosteroids in moderate to severe meningeal and cerebral tuberculosis to improve survival, although not all patients will benefit from adjunctive corticosteroid treatment.

Corticosteroid treatment in other forms of tuberculosis

Corticosteroids have been given for other forms of tuberculosis.

In the treatment of endobronchial tuberculosis, adjunctive therapy with corticosteroids has been shown to be beneficial.^{965,966} In children with bronchial obstruction due to hilar lymphadenopathy, resolution of symptoms was faster and complications less frequent in corticosteroid-treated children compared to controls.⁹⁶⁷

Patients with peripheral lymphatic tuberculosis are known to frequently develop new nodes and draining abscesses during chemotherapy which are bacteriologically sterile and thought to represent an immunologic reaction.⁵³⁵ It would be desirable to evaluate the potential usefulness of adjunctive corticosteroid treatment, yet no controlled trial has investigated this issue.

The use of corticosteroids in the treatment of genitourinary tuberculosis has been reported, ^{968,969} but the design of the studies was insufficient to draw conclusion on their efficacy in reducing ureteral strictures.

The role of surgery in the chemotherapy era

Surgery has played a major role in the history of treatment of tuberculosis, and thoracic surgery was actually largely developed around the treatment of pulmonary tuberculosis.^{679,970,971} Efficacious chemotherapy has removed the need for surgical intervention in the routine treatment of patients.

The usual indications for surgery in the treatment of tuberculosis are for the treatment of complications. This is true for pyopneumothorax, respiratory distress due to massive pleural effusion, extensive restrictive pleural thickening, restrictive pericarditis, obstructive hydrocephalus, long-tract neurological signs in tuberculous spondylitis, and ureteral obstruction. There are, with the exception of extensive drug resistance, virtually no indications for surgery for primary treatment of tuberculosis. The guidelines for such surgery follow those for any other cause of such complications.

In industrialized countries, surgery has also been used with some success as an adjunct in patients with strains resistant to all or virtually all medications.⁹⁷²⁻⁹⁷⁶ Such services are not usually available in national tuber-culosis control programs of low-income countries, and are fortunately still rarely needed in most countries.

What will be summarized here are indications that are frequent and do not require sophisticated surgical procedures.

Surgical treatment in respiratory tract tuberculosis

Tuberculous pyopneumothorax

The development of an empyema or, more precisely, a tuberculous pyopneumothorax is a well-recognized complication in patients with cavitary tuberculosis whose cavities are located near the pleura.^{977,978} In such patients, penetration of anti-tuberculosis medications into the pleural space and the empyema might be sub-optimal and may even lead to acquisition of drug resistance.⁹⁷⁹ Furthermore, in contrast to pleural effusions, resorption of an empyema is less likely to occur, thus draining of the pus is usually indicated.

In the field, the most simple and effective approach is insertion of a drain, laid in such a way that it leads over two or three ribs before penetrating into the pleural space. The drain should be sutured to the skin. The patient is offered a bed that is about one meter above the floor, and the drainage is led into a bottle filled with water serving as a water lock. In patients whose entire lung is collapsed, full expansion of the lung can be expected, often leaving pleural thickening, however. As decortication is not usually an option, this is the best possible result that might be expected in such cases.

Pleural tuberculosis

Massive pleural effusions often require draining to relieve the patient from respiratory distress. Care should be taken not to drain more than about one liter in a single session to prevent cardiovascular and electrolyte disturbances. Accompanied by adequate chemotherapy, resolution of the accumulated fluid is usually prompt. Some authorities recommend complete draining of the effusion, ⁹³⁷ but it is uncertain whether this is really required.

Surgical treatment in tuberculosis of the spine

As previously shown (Chapter 1), chemotherapy alone of tuberculosis of the spine yields excellent results. Only the "radical Hong Kong operation" improves the results of chemotherapy somewhat, but not importantly so.⁵⁶² Because of the sophistication required, this procedure is beyond the capacity in the periphery of national programs where adequate chemotherapy alone is the key to success.

Superficial abscess might be drained by needle aspiration, but might also recur after the procedure. $^{980}\,$

Appendix 2 Active agents other than essential drugs and drug classes (second-line drugs)

Numerous drugs other than the first-line anti-tuberculosis drugs discussed in Chapter 1, and often called "second-line drugs", have shown activity against *M. tuberculosis*. Generally, these medications are less efficacious, associated with a higher frequency of adverse drug events, and are more expensive. In most low-income countries, these drugs are not routinely available in national programs. Where they are, their use is best limited to specialists with experience in dealing with adverse drug events.

Nevertheless, the global emergence of multidrug resistance (resistance to at least isoniazid and rifampicin) has brought up the discussion about their use on a wider scale. ^{605,607-609,611,981-983} For this reason, brief summaries of these drugs are presented here.

Aminoglycosides (other than streptomycin)

Amikacin

Amikacin is a semi-synthetic aminoglycoside, synthesized by Kawaguchi and collaborators through acetylation of the 1-aminogroup of the 2-desox-istreptamine moiety of kanamycin A at the Bristol-Banyu research laboratories in Japan, ⁹⁸⁴ and reported in 1972. ⁹⁸⁵

Amikacin has broad activity, particularly against gram-negative bacteria⁹⁸⁶ and is active against *M. tuberculosis*.⁹⁸⁷ Some researchers have found it to be usually more active against *M. tuberculosis* than other aminoglycosides.^{224,283} Cross-resistance with other aminoglycosides may occur, ²⁸³ and is particularly frequent with kanamycin, and incomplete with the polypeptide capreomycin.⁹⁸⁸ Amikacin is frequently active against streptomycin-resistant strains of *M. tuberculosis*.⁹⁸⁹ However, it appears to have little early bactericidal activity.⁹⁹⁰ Like other aminoglycosides, amikacin is not absorbed orally and the usual route of administration is intramuscular or intravenous, at a dose of 7.5 mg/kg to 15 mg/kg (depending on the dosage interval). Like other aminoglycosides, amikacin affects the neuromuscular junction and may lead to neuromuscular blockade, ^{409,410,991} an effect that may be reduced by lithium, ⁹⁹² but not reversed by neostigmine.

Indomethacin may interact with amikacin in newborns by increasing serum levels to toxic concentrations.⁹⁹³ Neurotoxicity might be increased by muscle relaxants such as tubocourarine, succinylcholine or decamethonium.

Kanamycin

Kanamycin was isolated from *Streptomyces kanamyceticus* by Umezawa and collaborators in 1957.⁹⁹⁴ It is a mixture of kanamycin A, B, and C.⁹⁹⁵⁻⁹⁹⁷

Kanamycin is active against a range of gram-negative bacteria and mycobacteria including *M. tuberculosis*.⁹⁹⁸

Kanamycin is not absorbed orally, but intramuscular administration leads to peak serum levels within an hour and the serum half-life is about four to six hours.⁹⁹⁸ It is mainly excreted through the kidneys and thus, as with all aminoglycosides, dose adjustments are warranted in patients with renal failure.⁹⁹⁸

The usual dosage of kanamycin is 0.5 g to 1 g per day.⁹⁹⁸

Similar to other aminoglycosides, eighth cranial nerve toxicity is the most important. Auditory toxicity is more pronounced than vestibular toxicity ⁹⁹⁸ and is very frequent, affecting up to 20% of patients after three months, and up to 60% if treatment lasts for six months. ⁹⁹⁹ Like other aminoglycosides, kanamycin may cause neuromuscular blockade. ⁴⁰⁹ Other adverse drug events include renal toxicity and, rarely, allergies. ⁹⁹⁸

As with other aminoglycosides, resistance is thought to be acquired through a single extrachromosomal plasmid factor with multi-step selection. ¹⁰⁰⁰ Capreomycin resistant strains are not usually resistant to kanamycin. ⁴⁷⁶ The inverse seems to be the case for low, but not for high kanamycin resistance. ⁴⁷⁶

Capreomycin

Capreomycin, a polypeptide antibiotic, was isolated from *Streptomyces capreolus* by Herr and collaborators at the Lilly Research Laboratories in 1959.¹⁰⁰¹

Capreomycin is active against various species of mycobacteria, including M. tuberculosis. ^{1002,1003}

Similar to aminoglycosides, capreomycin causes auditory, vestibular, and renal toxicity.¹⁰⁰⁴ Also like aminoglycosides, it is not orally absorbed and the usual administration is intramuscular. Rare adverse drug events include hypokalemia.¹⁰⁰⁴

Cross-resistance between kanamycin and capreomycin is incomplete. 1005

Cycloserine

Cycloserine (originally called orientomycin) was first isolated in 1952 from a *Streptomyces* strain, designated strain K-300 by Kurosawa.¹⁰⁰⁶ The identity with the compound discovered two years later in the United States was elucidated by Shoji, ¹⁰⁰⁷ and Mitui and Imaizumi in 1957, ¹⁰⁰⁸ but not before Lederle Laboratories also isolated the compound and recognized its identity with oxamycin, isolated by the Merck Laboratories, ¹⁰⁰⁹⁻¹⁰¹¹ and the Pfizer Laboratories which also isolated the compound. ¹⁰¹² Cycloserine can be isolated from *Streptomyces orchidaceus*, *S. garyphalus*, or *S. lavendulae*.

Cycloserine is active against *M. tuberculosis* and several species of gram-positive bacteria. 1006

Cycloserine inhibits cell wall synthesis^{1013,1014} by inhibiting the synthesis of peptidoglycan by blocking action of D-alanine racemase and D-alanine:alanine synthase.⁴⁶

Cycloserine is rapidly absorbed after oral administration, with a peak serum level of 10 to 50 mg/L following administration of 0.75 g to 1 g after 0.5 to 4 hours.

The usual dosage is two to three times daily (250 mg/day), 1015 but it is often given as a single dose.

The main adverse drug events due to cycloserine are neurologic and psychiatric, ^{159,1016-1022} although it has also been used in the treatment of mentally ill tuberculosis patients without observation of major mental toxicity. ¹⁰²³ In a summary of several reports, cycloserine toxic adverse drug events were reported in over 20%. ¹⁰⁰⁶ Most frequently reported or observed were vertigo and disorientation. Neuropsychiatric changes including drowsiness, slurred speech, psychoses, epileptiform reactions, ¹⁰¹⁶ as well as electroencephalographic changes and coma, were frequently noted. These effects of cycloserine are probably due to an interaction with the action of some monoamine oxidase inhibitors, as shown in experimental animals. ¹⁰²⁴

Cardiac depression, pareses, paresthesias and headache, pruritic rashes, drug fever, liver enzyme elevations, and gastrointestinal disturbances were less frequently reported adverse drug events. Smaller doses, and administration twice rather than once per day, reduce the frequency of adverse drug events. Stevens-Johnson syndrome has been reported in an HIV-infected patient.¹⁰²⁵

Cycloserine appears to interact with alcohol, increasing the toxic effects of alcohol. ¹⁰²⁶

Determination of resistance to cycloserine is difficult, and the correspondence of laboratory results with clinical data is poor (figure 90).⁴⁶⁶

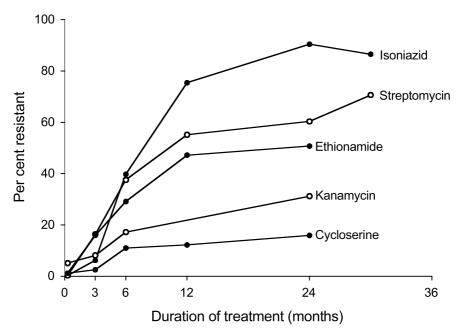


Figure 90. Proportion of strains of *M. tuberculosis* from resected lungs, *in vitro* resistant to anti-tuberculosis drugs, as a function of duration of treatment, the strain containing no susceptible organisms. Reproduced from⁴⁶⁶ by the permission of the publisher American Thoracic Society at the American Lung Association.

Para-aminosalicylic acid

In 1940, Bernheim demonstrated that salicylic acid and benzoic acid increased the oxygen consumption and carbon dioxide production of *M. tuberculosis*.¹⁰²⁷ Based on these observations, Lehmann investigated more than 50 derivatives of benzoic acid with the purpose of finding a substance possessing activities against *M. tuberculosis*. The most active compound he identified in the experiments was para-aminosalicylic acid, first published as preliminary results in the Lancet in 1946.¹⁰²⁸ Soon thereafter the first reports appeared, demonstrating its considerable anti-tuberculosis

activity in experimental models.²⁵⁶ The use of para-aminosalicylic acid became a core component in early combination therapy until its replacement by the better tolerated ethambutol.¹²²

It is likely that para-aminosalicylic acid, and not streptomycin, was the first anti-tuberculosis drug tested specifically against *M. tuberculosis*, as suggested in an editorial 1029 and correspondence of Lehman (reproduced with the permission of the South African Medical Journal): 1030,1031

"Dear Dr Dubovsky,

Your letter to the Director of the Central Laboratory at Sahlgrens Hospital was forwarded to me. It was the most remarkable letter I have received for many years. You are the first outside Sweden who has paid attention to the fact that PAS was in clinical use before streptomycin, eight months before ... Perhaps you wonder why I published the first paper on PAS so long after it was taken in clinical use. The reason was that as Ferrosan, a small company, had not taken out a patent on PAS, I didn't dare to publish the formula on PAS as other greater companies could take over the production of PAS ..."

The MIC of *M. tuberculosis* is 1 mg/L.¹⁰³²

In analogy with the observation that benzoic acid inhibits the respiration of tubercle bacilli, ¹⁰²⁷ para-aminosalicylic acid might be built into coenzyme F of the bacterium instead of para-aminobenzoic acid, and thereby inhibit growth. ⁴³⁰

Maximum serum concentrations with twice 4 g granular para-aminosalicylic acid are achieved within five to eight hours and remain above the minimum inhibitory concentration over the entire dosing interval.¹⁰³²

The granular form of para-aminosalicylic acid is better tolerated than the previously used tablet form. A dosage of 4 g twice daily of the granular form produces serum concentrations above the minimum inhibitory concentration over the entire dosing interval.¹⁰³² Good experiences with infusion therapy have also been reported.¹⁰³³

Para-aminosalicylic acid has been an unpleasant drug to take because of the bulk required and the frequency of adverse drug events, ¹⁰³⁴ which include gastrointestinal and cutaneous adverse drug events. ¹⁰³⁵ Para-aminosalicylic acid may cause hypothyroidism, ^{1036,1037} and intestinal malabsorption. ^{1038,1039} Among hematologic changes are thrombocytopenia in adults ^{1040,1041} and children. ¹⁰⁴²

Para-aminosalicylic acid has been reported to increase isoniazid blood levels.¹⁰⁴³⁻¹⁰⁴⁵ It may cause hypoglycemia in diabetics.¹⁰⁴⁶

Quinolones

Quinolones have a potential in the treatment of susceptible and drug-resistant tuberculosis. Quinolones that have been considered include ciprofloxacin, ¹⁰⁴⁷⁻¹⁰⁵⁶ clinafloxacin, ¹⁰⁵¹ difloxacin, ¹⁰⁵⁵ enoxacin, ¹⁰⁵⁵ fleroxacin, ¹⁰⁵⁷ gatifloxacin, ¹⁰⁵⁸ levofloxacin, ^{1051,1058-1060} lomefloxacin, ^{1061,1062} moxifloxacin, ^{1061,1063} norfloxacin, ¹⁰⁵⁵ ofloxacin, ^{1051,1053-1055,1064-1068} sitafloxacin, ¹⁰⁵⁸ sparfloxacin, ¹⁰⁵¹ temafloxacin, ¹⁰⁵¹ and tosufloxacin. ¹⁰⁵¹ Most clinical experience has been accumulated with ofloxacin and ciprofloxacin. Some of the quinolones have little or no activity against *M. tuberculosis*, while the potential of others is far greater. ¹⁰⁶⁹

Fluoroquinolones inhibit DNA gyrase of *M. tuberculosis*. ¹⁷⁶

The MICs of ciprofloxacin and ofloxacin are well below levels that can be achieved in serum.¹⁰⁵⁴ The early bactericidal activity of ciprofloxacin is, however, not as pronounced as that of isoniazid,¹⁰⁵⁶ and is inferior to that of ofloxacin.²⁷ Clinically, there is anecdotal evidence that even prolonged ciprofloxacin may not prevent reactivation of tuberculosis.¹⁰⁴⁷

Ciprofloxacin levels in bronchial biopsy specimens exceed those in the serum, indicating an accumulation of the compound in the lung parenchyma.¹⁰⁵²

A dosage of 600 mg to 800 mg of loxacin per day has been used successfully. $^{1067}\,$

In animals, quinolones induce changes in immature articular cartilage of weight-bearing joints, but these concerns have not been substantiated in children and adolescents.¹⁰⁷⁰ However, cases of arthropathy from ofloxacin among adults have been reported.¹⁰⁷¹

Antacids appear to lower serum concentrations of quinolones. 1072

Resistance to fluoroquinolones arises rapidly, and cross-resistance between quinolones is the rule.¹⁰⁷³ The most common cause of resistance results from mutations in the *gyrA* gene encoding the DNA gyrase,¹⁰⁷³ an enzyme required for replication and gene transcription.¹⁰⁷⁴ Quinolones should be used in combination with at least two other anti-tuberculosis drugs, as resistance might develop rapidly in a large proportion of patients.¹⁰⁷⁵

Rifamycins other than rifampicin

Rifabutin

Rifabutin is a semi-synthetic spiropiperidyl derivative of rifamycin S, 1076 which was synthesized in the research laboratories of Farmitalia Carlo Erba by Marsili *et al.*; its synthesis was announced in 1981. 1077

Rifabutin is active against a wide range of microorganisms, including gram-negative and gram-positive bacteria, and mycobacteria.^{1078,1079} In particular, among mycobacteria, it is more active against environmental species that are naturally resistant to rifampicin, ^{1080,1081} including *M. avium* complex.¹⁰⁸¹⁻¹⁰⁸⁵ While there is considerable cross-resistance with rifampicin, it is also active against a relative small subset of *M. tuberculosis* strains that have low resistance to rifampicin.¹⁰⁸³ However, this proportion is too small to make it a generally useful drug in rifampicin-resistant disease.¹⁰⁸⁶ Treatment results among patients with drug-susceptible organisms are similar to those obtained with rifampicin.¹⁰⁸⁷⁻¹⁰⁸⁹ However, studies on early bactericidal activity suggest that it is less active on extracellular bacilli than rifampicin.¹⁰⁹⁰

Rifabutin is more lipid soluble than rifampicin, thus tissue penetration is superior.¹⁰⁹¹ It has a longer terminal half-life, and is extensively metabolized.¹⁰⁹¹

The daily (and twice-weekly) recommended dosage of rifabutin is 5 mg/kg body weight.⁸⁹⁷

Adverse drug events reported with rifabutin treatment are similar to those with rifampicin treatment and include rash, hepatitis, fever, thrombocytopenia, orange-colored body fluids, arthralgia, uveitis, and leukopenia.^{897,1076} Some of these reactions may be potentiated through the interaction with anti-retroviral protease inhibitors.

Rifabutin induces hepatic metabolism, but not as markedly as rifampicin.¹⁰⁹¹ It does not affect the pharmacokinetics of antiretroviral drugs that are excreted in the urine.¹⁰⁹¹ A number of results from interaction studies show that rifabutin is a less potent inducer of the cytochrome P-450 family, and thus causes fewer clinically significant interactions than rifampicin, ¹⁰⁹² or they are less pronounced.²⁸¹ In particular, interactions with protease inhibitors are generally less than with rifampicin, ^{897,1093} and it is the rifamycin of choice for patients receiving highly active anti-retroviral therapy.

Resistance in sub-inhibitory concentrations is less rapidly acquired than with rifampicin.¹⁰⁹⁴ Similar to rifampicin, acquisition of resistance is frequently accompanied by mutations in the *rpoB* gene.¹⁰⁹⁵ However, up to 20% of rifampicin-resistant mutants with mutations in the *rpoB* gene are susceptible to rifabutin.¹⁰⁹⁶ This difference is not due to additional mechanisms of resistance, it is just that some of the mutations selected by rifampicin do not sufficiently modify the *rpoB* structure as to render this protein resistant to rifabutin (Telenti A, personal written communication, March 15, 2001).

Rifapentine

Rifapentine (cyclopentyl rifamycin SV) is a semisynthetic derivative of rifampicin, synthesized at the Lepetit laboratories in Italy. Its properties were first described in a publication in 1981.¹⁰⁹⁷

Rifapentine is comparable in its spectrum of activity to that of rifampicin.^{1098,1099} It is active in the experimental mouse model both against latent infection with *M. tuberculosis*¹¹⁰⁰ and clinical active disease.¹¹⁰¹

Rifapentine is an RNA synthesis inhibitor like rifampicin. 1099

The most conspicuous property of rifapentine is shown in a comparison of its pharmacokinetics with rifampicin. The serum elimination half-life is much longer in rifapentine¹⁷⁶ than rifampicin (figure 91).¹⁸¹ The serum elimination half-life $t\beta_{1/2}$ is 14 to 18 hours, ^{1102,1103} and is similar in adults and adolescents.¹¹⁰⁴ Intrapulmonary concentrations of rifapentine are below those in serum.¹¹⁰⁵ In contrast to rifampicin, higher peak levels are achieved following food intake than after fasting.¹¹⁰² Pharmacokinetics are not influenced by HIV status.¹¹⁰² A key issue that needs to be addressed is its high degree of plasma binding, which might require higher dosages than used so far.

The usual dosage is currently 600 mg twice-weekly.¹⁰⁹⁹ However, higher doses are now being studied.

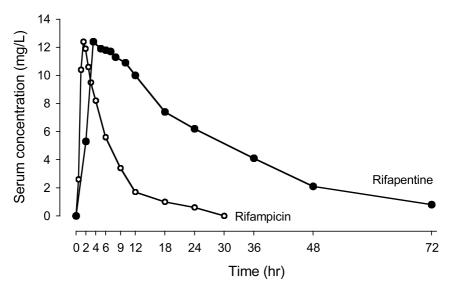


Figure 91. Comparative pharmacokinetics of rifampicin and rifapentine. Reproduced from^{181,1102} by the permission of the publisher ASM Press.

Adverse drug events similar to those associated with the use of rifampicin have been reported.¹⁰⁹⁹

Interactions that are expected most likely resemble those with rifampicin.

The pattern and mechanism of resistance to rifapentine is identical to that of rifampicin.

Thioamides

Following on from the discovery of the pyridine-containing isoniazid, numerous pyridine derivatives were tested, and the activity of thio-isonicotinamide against *M. tuberculosis* was found by several groups, 1106,1107 ethionamide, one of these thioamides, was introduced by the group of Liberman, Rist, and Grumbach. $^{1106-1108}$

Thioamides are active against *M. tuberculosis* and to a lesser extent against other mycobacteria. 1109

The mechanism of action of ethionamide is, like isoniazid, at the level of synthesis of mycolic acids.⁴⁶

Prothionamide is rapidly absorbed and rapidly excreted.¹¹¹⁰ Therefore, the daily dosage is usually divided into two doses. Ethionamide has excellent penetration into cerebrospinal fluid.⁵⁷⁰

The usual dosage of both ethionamide and prothionamide is 500 to 1,000 mg per day, divided into two doses.¹¹¹¹

The most important adverse drug event from thioamides are gastrointestinal disturbances and hepatotoxicity.¹¹¹²⁻¹¹¹⁹ It also appears to potentiate the hypothyroid effect of para-aminosalicylic acid. Comparisons between ethionamide and prothionamide seem to indicate that the latter might be less toxic than the former, ^{1111,1120} though the difference might not be important.

Although isoniazid and thioamide have the same parent compound, isonicotinic acid, isoniazid-resistant bacilli are often susceptible to ethionamide.¹¹⁰⁸

Drugs and drug classes with potential activity against *M. tuberculosis* **under investigation and development**

There can be little doubt about the necessity for the development of new anti-tuberculosis medications, given the limited amount of available choices.¹¹²¹ The Global Alliance for Tuberculosis Drug Development has

published a scientific blueprint for drug development that should assist in overcoming some of the barriers that impede the development, testing, and marketing of new compounds.¹⁰⁶⁹

Among the currently most promising candidates are long-acting rifamycins and fluoroquinolones (discussed above), oxazolidinone compounds, and nitroimidazopyrans. These and some other compounds under investigation are briefly summarized here.

Acetamides

Acetamides belong to a new class of compounds designed to inhibit the β -ketoacyl synthase reaction of fatty acid synthesis in mycobacteria.¹¹²² Because of their specific target, they exhibit virtually no activity against microorganisms other than mycobacteria. The MICs of the most potent compounds compare to those obtained with the most efficacious first-line drugs.¹¹²²

Amoxicillin plus clavulanic acid

M. tuberculosis possesses a beta-lactamase that might be responsible for its natural resistance to beta-lactam antibiotics.¹¹²³⁻¹¹²⁵ Thus, beta-lactam antibiotics are essentially inactive against tubercle bacilli. Clavulanic acid is a beta-lactamase blocker and, if given simultaneously with amoxicillin, makes the latter active against beta-lactam producing microorganisms. This combination has also been proposed and used in the treatment of drug-resistant tuberculosis.^{1126,1127}

Amoxicillin was synthesized in the Beecham Research Laboratories, patented in 1964, and described for the first time in 1970/71.^{1128,1129}

The addition of clavulanic acid to amoxicillin has shown *in vitro* activity against *M. tuberculosis*.¹¹³⁰⁻¹¹³³ Since then, several reports have appeared demonstrating successes in the treatment of patients with multidrug-resistant tuberculosis who also received amoxicillin plus clavulanic acid.¹¹³⁴⁻¹¹³⁷

The daily dosage might be 2 g of the combination. $^{\rm 402}$

The most important adverse drug event seen with beta-lactam antibiotics are hypersensitivity reactions, which might be immediate (urticaria, laryngeal edema, bronchospasm, hypotension, or local swelling), late (morbilliform rashes, serum sickness, or urticaria), or other than late reactions (toxic epidermal necrolysis, interstitial nephritis, pulmonary infiltration, vasculitis, hemolytic anemia, neutropenia, or thrombocytopenia).¹¹³⁸

Clarithromycin

Erythromycin, the prototype of the macrolide antibiotics, was first used to treat infections in 1952.¹¹³⁹

Clarithromycin is a macrolide that differs from erythromycin by the methylation of the hydroxyl group at position 6 on the lactone ring.¹¹⁴⁰

Clarithromycin has a wide anti-bacterial spectrum that includes mycobacteria. $^{1132,1139,1141-1147}$ Most frequently it has been used as prophylactic agent or against disease caused by *M. avium* complex. $^{1147-1154}$ While it shows *in vitro* activity against *M. tuberculosis* complex in human macrophages, 1145 the concentrations needed to inhibit growth appear to exceed those achievable in the serum and lung tissue of humans. 1155 It has therefore not been widely used in the treatment of tuberculosis.

Clarithromycin is usually rapidly absorbed and reaches C_{max} after two to three hours.¹¹⁴⁰ Its serum elimination half life $t\beta_{1/2}$ is 2.5 to 5 hours. It undergoes extensive hepatic metabolism. Because of its predominant renal excretion, dose adjustment might be necessary in patients with severe renal impairment.¹¹⁴⁰

Twice-daily 500 mg clarithromycin in AIDS patients was well tolerated, prevented *M. avium* complex disease and reduced mortality.¹¹⁴⁸ In the treatment of *M. avium* complex disease, a dosage of twice daily 1,000 mg has been given.^{1149,1156}

Fullerene derivatives

Fullerenes show an absolute lack of solubility in any polar solvent, and covalent attachment of solubilizing chains was therefore developed, resulting in the formation of water-soluble fulleropyrrolidines.¹¹⁵⁷ Certain such ionic fullerene derivatives have shown equally good activity against both susceptible and multidrug-resistant isolates of *M. tuberculosis*.

Nitroimidazopyrans

A series of nitroimidazopyrans, originally investigated as radiosensitizers for use in cancer chemotherapy, ¹¹⁵⁸ were shown to have *in vitro* and *in vivo* activity against *M. tuberculosis*. ^{1141,1159} However, the original compounds were shown to be mutagenic. ¹¹⁶⁰ Newer derivatives showed substantial activity against *M. tuberculosis* and lacked the mutagenicity shown previously with bicyclic nitroimidazoles. ¹¹⁶¹ There is considerable *in vivo* activity in the mouse model against *M. tuberculosis*, which is comparable

to that of isoniazid.¹¹⁶² It appears that the action is on both protein and lipid synthesis,¹¹⁶¹ inhibiting fatty acid and mycolic acid synthesis.¹¹⁶³

Similar to the nitroimidazoles (to which metronidazole belongs), nitrofurans show substantial *in vitro* bactericidal activity against bacilli held in a hypoxic stationary phase.¹¹⁶⁴

Because the recently synthesized nitroimidazopyran compound acts against multidrug-resistant tubercle bacilli, ¹¹⁶¹ it might prove a valuable agent in the future.

Oxazolidinones

Oxazolidinones are a class of protein synthesis inhibitors and include linezolid and eperezolid. ^{1165,1166}

Oxazolidinones have promising activity against a range of microorganisms including, and other than, *M. tuberculosis*.¹¹⁶⁶

Oxazolidinones appear to inhibit a step in the protein synthesis.¹¹⁶⁷ It has been proposed that they inhibit protein synthesis by binding to the 50S ribosomal subunit.¹¹⁶⁸

In experimental animals, oxazlidinones appear to be rapidly absorbed. ¹¹⁶⁶

Paromomycin

Paromomycin is an aminocyclosidic antibiotic complex, isolated from *S. rimosus* forma *paromomycinus* in 1959 in the Parke-Davies laboratories¹¹⁶⁹ in the same year as aminosidin was isolated from *S. chrestomyceticus* in the Farmitalia laboratories. It is also identical to catenulin, which was isolated from *S. catenulae* in 1952 in the Pfizer laboratories. Paromomycin is an antibiotic complex consisting of at least six antibiotics, and belongs to the neomycin family.¹¹⁷⁰

Its potential in the treatment of tuberculosis lies with the advantage that there is little cross-resistance with either streptomycin or with amikacin/kanamycin. Its early bactericidal activity indicates that it is at least as effective as amikacin.¹¹⁷¹ Its toxicity (similar to that of neomycin) may, however, preclude its prolonged parenteral use.

Phenothiazines

Phenothiazines are derivatives of methylene blue and are used in the management of psychosis. Originally, Paul Ehrlich had reported that methylene blue immobilized bacteria, and their evaluation as potential antimicrobial agents was thus only natural.¹¹⁷²⁻¹¹⁷⁴ However, the concentrations needed to exert activity by far exceed what is achievable within the non-toxic range. The argument for considering phenothiazines for the treatment of tuberculosis stems from the fact that pulmonary macrophages concentrate chlorpromazine (the first phenothiazine developed) 100-fold above what is found in plasma, concentrations that are active against mycobacteria *in vitro* ¹¹⁷² and *in vivo*.¹¹⁷⁵ Thioridazine, a well tolerated phenothiazine, has been shown to be active against both susceptible and resistant tubercle bacilli.¹¹⁷⁶⁻¹¹⁷⁸ Chlorpromazine has a titrable ability to slow the growth of intracellular tubercle bacilli *in vitro*.¹¹⁷⁹ Phenothiazines have not yet been tested for their activity against tuberculosis in humans.

Tuberactinomycin

Tuberactinomycin, a polypeptide, was isolated in the Toyo Jozo research laboratories in Japan from *Streptomyces griseoverticullatus* var. *tuberacticus* in 1966.¹¹⁸⁰ and was shown to be active against both kanamycin-susceptible and -resistant strains.^{1181,1182} Tuberactinomycin-N was semisynthetically derived from this compound and found to have stronger antimycobacterial activity and to be associated with less ototoxicity.^{1183,1184} Its use has been largely limited to East Asia, where it was found to be a useful alternative to capreomycin in the treatment of multidrug-resistant, aminoglycoside-resistant tuberculosis.

Appendix 3 Current vaccine development strategies

The incomplete protection that BCG provides against tuberculosis and its dismally disappointing effects in some areas have challenged researchers to develop a vaccine with better and more consistent performance. It is uncertain whether a much better vaccine can be developed in the near future, as the development of vaccines against bacteria that do not exert their pathogenicity through toxins has been fraught with difficulties. Vaccine development strategies currently being pursued include: ¹¹⁸⁵⁻¹¹⁸⁸

- Immunotherapy;
- Vaccination with saprophytic (environmental) mycobacteria;
- Auxotrophs;
- DNA vaccines;
- Recombinants;
- Subunits.

Immunotherapy with M. vaccae

M. vaccae is an environmental mycobacterium not known to cause disease in humans. A killed suspension of *M. vaccae* has been proposed, not to vaccinate against future tuberculosis, but to increase therapeutic response in the treatment of clinically manifest tuberculosis.¹¹⁸⁹

Numerous anecdotes have been published to illustrate its putative effects, but a clinical trial utilizing rigorous scientific standards has not shown any beneficial effect in addition to chemotherapy alone.¹¹⁹⁰ Another controlled clinical trial indicates that immunotherapy with *M. vaccae* may be effective. In that trial, sputum culture conversion at one month (but not at two months) was significantly higher among persons receiving *M. vaccae* compared to controls. In addition, radiographic improvement was swifter.¹¹⁹¹ In yet another trial, no relevant differences during treatment and a four-year follow-up were found.¹¹⁹²

However, an effect is difficult to demonstrate in the case of fully drugsusceptible tuberculosis, which responds superbly to chemotherapy. What is perhaps needed is to study its value in the treatment of multiple drugresistant tuberculosis, where the unequivocal outcome of death is a frequent enough event to permit a more definitive evaluation of the claims for improved immunologic response.

Vaccination with saprophytic (environmental) mycobacteria

The experiments by Palmer and Long have shown that various species of environmental mycobacteria provide some protection against experimental tuberculosis, but to different degrees and never exceeding that of BCG.⁸²⁶ As mentioned above, the trial in the United Kingdom and the observations in Malawi have lent further credibility to the hypothesis that certain environmental species might offer some protection against tuberculosis. This line of experimentation has been further pursued and evidence accumulated that animals vaccinated with environmental mycobacteria have an increased resistance to a subsequent challenge with virulent tubercle bacilli compared to non-vaccinated animals.⁸²⁸ So far, none of these vaccinations have been superior to BCG vaccination.

Auxotrophs

Another approach has been the development of so-called auxotrophic vaccines, where BCG and *M. tuberculosis* have been used to generate selected auxotrophs by insertional mutagenesis.¹¹⁸⁵ The advantage of such a vaccine might be that it would gradually die in the host (an advantage in immunocompromised hosts) and that a weakened auxotrophic *M. tuberculosis* might be more immunogenic than BCG.¹¹⁸⁵ However, survival time of the vaccine might be critical and has not yet been characterized sufficiently well to demonstrate superiority over BCG vaccination in experimental models,¹¹⁹³ but certain mutations in genes involved in amino-acid biosynthesis have been promising in experimental models.¹¹⁹⁴

DNA vaccines

Most efforts currently being undertaken are in the development of a DNA vaccine,¹¹⁹⁵⁻¹²⁰⁴ but none of the experimental models has yet shown superiority to BCG vaccine. There is, however, some experimental evidence that

this type of vaccine may not only protect against subsequent infection with *M. tuberculosis*, but may stimulate the immune response even among experimental animals with active disease.¹²⁰⁵ There are indications that the combination of priming with a DNA vaccine followed by a booster with BCG might induce higher protective efficacy in mice than BCG vaccination alone.¹²⁰⁶

Recombinants

Recombinant vaccines use existing microorganisms, e.g., vaccinia viruses¹²⁰⁷ or BCG, ^{1208,1209} which are genetically modified to produce additional antigens thought to enhance the immune response.¹¹⁸⁵ This approach is fairly recent and needs further research, which will most likely be aided by the deciphering of the entire sequence of the *M. tuberculosis* genome.¹²¹⁰ *In vitro*, BCG engineered to secrete recombinant human interferon-alpha was substantially more active than unaltered BCG in inducing interferon gamma in human peripheral blood mononuclear cells¹²⁰⁹ In a guinea pig model, such a recombinant vaccine was superior than two BCG strains in preventing gross lesions and dissemination.¹²¹¹

Subunits

Particular components (subunits) of *M. tuberculosis* may be better suited to inducing protective immunity than an entire organism. Recent research is thus evaluating the protective efficacy of such subunits.¹²¹² Preliminary experimental studies appear to be promising, providing protection similar to that obtained with BCG vaccination.^{1213,1214} Subunits are potentially specific and safe. A disadvantage of subunit vaccines is their limited persistence and thus potentially reduced duration of immune response.¹²¹⁵ In experimental mice models, re-challenge with a mycloyl transferase protein significantly boosted the protection against challenge with *M. tuberculosis* in animals whose immune protection had waned following BCG vaccination at birth.¹²¹⁶

References

- 1. Rieder HL. Epidemiologic basis of tuberculosis control. Paris: International Union Against Tuberculosis and Lung Disease, 1999; pp. 1-162.
- Rieder HL, Chonde TM, Myking H, Urbanczik R, Laszlo A, Kim SJ, Van Deun A, Trébucq A. The public health service national tuberculosis reference laboratory and the national laboratory network. Minimum requirements, role and operation in a low-income country. Paris: International Union Against Tuberculosis and Lung Disease, 1998; pp. 1-112.
- International Union Against Tuberculosis and Lung Disease. Technical guide. Sputum examination for tuberculosis by direct microscopy in low income countries. 5 ed. Paris: International Union Against Tuberculosis and Lung Disease, 2000; pp. 1-25.
- 4. World Health Organization. Laboratory services in tuberculosis control. Part I: Organization and management. Geneva: World Health Organization, 1998.
- 5. World Health Organization. Laboratory services in tuberculosis control. Part II: Microscopy. Geneva: World Health Organization, 1998.
- 6. World Health Organization. Laboratory services in tuberculosis control. Part III: Culture. Geneva: World Health Organization, 1998.
- 7. Crofton J, Horne N, Miller F. Clinical tuberculosis. 2 ed. London and Basingstoke: Macmillan Education Ltd, 1999; pp. 1-222.
- Enarson DA, Rieder HL, Arnadottir T, Trébucq A. Management of tuberculosis. A guide for low income countries. 5 ed. Paris: International Union Against Tuberculosis and Lung Disease, 2000; pp. 1-89.
- 9. Rieder HL. Opportunity for exposure and risk of infection: the fuel for the tuberculosis pandemic. (Editorial). Infection 1995; 23: 1-4.
- Rieder HL. Case finding in high- and low-prevalence countries. *In:* Reichman LB, Hershfield ES, Eds. Tuberculosis. A comprehensive international approach. New York Basel: Marcel Dekker, Inc., 2000; 323-339.
- 11. Grzybowski S, Barnett GD, Styblo K. Contacts of cases of active pulmonary tuberculosis. Bull Int Union Tuberc 1975; 50: 90-106.
- 12. Sutherland I. The epidemiology of tuberculosis is prevention better than cure? Bull Int Union Tuberc Lung Dis 1981; 56(3-4): 127-34.
- World Health Organization. Treatment of tuberculosis: guidelines for national programmes. Second edition. WHO/TB/97.220: 1-66. Geneva: WHO, 1997.
- 14. Peloquin CA. Serum concentrations of the antimycobacterial drugs. (Editorial). Chest 1998; 113: 1154-5.

- 15. Goldman AL, Braman SS. Isoniazid: a review with emphasis on adverse effects. Chest 1972; 62: 71-7.
- Knowles S, Uetrecht J, Shear NH. Idiosyncratic drug reactions: the reactive metabolite syndromes. Lancet 2000; 356: 1587-91.
- 17. Meyer H, Mally J. Über Hydrazinderivate der Pyridincarbonsäuren. Monatshefte für Chemie und verwandte Teile anderer Wissenschaften 1912; 23: 393-414.
- 18. Domagk G, Offe HA, Siefken W. Ein weiterer Beitrag zur experimentellen Chemotherapie der Tuberkulose (Neoteben). Deutsch Med Wschr 1952; 77: 573-8.
- Benson WM, Stefko PL, Roe MD. Pharmacologic and toxicologic observations on hydrazine derivatives of isonictoinic acid (RimifonTM, MarsilidTM). Am Rev Tuberc 1952; 65: 376-91.
- Bernstein J, Lott WA, Steinberg BA, Yale HL. Chemotherapy of experimental tuberculosis. V. Isonicotinic acid hydrazide (Nydrazid[™]) and related compounds. Am Rev Tuberc 1952; 65: 357-64.
- Suo J, Chang CR, Lin TP, Heifets LB. Minimal inhibitory concentrations of isoniazid, rifampin, ethambutol, and streptomycin against *Mycobacterium tuberculo*sis strains isolated before treatment of patients in Taiwan. Am Rev Respir Dis 1988; 138: 999-1001.
- 22. Inderlied CB, Salfinger M. Antimicrobial agents and susceptibility tests: mycobacteria. *In:* Murray PR, Baron EJ, Pfaller MA, Tenover FC, Yolken RH, Eds. Manual of clinical microbiology. Washington DC: ASM Press, 1995; 1385-1404.
- Lee CN, Heifets B. Determination of minimal inhibitory concentrations of antituberculosis drugs by radiometric and conventional methods. Am Rev Respir Dis 1987; 136: 349-52.
- 24. Jindani A, Aber VR, Edwards EA, Mitchison DA. The early bactericidal activity of drugs in patients with pulmonary tuberculosis. Am Rev Respir Dis 1980; 121: 939-49.
- 25. Jindani A. The effect of single and multiple drugs on the viable count of *M. tuber-culosis* in the sputum of patients with pulmonary tuberculosis during the early days of treatment. Thesis, University of London, 1979.
- 26. Hafner R, Cohn JA, Wright DJ, Dunlap NE, Egorin MJ, Enama ME, Muth K, Peloquin CA, Mor N, Heifets LB, DATRI 008 Study Group. Early bactericidal activity of isoniazid in pulmonary tuberculosis. Optimization of methodology. Am J Respir Crit Care Med 1997; 156: 918-23.
- Sirgel FA, Donald PR, Odhiambo J, Githui W, Umapathy KC, Paramasivan CN, Tam CM, Kam KM, Lam CW, Sole KM, Mitchison DA. A multicentre study of the early bactericidal activity of anti-tuberculosis drugs. J Antimicrob Chemother 2000; 45: 859-70.
- 28. World Health Organization. A concurrent comparison of home and sanatorium treatment of pulmonary tuberculosis in South India. Bull World Health Organ 1959; 21: 51-144.

- 29. Brooks SM, Lassiter NL, Young EC. A pilot study concerning the infection risk of sputum positive tuberculous patients on chemotherapy. Am Rev Respir Dis 1973; 108: 799-804.
- 30. Gunnels JJ, Bates JH, Swindoll H. Infectivity of sputum-positive tuberculous patients on chemotherapy. Am Rev Respir Dis 1974; 109: 323-30.
- Koch-Weser D, Ebert RH, Barclay WR, Lee VS. Studies on the metabolic significance of acid-fastness of tubercle bacilli. J Lab Clin Med 1953; 42: 828-9.
- 32. Winder FG, Collins PB. Inhibition by isoniazid of synthesis of mycolic acids in *Mycobacterium tuberculosis*. J Gen Microbiol 1970; 63: 41-8.
- 33. Sacchettini JC, Blanchard JS. The structure and function of the isoniazid target in *M. tuberculosis*. Res Microbiol 1996; 147: 36-43.
- Takayama K, Wang L, David HL. Effect of isoniazid on the in vivo mycolic acid synthesis, cell growth, and viability of *Mycobacterium tuberculosis*. Antimicrob Agents Chemother 1972; 2: 29-35.
- 35. Wang L, Takayama K. Relationship between the uptake of isoniazid and its action on in vivo mycolic acid synthesis. Antimicrob Agents Chemother 1972; 2: 438-41.
- Takayama K, Schnoes HK, Armstrong EL, Boyle RW. Site of inhibitory action of isoniazid in the synthesis of mycolic acids in *Mycobacterium tuberculosis*. J Lipid Res 1975; 16: 308-17.
- Davidson LA, Takayama K. Isoniazid inhibition of the synthesis of monounsaturated long-chain fatty acids in *Mycobacterium tuberculosis* H37Ra. Antimicrob Agents Chemother 1979; 16: 104-5.
- Middlebrook G. Isoniazid-resistance and catalase activity of tubercle bacilli. A preliminary report. Am Rev Tuberc 1954; 69: 471-2.
- 39. Winder F. Catalase and peroxidase in mycobacteria. Possible relationship to the mode of action of isoniazid. Am Rev Respir Dis 1960; 81: 68-78.
- 40. Youati J. A review of the action of isoniazid. Am Rev Respir Dis 1969; 99: 729-50.
- 41. Zhang Y, Heym B, Allen B, Young D, Cole S. The catalase-peroxidase gene and isoniazid resistance of *Mycobacterium tuberculosis*. Nature 1992; 358: 591-3.
- 42. Heym B, Zhang Y, Poulet S, Young D, Cole ST. Characterization of the *KatG* gene encoding a catalase-peroxidase required for the isoniazid susceptibility of *Mycobacterium tuberculosis*. J Bacteriol 1993; 175: 4255-9.
- Stoeckle MY, Guan L, Riegler N, Weitzman I, Kreiswirth B, Kornblum J, Laraque F, Riley LW. Catalase-peroxidase gene sequences in isoniazid-sensitive and -resistant strains of *Mycobacterium tuberculosis* from New York City. J Infect Dis 1993; 168: 1063-5.

- 44. Heym B, Alzari PM, Honoré N, Cole ST. Misssense mutations in the catalaseperoxidase gene, *katG*, are associated with isoniazid resistance in *Mycobacterium tuberculosis*. Mol Microbiol 1995; 15: 235-45.
- 45. Somoskövi A, Parsons LM, Salfinger M. The molecular basis of resistance to isoniazid, rifampin, and pyrazinamide in *Mycobacterium tuberculosis*. Respir Res 2001; 2: 164-8.
- 46. Zhang Y, Telenti A. Genetics of drug resistance in *Mycobacterium tuberculosis*. *In:* Hatfull GF, Jacobs WR, Jr., Eds. Molecular genetics of mycobacteria. Washington, DC: ASM Press, 2000; 235-254.
- 47. Slayden RA, Barry CE, III. The genetics and biochemistry of isoniazid resistance in *Mycobacterium tuberculosis*. Microbes Infection 2000; 2: 659-69.
- 48. Banerjee A, Dubnau E, Quemard A, Balasubramanian V, Um KS, Wilson T, Collis D, de Lisle G, Jacobs WR, Jr. *inhA*, a gene encoding a target for isoniazid and ethionamide in *Mycobacterium tuberculosis*. Science 1994; 263: 227-30.
- 49. Musser JM, Kapur V, Williams DL, Kreiswirth BN, van Soolingen D, van Embden JDA. Characterization of the catalase-peroxidase gene (*katG*) and *inhA* locus in isoniazid-resistant and -susceptible strains of *Mycobacterium tuberculosis* by automated DNA sequencing: restricted array of mutations associated with drug resistance. J Infect Dis 1996; 173: 196-202.
- 50. Parsons LM, Driscoll JR, Taber HW, Salfinger M. Drug resistance in tuberculosis. Infect Dis Clin N Am 1997; 11: 905-28.
- 51. Heym B, Stavropoulos E, Honoré N, Domenech P, Saint-Joanis B, Wilson TM, Collins DM, Colston MJ, Cole ST. Effects of overexpression of the alkyl hydroperoxide reductase *ahpC* on the virulence and isoniazid resistance of *Mycobacterium tuberculosis*. Infect Immun 1997; 65: 1395-401.
- 52. Canetti G, Grosset J. Teneur de souches sauvages de *Mycobacterium tuberculosis* en variants résistants à l'isoniazide et en variants résistants à la streptomycine sur milieu de Loewenstein-Jensen. Ann Inst Pasteur 1961; 101: 28-46.
- 53. David HL. Probability distribution of drug-resistant mutants in unselected populations of *Mycobacterium tuberculosis*. Appl Microbiol 1970; 20: 810-4.
- 54. Mitchison DA. Drug resistance in mycobacteria. Br Med Bull 1984; 40: 84-90.
- 55. Peloquin CA, Namdar R, Dodge AA, Nix DE. Pharmacokinetics of isoniazid under fasting conditions, with food, and with antacids. Int J Tuberc Lung Dis 1999; 3: 703-10.
- 56. Davidson PT, Hanh LQ. Antituberculosis drugs. Clin Chest Med 1986; 7: 425-38.
- 57. Kergueris MF, Bourin M, Larousse C. Pharmacokinetics of isoniazid: influence of age. Eur J Clin Pharmacol 1986; 30: 335-40.
- 58. Weber WW, Hein DW. Clinical pharmacokinetics of isoniazid. Clin Pharmacokinetics 1979; 4: 401-22.

- Sarma GR, Kailasam S, Nair NGK, Narayana ASL, Tripathy SP. Effect of prednisolone and rifampin on isoniazid metabolism in slow and rapid inactivators of isoniazid. Antimicrob Agents Chemother 1980; 18: 661-6.
- Parkin DP, Vandenplas S, Botha FJH, Vandenplas ML, Seifart HI, van Helden PD, van der Walt BJ, Donald PR, van Jaarsveld PP. Trimodality of isoniazid elimination. Phenotype and genotype in patients with tuberculosis. Am J Respir Crit Care Med 1997; 155: 1717-22.
- Donald PR, Gent WL, Seifart HI, Lamprecht JH, Parkin DP. Cerebrospinal fluid isoniazid concentrations in children with tuberculous meningitis: the influence of dosage and acetylation status. Pediatrics 1992; 89: 247-50.
- Siskind MS, Thienemann D, Kirlin L. Isoniazid-induced neurotoxicity in chronic dialysis patients: report of three cases and a review of the literature. Nephron 1993; 64: 303-6.
- 63. Blanchard PD, Yao JDC, McAlpine DE, Hurt RD. Isoniazid overdose in the Cambodian population of Olmsted County, Minnesota. JAMA 1986; 256: 3131-3.
- 64. Asnis DS, Bhat JG, Melchert AF. Reversible seizures and mental status changes in a dialysis patient on isoniazid preventive therapy. Ann Pharmacother 1993; 27: 444-6.
- 65. Nolan CM, Elarth AM, Barr HW. Intentional isoniazid overdosage in young Southeast Asian refugee women. Chest 1988; 93: 803-6.
- 66. Spalding CT, Buss WC. Toxic overdose of isoniazid, rifampicin and ethambutol. Eur J Clin Pharmacol 1986; 30: 381-2.
- 67. Shah BR, Santucci K, Sinert R, Steiner P. Acute isoniazid neurotoxicity in an urban hospital. Pediatrics 1995; 95: 700-4.
- 68. Martinjak-Dvorsek I, Gorjup V, Horvat M, Noc M. Acute isoniazid neurotoxicity during preventive therapy. Crit Care Med 2000; 28: 567-8.
- 69. Gnam W, Flint A, Goldbloom D. Isoniazid-induced hallucinosis: response to pyridoxine. (Correspondence). Psychosomatics 1993; 34: 537-9.
- 70. Ibrahim ZY, Menke JJ. Comment: Isoniazid-induced psychosis. (Correspondence). Ann Pharmacother 1994; 28: 1311.
- Zander Olsen P, Tørning K. Isoniazid and loss of memory. Scand J Respir Dis 1968; 49: 1-8.
- 72. Jimenez-Lucho VE, Del Busto R, Odel J. Isoniazid and ethambutol as a cause of optic neuropathy. Eur J Respir Dis 1987; 71: 42-5.
- 73. Ishii N, Nishihara Y. Pellagra encephalopathy among tuberculous patients: its relation to isoniazid therapy. J Neurology, Neurosurg Psych 1985; 48: 628-34.
- 74. Muratake T, Watanabe H, Hayashi S. Isoniazid-induced pellagra and the *N*-acetyl-transferase gene genotype. (Correspondence). Am J Psychiatry 1999; 156: 660.
- 75. Bottomley SS. Sideroblastic anaemia. Clin Haematol 1982; 11: 389-409.

- 76. McCurdy PR, Donohoe RF. Pyridoxine-responsive anemia conditioned by isonicotinic acid hydrazide. Blood 1966; 27: 352-62.
- 77. Hankins DG, Saxena K, Faville RJ, Warren BJ. Profound acidosis caused by isoniazid ingestion. Am J Emerg Med 1987; 5: 165-6.
- 78. Sievers ML, Herrier RN. Treatment of acute isoniazid toxicity. Am J Hosp Pharm 1975; 32: 202-6.
- 79. Snider DE. Pyridoxine supplementation during isoniazid therapy. Tubercle 1980; 61: 191-6.
- 80. Wason S, Lacouture PG, Lovejoy FH. Single high-dose pyridoxine treatment for isoniazid overdose. JAMA 1981; 246: 1102-4.
- 81. Chan TYK. Pyridoxine ineffective in isoniazid-induced psychosis. (Correspondence). Ann Pharmacother 1999; 33: 1123-4.
- 82. Siefkin AD, Albertson TE, Corbett MG. Isoniazid overdose: pharmacokinetics and effects of oral charcoal in treatment. Human Toxicol 1987; 6: 1-5.
- 83. Scolding N, Ward MJ, Hutchings A, Routledge PA. Charcoal and isoniazid pharmacokinetics. Human Toxicol 1986; 5: 285-6.
- 84. Orlowski JP, Paganini EP, Pippenger CE. Treatment of a potentially lethal dose isoniazid ingestion. Ann Emerg Med 1988; 17: 73-6.
- 85. Rothfield NF, Bierer WF, Garfield JW. Isoniazid induction of antinuclear antibodies. Ann Intern Med 1978; 88: 650-2.
- 86. Price EJ, Venables PJW. Drug-induced lupus. Drug Safety 1995; 12: 283-90.
- 87. Robinson MG, Foadi M. Hemolytic anemia with positive Coombs' test. Association with isoniazid therapy. JAMA 1969; 208: 656-8.
- 88. Ferguson A. Agranulocytosis during isoniazid therapy. (Correspondence). Lancet 1952; 2: 1179.
- Mehrotra TN, Gupta SK. Agranulocytosis following isoniazid. Report of a case. Ind J Med Sci 1973; 27: 292-3.
- 90. Mielke HG. Aplastische Anämie (Erythroblastophthise) nach INH-Behandlung. Folia Haematol Neue Folge 1958; 2: 1-10.
- 91. Goodman SB, Block MH. A case of red cell aplasia occurring as a result of antituberculous therapy. Blood 1964; 24: 616-23.
- Claiborne RA, Dutt AK. Isoniazid-induced pure red cell aplasia. Am Rev Respir Dis 1985; 131: 947-9.
- Holdiness MR. A review of blood dyscrasias induced by the antituberculosis drugs. Tubercle 1987; 68: 301-9.
- 94. FitzGerald JM, Turner MT, Dean S, Elwood RK. Alopecia side-effect of antituberculosis drugs. (Correspondence). Lancet 1996; 347: 472.
- 95. Gabrail NY. Severe febrile reaction to isoniazid. Chest 1987; 91: 620-1.

- 96. Asai S, Shimoda T, Hara K, Fujiwara K. Occupational asthma caused by isonicontinic acid hydrazide (INH) inhalation. J Allerg Clin Immunol 1987; 80: 578-82.
- Polosa R, Colombrita R, Prosperini G, Cacciola R. A case of acute deterioration in asthma symptoms induced by isoniazid prophylaxis. Respir Med 1997; 91: 438-40.
- 98. Bomb BS, Purohit SD, Bedi HK. Stevens-Johnson syndrome caused by isoniazid. Tubercle 1976; 57: 229-30.
- 99. Yamasaki R, Yamasaki M, Kawasaki Y, Nagasako R. Generalized pustular dermatosis caused by isoniazid. Br J Dermatol 1985; 112: 504-6.
- 100. Holdiness MR. Contact dermatitis to antituberculosis drugs. Contact Dermatitis 1986; 15: 282-8.
- Kopanoff DE, Snider DE, Caras GJ. Isoniazid-related hepatitis. Am Rev Respir Dis 1978; 117: 991-1001.
- 102. Gal AA, Klatt EC. Fatal isoniazid hepatitis in a child. (Correspondence). Pediatr Infect Dis 1986; 5: 490-1.
- 103. Murphy R, Swartz R, Watkins PB. Severe actetominophen toxicity in a patient receiving isoniazid. (Correspondence). Ann Intern Med 1990; 113: 799-800.
- 104. Moulding TS, Redeker AG, Kanel GC. Acetominophen, isoniazid, and hepatic toxicity. Ann Intern Med 1991; 114: 431.
- 105. Stephenson I, Qualie M, Wiselka MJ. Hepatic failure and encephalopathy attributed to an interaction between acetaminophen and rifampicin. (Correspondence). Am J Gastroenterol 2001; 96: 1310-1.
- 106. Tuberculosis Chemotherapy Centre Madras. The prevention and treatment of isoniazid toxicity in the therapy of pulmonary tuberculosis. 1. An assessment of two vitamin B preparations and glutamic acid. Bull World Health Organ 1963; 28: 455-75.
- 107. Tuberculosis Chemotherapy Centre Madras. The prevention and treatment of isoniazid toxicity in the therapy of pulmonary tuberculosis. 2. An assessment of the prophylactic effect of pyridoxine in low dosage. Bull World Health Organ 1963; 29: 457-81.
- 108. McCune R, Deuschle K, McDermott W. The delayed appearance of isoniazid antagonism by pyridoxine in vivo. Am Rev Tuberc Pulm Dis 1957; 76: 1100-5.
- O'Brien RJ, Long MW, Cross FS, Lyle MA, Snider DE, Jr. Hepatotoxicity from isoniazid and rifampin among children treated for tuberculosis. Pediatrics 1983; 72: 491-9.
- 110. Riska N. Hepatitis cases in isoniazid treated groups and in a control group. Bull Int Union Tuberc 1976; 51: 203-8.
- 111. Comstock GW, Edwards PQ. The competing risks of tuberculosis and hepatitis for adult tuberculin reactors. (Editorial). Am Rev Respir Dis 1975; 111: 573-7.

- 112. Van den Brande P, van Steenbergen W, Vervoort G, Demedts M. Aging and hepatotoxicity of isoniazid and rifampin in pulmonary tuberculosis. Am J Respir Crit Care Med 1995; 152: 1705-8.
- 113. Pande JN, Singh SPN, Khilnani GC, Tandon RK. Risk factors for hepatotoxicity from antituberculosis drugs: a case-control study. Thorax 1996; 51: 132-6.
- 114. Nolan CM, Goldberg SV, Buskin SE. Hepatotoxicity associated with isoniazid preventive therapy. A 7-year survey from a public health tuberculosis clinic. JAMA 1999; 281: 1014-8.
- 115. Leonin TA, Julian EV, Baluis CR. A review of the hepatotoxic effects of anti-TB drugs at the Veterans Memorial Medical Center. Chest 1979; 11: 140-8.
- 116. Maddrey WC, Boitnott JK. Isoniazid hepatitis. Ann Intern Med 1973; 79: 1-12.
- 117. Lauterburg BH, Smith CV, Todo EL, Mitchell JR. Pharmacokinetics of the toxic hydrazino metabolites formed from isoniazid in humans. J Pharmacol Experim Therapeutics 1985; 235: 566-70.
- 118. Gangadharam PRJ. Isoniazid, rifampin, and hepatotoxicity. (Editorial). Am Rev Respir Dis 1986; 133: 963-5.
- 119. Martinez-Roig A, Cami J, Llorens-Terol J, de la Torre R, Perich F. Acetylation phenotype and hepatotoxicity in the treatment of tuberculosis in children. Pediatrics 1986; 77: 912-5.
- 120. Girling DJ. The hepatic toxicity of antituberculosis regimens containing isoniazid, rifampicin and pyrazinamide. Tubercle 1978; 59: 13-32.
- 121. Ellard GA, Girling DJ, Nunn AJ. The hepatotoxicity of isoniazid among three acetylator phenotypes. (Corrspondence). Am Rev Respir Dis 2001; 123: 568-70.
- 122. Fox W, Ellard GA, Mitchison DA. Studies on the treatment of tuberculosis undertaken by the British Medical Research Council Tuberculosis Units, 1946-1986, with relevant subsequent publications. Int J Tuberc Lung Dis 1999; 3 (suppl 2): S231-S279.
- 123. International Union Against Tuberculosis Committee on Prophylaxis. Efficacy of various durations of isoniazid preventive therapy for tuberculosis: five years of follow-up in the IUAT trial. Bull World Health Organ 1982; 60: 555-64.
- 124. Askgaard DS, Wilcke T, Dossing M. Hepatotoxicity caused by the combined action of isoniazid and rifampicin. Thorax 1995; 50: 213-4.
- 125. Campbell IA. Toxicity of isoniazid and rifampicin in combination. (Correspondence). Thorax 1994; 50: 814.
- 126. Muakkassah SF, Bidlack WR, Yang WCT. Mechanism of the inhibitory action of isoniazid on microsomal drug metabolism. Biochem Pharmacol 1981; 30: 1651-8.
- 127. Baciewicz AM, Self TH. Isoniazid interactions. South Med J 1985; 78: 714-8.
- 128. Kottegoda SR. Cheese, wine, and isoniazid. (Correspondence). Lancet 1985; 2: 1074.

- 129. Hauser MJ, Baier H. Interaction of isoniazid with foods. Drug Intell Clin Pharm 1982; 16: 617-8.
- 130. Self TH, Chrisman CR, Baciewicz AM, Bronze MS. Isoniazid drug and food interactions. Am J Med Sci 1999; 317: 304-11.
- 131. Uragoda CG, Kottegoda SR. Adverse reactions to isoniazid on ingestion of fish with a high histamine content. Tubercle 1977; 58: 83-9.
- 132. O'Sullivan TL. Drug-food interaction with isoniazid resembling anaphylaxis. (Correspondence). Ann Pharmacother 1997; 31: 928-9.
- 133. Uragoda CG. Histamine poisoning in tuberculous patients after ingestion of tuna fish. Am Rev Respir Dis 1980; 121: 157-9.
- 134. Morinaga S, Kawasaki A, Hirata H, Suzuki S, Mizushima Y. Histamine poisoning after ingestion of spoiled raw tuna in a patient taking isoniazid. Intern Med 1997; 36: 198-200.
- 135. Smith CK, Durack DT. Isoniazid and reaction to cheese. (Correspondence). Ann Intern Med 1978; 88: 520-1.
- 136. Acres SE, Paulson E. Histamine poisoning in a patient on isoniazid. Canada Comm Dis Rep 1980; 7: 79-80.
- 137. Uragoda CG, Lodha SC. Histamine intoxication in a tuberculous patient after ingestion of cheese. Tubercle 1979; 60: 59-61.
- 138. Lejonc JL, Gusmini D, Brochard P. Isoniazid and reaction to cheese. (Correspondence). Ann Intern Med 1979; 91: 793.
- Boman G, Borg O, Hanngren Å, Mamlborg AS, Sjöqvist F. Pharmacokinetic interactions between the tuberculostatic rifampicin, para-aminosalicylic acid and isoniazid. (Abstract). Acta Pharmacol Toxicol (Copenh) 1970; 28(suppl 1): No. 4.
- 140. Grange JM, Winstanley PA, Davies PDO. Clinically significant drug interactions with antituberculosis agents. Drug Safety 1994; 11: 242-51.
- 141. Berkowitz FE, Henderson SL, Fajman N, Schoen B, Naughton M. Acute liver failure caused by isoniazid in a child receiving carbamazepine. Int J Tuberc Lung Dis 1998; 2: 603-6.
- 142. Dockweiler U. Isoniazid-induced valproic-acid toxicity, or vice versa. (Correspondence). Lancet 1987; 2: 152.
- Höglund P, Nilsson LG, Paulsen O. Interaction between isoniazid and theophylline. Eur J Respir Dis 1987; 70: 110-6.
- 144. Engelhard D, Stutman HR, Marks MI. Interaction of ketoconazole with rifampin and isoniazid. N Engl J Med 1984; 311: 1681-3.
- Nuñez-Vergara LJ, Yudelevich J, Squella JA, Speisky H. Drug-aldehyde interactions during ethanol metabolism *in vitro*. Alcohol Alcoholism 1991; 26: 139-46.
- 146. Rosenthal AR, Self TH, Baker ED, Linden RA. Interaction of isoniazid and warfarin. JAMA 1977; 238: 2177.

- 147. Murray FJ. Outbreak of unexpected reactions among epileptics taking isoniazid. Am Rev Respir Dis 1962; 86: 729-32.
- 148. Kay L, Kampmann JP, Svendsen T, Vergman B, Hansen JEM, Skovsted L, Kristensen M. Influence of rifampicin and isoniazid on the kinetics of phenytoin. Br J Clin Pharmac 1985; 20: 323-6.
- Walubo A, Aboo A. Phenytoin toxicity due to concomitant anti-tuberculosis therapy. S Afr Med J 1995; 85: 1175-6.
- 150. Wright JM, Stokes EF, Sweeney VP. Isoniazid-induced carbamazepine toxicity and vice versa. N Engl J Med 1982; 307: 1325-7.
- 151. García B, Zaborras E, Areas V, Obeso G, Jiménez I, de Juana P, Bermejo T. Interaction between isoniazid and carbamazepine potentiated by cimetidine. (Correspondence). Ann Pharmacother 1992; 26: 841-2.
- Pippenger CE. Clinically significant carbamazepine drug interactions: an overview. Epilepsia 2001; 28(suppl 3): S71-S76.
- 153. Hoyt Block S. Carbamazepine-isoniazid interaction. Pediatrics 1982; 69: 494-5.
- 154. van Wieringen A, Vrijlandt CM. Ethosuximide intoxication caused by the interaction with isoniazid. Neurology 1983; 33: 1227-8.
- 155. Leimenstoll G, Schlegelberger T, Fulde R, Niedermayer W. Interaktion von Ciclosporin und Ethambutol-Isoniazid. Deutsche Med Wschr 1988; 113: 514-5.
- 156. Patsalos PN, Duncan JS. Antiepileptic drugs. A review of clinically significant interactions. Drug Safety 1993; 9: 156-84.
- 157. Jonville AP, Gauchez AS, Autret E, Billard C, Barbier P, Nsabiyumva F, Breteau M. Interaction between isoniazid and valproate: a case of valproate overdosage. Eur J Clin Pharmacol 1991; 40: 197-8.
- 158. Carrión C, Espinosa E, Herrero A, García B. Possible vincristine-isoniazid interaction. (Correspondence). Ann Pharmacother 1995; 29: 201.
- 159. Abernethy DR, Greenblatt DJ, Ochs HR, Shadex RI. Benzodiazepine drug-drug interactions commonly occurring in clinical practice. Curr Med Res Opin 1984; 8(suppl 4): 80-93.
- 160. Ochs HR, Greenblatt DJ, Roberts GB, Dengler HJ. Diazepam interaction with antituberculosis drugs. Clin Pharmacol Ther 1981; 29: 671-8.
- Ochs HR, Greenblatt DJ, Knüchel M. Differential effect of isoniazid on triazolam oxidation and oxazepam conjugation. Br J Clin Pharmac 1983; 16: 743-6.
- 162. Takeda M, Nishinuma K, Yamashita S, Matsubayashi T, Tanino S, Nishimura T. Serum haloperidol levels of schizophrenics receiving treatment for tuberculosis. Clin Neuropharmacol 1986; 9: 386-97.
- 163. DiMartini A. Isoniazid, tricyclics and the "cheese reaction". Intern J Pschopharmacol 1995; 10: 197-8.
- 164. Torrent J, Izquierdo I, Cabezas R, Jané F. Theophylline-isoniazid interaction. DICP Ann Pharmacother 1989; 23: 143-5.

- 165. Ahn HC, Yang JH, Lee HB, Rhee YK, Lee YC. Effect of combined therapy of oral anti-tubercular agents on theophylline pharmacokinetics. Int J Tuberc Lung Dis 2000; 4: 784-7.
- 166. Judd FK, Mijch AM, Cockram A, Norman TR. Isoniazid and anti-depressants: is it a cause for concern? Intern Clin Psychopharmacol 1994; 9: 123-5.
- Malek-Ahmadi P, Chavez M, Contreras SA. Coadministration of isoniazid and antidepressant drugs. (Correspondence). J Clin Psychiatry 1996; 57: 550.
- 168. Gannon R, Pearsall W, Rowley R. Isoniazid, meperidine, and hypothension. (Correspondence). Ann Intern Med 1983; 99: 415.
- Morgan JP. Isoniazid and levodopa. (Correspondence). Ann Intern Med 1980; 92: 434.
- 170. Mazze RI, Woodruff RE, Heerdt ME. Isoniazid-induced enflurane defluorination in humans. Anesthesiology 1982; 57: 5-8.
- 171. Sensi P, Margalith P, Timbal MT. Rifomycin, a new antibiotic preliminary report. (Correspondence). Farmaco Ed Sci 1959; 14: 146-7.
- 172. Sensi P. A family of new antibiotics, the rifamycins. Res Prog Org Biol Chem 1964; 1: 338-421.
- 173. Oppolzer W, Prelog V, Sensi P. Konstitution des Rifamycins B und verwandter Rifamycine. Experientia 1964; 20: 336-9.
- 174. Maggi N, Pasqualucci CR, Ballotta R, Sensi P. Rifampicin: a new orally active rifamycin. Chemotherapia 1966; 11: 285-92.
- 175. Kenny MT, Strates B. Metabolism and pharmacokinetics of the antibiotic rifampin. Drug Metabolism Rev 1981; 12: 159-218.
- 176. Telenti A. Genetics of drug resistant tuberculosis. Thorax 1998; 53: 793-7.
- 177. Miller LP, Crawford JT, Shinnick TM. The *rpoB* gene of *Mycobacterium tuber-culosis*. Antimicrob Agents Chemother 1994; 38: 805-11.
- 178. Telenti A, Lowrie D, Matter L, Imboden P, Cole S, Schopfer K, Marchesi F, Colston MJ, Bodmer T. Detection of rifampicin-resistance mutations in *Mycobacterium tuberculosis*. Lancet 1993; 341: 647-50.
- 179. Cole ST. Rifamycin resistance in mycobacteria. Res Microbiol 1996; 147: 48-52.
- 180. Acocella G. Clinical pharmacokinetics of rifampicin. Clin Pharmacokinetics 1978; 3: 108-27.
- 181. Peloquin CA, Jaresko GS, Yong CL, Keung ACF, Bulpitt AE, Jelliffe RW. Population pharmacokinetic modeling of isoniazid, rifampin, and pyrazinamide. Antimicrob Agents Chemother 1997; 41: 2670-9.
- 182. Pähkla R, Lambert J, Ansko P, Winstanley P, Davies PDO, Kiivet RA. Comparative bioavailability of three different preparations of rifampicin. J Clin Pharm Ther 1999; 24: 219-25.

- 183. Acocella G, Conti R, Luisetti M, Pozzi E, Grassi C. I. Absorption and metabolism of the compounds used in the initial intensive phase of the short-course regiments: single administration study. Am Rev Respir Dis 1985; 132: 510-5.
- 184. Maggi N, Furesz S, Pallanza R, Pelizza G. Rifampicin desacetylation in the human organism. Arzneim -Forsch /Drug Res 1969; 19: 651-4.
- 185. Scotti R. Sex difference in blood levels of some antibiotics. Chemotherapy 1973; 18: 205-11.
- 186. Siegler DI, Burley DM, Bryant M, Citron KM, Standen SM. Effect of meals on rifampicin absorption. Lancet 1971; 2: 197-8.
- 187. Peloquin CA, Namdar R, Singleton MD, Nix DE. Pharmacokinetics of rifampin under fasting conditions, with food, and with antacids. Chest 1999; 115: 12-8.
- 188. Purohit SD, Sarkar SK, Gupta ML, Jain DK, Gupta PR, Mehta YR. Dietary constituents and rifampicin absorption. (Correspondence). Tubercle 1987; 68: 151-2.
- Chan K. Rifampicin concentrations in cerebrospinal fluid and plasma of the rabbit by high performance liquid chromatography. Meth Find Exptl Clin Pharmacol 1986; 8: 721-6.
- 190. Cavenaghi R. Rifampicin raw material characteristics and their effect on bioavailability. Bull Int Union Tuberc Lung Dis 1989; 64: 36-41.
- 191. Blomberg B, Spinaci S, Fourie B, Laing R. The rationale for recommending fixed-dose combination tablets for treatment of tuberculosis. Bull World Health Organ 2001; 79: 61-8.
- 192. Pelizza G, Nebuloni M, Ferrari P, Gallo GG. Polymorphism of rifampicin. Farmaco Ed Sci 1977; 32: 471-81.
- 193. Henwood SQ, de Villiers MM, Liebenberg W, Lötter AP. Solubility and dissolution properties of generic rifampicin raw materials. Drug Dev Industr Pharm 2000; 26: 403-8.
- 194. Girling DJ, Hitze KL. Adverse reactions to rifampicin. Bull World Health Organ 1979; 57: 45-9.
- 195. Curci G, Bergamini N, Delli Veneri F, Ninni A, Ninni V. Sul comportamento della cinetica della rifampicina e del tasso bilirubinemico dopo somministrazione isolata of ripetuta di differenti dosi per kg di peso corporeo nell'uomo. Arch Monaldi 1970; 25: 427-40.
- 196. McColl KEL, Thompson GG, El Omar E, Moore MR, Park BK, Brodie MJ. Effect of rifampicin on haem and bilirubin metabolism in man. Br J Pharmacol 1987; 23: 553-9.
- 197. Sarma GR, Immanuel C, Kailasam S, Narayana ASL, Venkatesan P. Rifampininduced release of hydrazine from isoniazid. A possible cause of hepatitis during treatment of tuberculosis with regimens containing isoniazid and rifampin. Am Rev Respir Dis 1986; 133: 1072-5.
- 198. Steele MA, Burk RF, DesPrez RM. Toxic hepatitis with isoniazid and rifampin. A meta-analysis. Chest 1991; 99: 465-71.

- 199. Burke M, Logan J. Hepatic dysfunction in tuberculous patients treated with rifampicin and isoniazid. J Irish Medical Ass 1979; 72: 430-4.
- Schaberg T, Rebhan K, Lode H. Risk factors for side-effects of isoniazid, rifampin and pyrazinamide in patients hospitalized for pulmonary tuberculosis. Eur Respir J 1996; 9: 2026-30.
- 201. Bistritzer T, Barzilay Z, Jonas A. Isoniazid-rifampin-induced fulminant liver disease in an infant. J Pediatr 1980; 97: 480-2.
- 202. Tsagaropoulou-Stinga H, Mataki-Emmanouilidou T, Karida-Kavalioti S, Manios S. Hepatotoxic reactions in children with severe tuberculosis treated with isoniazidrifampin. Pediatr Infect Dis 1985; 4: 270-3.
- 203. Van Aalderen WM, Knoester H, Knol K. Fulminant hepatitis during treatment with rifampicin, pyrazinamide and ethambutol. Eur J Pediatr 1987; 146: 290-1.
- 204. Ozick LA, Jacob L, Comer GM, Lee TP, Ben-Zvi J, Donelson SS, Felton CP. Hepatotoxicity from isoniazid and rifampin in inner-city AIDS patients. Am J Gastroenterol 1995; 90: 1978-80.
- 205. de A Nishioka S. Antituberculosis drugs and hepatotoxicity. (Correspondence). Am J Gastroenterol 1996; 91: 1471.
- 206. Ungo JR, Jones D, Ashkin D, Hollender ES, Bernstein D, Albanese AP, Pitchenik AE. Antituberculosis drug-induced hepatotoxicity. The role of hepatitis C virus and the human immunodeficiency virus. Am J Respir Crit Care Med 1998; 157: 1871-6.
- 207. Hwang SJ, Wu JC, Lee CH, Yen FS, Lu CL, Lin TP, Lee SD. A prospective clinical study of isoniazid-rifampicin-pyrazinamide-induced liver injury in an area endemic for hepatitis B. J Gastroenterol Hepatol 1997; 12: 87-91.
- 208. Wu JC, Lee SD, Yeh PF, Chan CY, Wang YJ, Huang YS, Tsai YT, Lee PY, Ting LP, Lo KW. Isoniazid-rifampin-induced hepatitis in hepatitis C carriers. Gastroenterology 1990; 98: 502-4.
- 209. Katz MD, Lor E. Acute interstitial nephritis associated with intermittent rifampin use. Drug Intell Clin Pharm 1986; 20: 789-92.
- Murray AN, Cassidy MJD, Templecamp C. Rapidly progressive glomerulonephritis associated with rifampicin therapy for pulmonary tuberculosis. Nephron 1987; 46: 373-6.
- Walker-Renard P. Pruritus associated with intravenous rifampin. Am Rev Respir Dis 1991; 144: 750-5.
- 212. Goldin HM, Schweitzer WJ, Bronson DM. Rifampin and exfoliative dermatitis. (Correspondence). Ann Intern Med 1987; 107: 789.
- 213. Okano M, Kitano Y, Igarashi T. Toxic epidermal necrolysis due to rifampicin. (Correspondence). J Am Acad Dermatol 1987; 17: 303-4.
- 214. Nyirenda K, Gill GV. Stevens-Johnson syndrome due to rifampicin. (Correspondence). BMJ 1977; 2: 1189.

- 215. Prazuck T, Fisch A, Simonnet F, Noat G. Lyell's syndrome associated with rifampicin therapy of tuberculosis in an AIDS patient. (Correspondence). Scand J Infect Dis 1990; 22: 629.
- 216. Kuaban C, Bercion R, Koulla-Shiro S. Current HIV seroprevalence rate and incidence of adverse skin reactions in adults with pulmonary tuberculosis receiving thiacetazone-free antituberculosis treatment in Yaounde, Cameroon. Centr Afric J Med 1998; 44: 34-7.
- 217. Arora VK, Bedi RS, Arora R. Rifampicin induced menstrual disturbances. Ind J Chest Dis All Sci 1987; 29: 63-4.
- 218. Wurtz RM, Abrams D, Becker S, Jacobson MA, Mass MM, Marks SH. Anaphylactoid drug reactions to ciprofloxacin and rifampicin in HIV-infected patients. (Correspondence). Lancet 1989; 1: 955-6.
- 219. Martínez E, Collazos J, Mayo J. Shock and cerebral infarct after rifampin reexposure in a patient infected with human immunodeficiency virus. Clin Infect Dis 1998; 27: 1329-30.
- 220. Van Assendelft AHW. Leucopenia in rifampicin chemotherapy. J Antimicrob Chemother 1985; 16: 407-8.
- 221. Conen D, Blumberg A, Weber S, Schubothe H. Hämolytische Krise und akutes Nierenversagen unter Rifampicin. Schweiz Med Wochenschr 1979; 109: 558-62.
- 222. Kindelan JM, Serrano I, Jurado R, Villanueva JL, Garcia-Lazaro M, Garcia-Herola A, Torre Cisneros J. Rifampin-induced severe thrombocytopenia in a patient with pulmonary tuberculosis. (Correspondence). Ann Pharmacother 1994; 28: 1304-5.
- 223. Bachs L, Parés A, Piera C, Elena M, Rodés J. Comparison of rifampicin with phenobarbitone for treatment of pruritus in biliary cirrhosis. (Correspondence). Lancet 1989; 1: 574-6.
- 224. Klaui H, Leuenberger P. Pseudomembranous colitis due to rifampicin. (Correspondence). Lancet 1981; 2: 1294.
- 225. Nakajima A, Yajima S, Shirakura T, Ito T, Kataoka Y, Ueda K, Nagoshi D, Kanemoto H, Matsubashi N. Rifampicin-associated pseudomembranous colitis. J Gastroenterol 2000; 35: 299-303.
- 226. Lange P, Oun H, Fuller S, Turney JH. Eosinophilic colitis due to rifampicin. (Correspondence). Lancet 1994; 344: 1296-7.
- 227. Berning SE, Iseman MD. Rifamycin-induced lupus syndrome. Lancet 1997; 349: 1521-2.
- 228. Jenkins P, Emerson PA. Myopathy induced by rifampicin. BMJ 1981; 283: 105-6.
- 229. Dutt AK, Moers D, Stead WW. Undesirable side effects of isoniazid and rifampin in largely twice-weekly short-course chemotherapy for tuberculosis. Am Rev Respir Dis 1983; 128: 419-24.

- 230. Morgan JR, Clarke KW, Brear SG. Phenomenon of rifampicin-induced discolouration of body fluids. (Correspondence). Respir Med 1993; 84: 320-1.
- 231. Lyons RW. Orange contact lenses from rifampin. (Correspondence). N Engl J Med 1979; 300: 372-3.
- Poitrineau Y, Barthelemy J, Rouby D, Fauron P, Barrau P, Beligon C. Intoxication mortelle par la rifampicine. A propos d'une observation. Therapie 1993; 48: 271-3.
- 233. Holdiness MR. Rifampicin adverse cutaneous reaction. (Correspondence). Int J Dermatol 1986; 25: 72-3.
- 234. Bolan G, Laurie RE, Broome CV. Red man syndrome: inadvertent administration of an excessive dose of rifampin to children in a day-care center. Pediatrics 1986; 77:633-5.
- 235. Damkier P, Hansen LL, Brøsen K. Rifampicin treatment greatly increases the apparent oral clearance of quinidine. Pharmacol Toxicol 1999; 85: 257-62.
- 236. Raghupati Sarma G, Acharyulu GS, Kannapiran M, Krishna Murthy PV, Gurumurthy P, Tripathy SP. Role of rifampicin in arthralgia induced by pyrazinamide. Tubercle 1983; 64: 93-100.
- 237. Baciewicz AM, Self TH. Rifampin drug interactions. Arch Intern Med 1984; 144: 1667-71.
- 238. Acocella G, Conti R. Interaction of rifampicin with other drugs. Tubercle 1980; 61: 171-7.
- 239. Baciewicz AM, Self TH, Bekemeyer WB. Update on rifampin drug interactions. Arch Intern Med 1987; 147: 565-8.
- Strayhorn VA, Baciewicz AM, Self TH. Update on rifampin drug interactions, III. Arch Intern Med 1997; 157: 2453-8.
- 241. Bhatia RS, Uppal R, Malhi R, Behera D, Jindal SK. Drug interaction between rifampicin and cotrimoxazole in patients with tuberculosis. Hum Exp Toxicol 1991; 10: 419-21.
- 242. Jaruratanasirikul S, Sriwiriyajan S. Effect of indinavir on the pharmacokinetics of rifampicin in HIV-infected patients. Pharm Pharmacol 2001; 53: 409-12.
- 243. Twum-Barima Y, Carruthers SG. Quinidine-rifampin interaction. N Engl J Med 1981; 304: 1466-9.
- 244. Bussey HL, Merritt GJ, Hill EG. The influence of rifampin on quinidine and digoxin. Arch Intern Med 1984; 144: 1021-3.
- 245. Mauro VF, Somani P, Temesy-Armos PN. Drug interaction between lorcainide and rifampicin. Eur J Clin Pharmacol 1987; 31: 737-8.
- 246. Hauser AR, Lee C, Teague RB, Mullins C. The effect of rifampin on theophylline disposition. (Abstract). Clin Pharmacol Ther 1983; 33: 254.
- 247. Straughn AB, Henderson RP, Lieberman PL, Self TH. Effect of rifampin on theophylline disposition. Therapeutic Drug Monitoring 1984; 6: 153-6.

- 248. Powell-Jackson PR, Jamieson AP, Gray BJ, Moxham J, Williams R. Effect of rifampicin administration on theophylline pharmacokinetics in humans. Am Rev Respir Dis 1985; 131: 939-40.
- 249. Michot F, Bürgi M, Büttner J. Rimactan (Rifampizin) und Antikoagulantientherapie. Schweiz Med Wochenschr 1970; 100: 583-4.
- 250. Beran G. Der Einfluss der Rifampizintherapie auf die orale Antikoagulation mit Acenoumarol. Prax Pneumol 1970; 26: 350-3.
- 251. Broekhout-Mussert RJ, Bieger R, van Brummelen P, Lemkes HHPJ. Inhibition by rifampin of the anticoagulant effect of phenprocoumon. JAMA 1974; 229: 1903-4.
- 252. Held H. Interaktion von Rifampicin mit Phenprocoumaron. Beobachtungen bei tuberkulosekranken Patienten. Dtsch Med Wschr 1979; 104: 1311-4.
- 253. O'Reilly RA. Interaction of sodium warfarin and rifampin. Studies in man. Ann Intern Med 1974; 81: 337-40.
- 254. O'Reilly RA. Interaction of chronic daily warfarin therapy and rifampin. Ann Intern Med 1975; 83: 506-7.
- 255. Romankiewicz JA, Ehrman M. Rifampin and warfarin: a drug interaction. Ann Intern Med 1975; 82: 224-5.
- 256. Self TH, Mann RB. Interaction of rifampin and warfarin. Chest 1975; 67: 490-1.
- 257. Fox P. Warfarin-rifampicin interaction. (Correspondence). Med J Austr 1982; 1: 60.
- 258. Syvälahti EKG, Pihlajamäki KK, Iisalo EJ. Rifampicin and drug metabolism. (Correspondence). Lancet 1974; 2: 232-3.
- 259. Zilly W, Breimer DD, Richter E. Induction of drug metabolism in man after rifampicin treatment measured by increased hexobarbital and tolbutamide clearance. Eur J Clin Pharmacol 1975; 9: 219-27.
- 260. Kihara Y, Otsuki M. Interaction of gliazide and rifampicin. (Correspondence). Diabetes Care 2000; 23: 1204-5.
- Niemi M, Kivistö KT, Backman JT, Neuvonen PJ. Effect of rifampicin on the pharmacokinetics and pharmacodynamics of glimepiride. J Clin Pharmacol 2000; 50: 591-5.
- 262. Niemi M, Backman JT, Neuvonen M, Neuvonen PJ, Kivistö KT. Effects of rifampin on the pharmacokinetics and pharmacodynamics of glyburide and glipizide. Clin Pharmacol Ther 2001; 69: 400-6.
- 263. Lazar JD, Wilner KD. Drug interactions with fluconazole. Rev Infect Dis 1990; 12 (Suppl 3): S327-S333.
- 264. Harvey CJ, Lloyd ME, Bateman NT, Hughes GRV. Influence of rifampicin on hydroxychloroquine. (Correspondence). Clin Experiment Rheumatol 1995; 13: 536.
- 265. Osborn JE, Pettit MJ, Graham P. Interaction between rifampicin and quinine: case report. (Correspondence). Pharm J 1989; 243: 704.

- 266. Ridtitid W, Wongnawa M, Mahatthanatrakul W, Chaipol P, Sunbhanich M. Effect of rifampicin on plasma concentrations of mefloquine in healthy volunteers. J Pharm Pharmacol 2001; 52: 1265-9.
- Prober CG. Effect of rifampin on chloramphenicol levels. (Correspondence). N Engl J Med 1985; 312: 788-9.
- 268. Centers for Disease Control and Prevention. Clinical update: impact of HIV protease inhibitors on the treatment of HIV-infected tuberculosis patients with rifampin. Morb Mortal Wkly Rep 1996; 45: 921-5.
- 269. Burman WJ, Gallicano K, Peloquin C. Therapeutic implications of drug interactions in the treatment of human immunodeficiency virus-related tuberculosis. Clin Infect Dis 1999; 28: 419-30.
- 270. Dean GL, Back DJ, de Ruiter A. Effect of tuberculosis therapy on nevirapine trough plasma concentrations. (Correspondence). AIDS 1999; 13: 2489-90.
- 271. Burger DM, Meenhorst PL, Koks CHW, Beijnen JH. Pharmacokinetic interaction between rifampin and zidovudine. Antimicrob Agents Chemother 1993; 37: 1426-31.
- 272. Burger DM, Meenhorst PL, ten Napel CHH, Mulder JW, Neef C, Koks CHW, Bult A, Beijnen JH. Pharmacokinetic variability in HIV-infected individuals: subgroup analysis and drug interactions. AIDS 1994; 8: 1683-9.
- 273. Ohnhaus EE, Brockmeyer N, Dylewicz P, Habicht H. The effect of antipyrine and rifampin on the metabolism of diazepam. Clin Pharmacol Ther 1987; 42: 148-56.
- 274. Herman RJ, Nakamura K, Wilkinson GR, Wood AJJ. Induction of propanolol metabolism by rifampicin. Br J Clin Pharmac 1983; 16: 565-9.
- 275. Rahn KH, Mooy J, Böhm R, van der Vet A. Reduction of bioavailability of verapamil by rifampicin. (Correspondence). N Engl J Med 1985; 312: 920-1.
- 276. Barbarash RA. Verapamil-rifampin interaction. Drug Intell Clin Pharm 1985; 19: 559-60.
- 277. Mooy J, Böhm R, van Baak M, van Kemenade J, van der Vet A, Rahm KH. The influence of antituberculosis drugs on the plasma level of verapamil. Eur J Clin Pharmacol 1987; 32: 107-9.
- 278. Tada Y, Tsuda Y, Otsuka T, Nagasawa K, Kimbura H, Kusaba T, Sakata T. Case report: nifedipine-rifampicin interaction attenuates effect on blood pressure in a patients with essential hypertension. Am J Med Sci 1992; 303: 25-7.
- 279. Novi C, Bissoli F, Simonati V, Volpini T, Baroli A, Vignati G. Rifampin and digoxin: possible interaction in a dialysis patient. (Correspondence). JAMA 1980; 244: 2521.
- 280. Gault H, Longerich L, Dawe M, Fine A. Digoxin-rifampin interaction. Clin Pharmacol Ther 1984; 35: 750-4.

- 281. LeBel M, Masson E, Guilbert E, Colborn D, Pacquet F, Allard S, Vallée F, Narang PK. Effects of rifabutin and rifampicin on the pharmacokinetics of ethinylestradiol and norethindrone. J Clin Pharmacol 1998; 38: 1042-50.
- 282. Udwadia ZW, Sridhar G, Beveridge CJ, Soutar C, McHardy GJR, Leitch AG. Catastrophic deterioration in asthma induced by rifampicin in steroid-dependent asthma. Respir Med 1993; 87: 629.
- 283. Powell-Jackson PR, Gray BJ, Heaton RW, Costello JF, Williams R, English J. Adverse effect of rifampicin administration on steroid-dependent asthma. Am Rev Respir Dis 1983; 128: 307-10.
- 284. Atkin SL, Masson EA, Bodmer CW, Walker BA, White MC. Increased insulin requirement in a patient with type 1 diabetes on rifampicin. (Correspondence). Diabetic Medicine 1993; 10: 392.
- 285. Takasu N, Yamada T, Miura H, Sakamoto S, Korenaga M, Nakajima K, Kanayama M. Rifampicin-induced early phase hyperglycemia in humans. Am Rev Respir Dis 1982; 125: 23-7.
- 286. Nolan SR, Self TH, Norwood JM. Interaction between rifampin and levothyroxine. Southern Med J 1999; 92: 529-31.
- 287. Van Buren D, Wideman CA, Gibbons S, Van Buren CT, Jarowenko M, Flechner SM, Frazier OH, Cooley DA, Kahan BD. The antagonistic effect of rifampin upon cyclosporine bioavailability. Transplant Proc 1984; 16: 1642-5.
- 288. Daniels NJ, Dover JS, Schachter RK. Interaction between cyclosporin and rifampicin. (Correspondence). Lancet 1984; 2: 639.
- 289. Coward RA, Raftery AT, Brown CB. Cyclosporin and antituberculous therapy. (Correspondence). Lancet 1985; 1: 1342-3.
- 290. Freitag VL, Skifton RD, Lake KD. Effect of short-term rifampin on stable cyclosporine concentrations. (Correspondence). Ann Pharmacother 1999; 33: 871-2.
- 291. Kiuchi T, Tanaka K, Inomata Y, Uemoto S, Satomura K, Egawa H, Uyama S, Sano K, Okajima H, Yamaoka Y. Experience with tacrolimus-based immunosuppression in living-related liver transplantation complicated with graft tuberculosis: interaction with rifampicin and side effects. Transplant Proc 1996; 28: 3171-2.
- 292. Kreek MJ, Garfield JW, Gutjahr CL, Giusti LM. Rifampin-induced methadone withdrawal. N Engl J Med 1976; 294: 1104-6.
- 293. Raistrick D, Hay A, Wolff K. Methadone maintenance and tuberculosis treatment. BMJ 1996; 313: 925-6.
- 294. Schlatter J, Madras JL, Saulnier JL, Poujade F. Interactions médicamenteuses avec la méthadone. Presse Méd 1999; 28: 1381-4.
- 295. Chouraqui JP, Bessard G, Favier M, Kolodie L, Rambaud P. Hémorrhagie par avitaminose K chez la femme enceinte et le nouveau-né. Thérapie 1982; 37: 447-50.

- 296. Brodie MJ, Boobis AR, Hillyard CI, Abeyasekera G, Stevenson JC, MacIntyre I, Park K. Effect of rifampicin and isoniazid on vitamin D metabolism. Clin Pharmacol Ther 1982; 32: 525-30.
- 297. Shafer JL, Houston JB. The effect of rifampicin on sulfapyridine plasma concentrations following sulphalazine administration. Br J Clin Pharmac 1992; 19: 526-8.
- 298. Kushner S, Dalalian H, Sanjuro JL, Bach FL, Jr, Safir SR, Smith VK, Jr, Williams JH. Experimental chemotherapy of tuberculosis. II. The synthesis of pyrazinamides and related compounds. J Am Chem Soc 1952; 74: 3617-21.
- 299. Solotorovsky M, Gregory FJ, Isonson EJ, Bugie EJ, O'Neill RC, Pfister K 3rd. Pyrazinoic acid amide - an agent active against experimental murine tuberculosis. (19447). Proc Soc Experiment Biol Med 1952; 79: 563-55.
- 300. Dalmer O, Walter E. Firma E. Merck in Darmstadt. Verfahren zur Herstellung von Abkömmlingen der Pyrazinmonocarbonsäure. Germany patent 632 257 Klasse 12 p Gruppe 6 M 127990 IV a/12 p. 1934.
- Zierski M. Pharmakologie, Toxikologie und klinische Anwendung von Pyrazinamid. Praxis Klin Pneumol 1981; 35: 1075-105.
- 302. McDermott W, Tompsett R, Stern K. Activation of pyrazinamide and nicotinamide in acidic environments in vitro. Am Rev Tuberc 1954; 70: 748-54.
- 303. Zhang Y, Scorpio A, Nikaido H, Sun Z. Role of acid pH and deficient efflux of pyrazinoic acid in unique susceptibility of *Mycobacterium tuberculosis* to pyrazinamide. J Bacteriol 1999; 181: 2044-9.
- Rastogi N, Potar MC, David HL. Pyrazinamide is not effective against intracellularly growing *Mycobacterium tuberculosis*. Antimicrob Agents Chemother 1988; 32: 287.
- 305. Salfinger M, Crowle AJ, Barth Reller L. Pyrazinamide and pyrazinoic acid activity against tubercle bacilli in cultured human macrophages and in the BACTEC system. J Infect Dis 1990; 162: 201-7.
- 306. Raynaud C, Lanéelle MA, Senaratne RH, Draper P, Lanéelle G, Daffé M. Mechanisms of pyrazinamide resistance in mycobacteria: importance of lack of uptake in addition to lack of pyrazinamidase activity. Microbiology 1999; 145: 1359-67.
- 307. Cole ST, Telenti A. Drug resistance in *Mycobacterium tuberculosis*. Eur Respir J 1995; 8(suppl 20): 701s-13s.
- 308. Scorpio A, Zhang Y. Mutations in *pncA*, a gene encoding pyrazinamidase / nicotinamidase, cause resistance to the antituberculous drug pyrazinamide in tuber-cle bacillus. Nature Med 1996; 2: 662-7.
- 309. Mestdagh M, Fonteyne PA, Realini L, Rossau R, Jannes G, Mijs W, De Smet KAL, Portaels F, Van den Eeckhout E. Relationship between pyrazinamide, loss of pyrazinamidase activity, and mutations in the *pncA* locus in multidrug-resistant clinical isolates of *Mycobacterium tuberculosis*. Antimicrob Agents Chemother 1999; 43: 2317-9.

- 310. Yeager RL, Munroe WGC, Dessau FI. Pyrazinamide (Aldinamide) in the treatment of pulmonary tuberculosis. Am Rev Tuberc 1952; 65: 523-46.
- 311. Kataria YP. Observations on human infection with *Mycobacterium bovis*. Tubercle 1969; 50: 14-21.
- 312. Ellard GA, Humphries MJ, Gabriel M, Theoh R. Penetration of pyrazinamide into the cerebrospinal fluid in tuberculous meningitis. BMJ 1987; 294: 284-5.
- 313. Donald PR, Seifart H. Cerebrospinal fluid pyrazinamide concentrations in children with tuberculous meningitis. Pediatr Infect Dis 1988; 7: 469-71.
- 314. Ellard GA. Absorption, metabolism and excretion of pyrazinamide in man. Tubercle 1969; 50: 144-58.
- 315. Peloquin CA, Bulpitt AE, Jaresko GS, Jelliffe RW, James GT, Nix DE. Pharmacokinetics of pyrazinamide under fasting conditions, with food, and with antacids. Pharmacotherapy 1998; 18: 1205-11.
- 316. Schwartz WS, Moyer RE. The chemotherapy of pulmonary tuberculosis with pyrazinamide used alone and in combination with streptomycin, para-aminosalicylic acid, or isoniazid. Am Rev Tuberc 1954; 70: 413-23.
- 317. McDermott W, Ormond L, Muschenheim C, Deuschle K, McCune RM, Jr, Tompsett R. Pyrazinamide-isoniazid in tuberculosis. Am Rev Tuberc 1954; 69: 319-33.
- 318. Ferebee SH, Mount FW. Chemotherapy of tuberculosis, progress and promise. Publ Health Rep 1957; 72: 412-20.
- 319. Mount FW, Wunderlich GS, Murray SJ, Ferebee SH. Hepatic toxicity of pyrazinamide used with isoniazid in tuberculous patients. A United States Public Health Service Tuberculosis Therapy Trial. Am Rev Respir Dis 1959; 80: 371-87.
- 320. van der Kooi K, Mottet JJ, Regamey C. Isoniazid is not always the cause of hepatitis during treatment of tuberculosis. (Correspondence). Clin Infect Dis 1994; 19: 988-9.
- 321. Türktas H, Ünsal M, Tülek N, Örüç O. Hepatotoxicity of antituberculosis therapy (rifampicin, isoniazid and pyrazinamide) or viral hepatitis. Tubercle Lung Dis 1994; 75: 58-60.
- 322. Thompson NP, Caplin ME, Hamilton MI, Gillespie SH, Clarke SW, Burroughs AK, McIntyre N. Anti-tuberculosis medications and the liver: dangers and recommendations in management. Eur Respir J 1995; 8: 1384-8.
- 323. Ormerod LP, Horsfield N. Frequency and type of reactions to antituberculosis drugs: observations in routine treatment. Tubercle Lung Dis 1996; 77: 37-42.
- 324. Philipps S. Pyrazinamide-isoniazid: its apparent influence on the reactivation rate in pulmonary tuberculosis. Am Rev Respir Dis 1967; 95: 503-5.
- 325. Jenner PJ, Ellard GA, Allan WGL, Singh D, Girling DJ, Nunn AJ. Serum uric acid concentrations and arthralgia among patients treated with pyrazinamide-containing regimens in Hong Kong and Singapore. Tubercle 1981; 62: 175-9.

- 326. Kannapiran M, Krishnamurthy PV, Raghupatti Sarma G. Uric acid disposition during intermittent chemotherapy of pulmonary tuberculosis with regimens containing pyrazinamide and rifampicin. Indian J Med Res 1985; 82: 116-21.
- 327. Ellard GA, Haslam RM. Observations on the reduction of the renal elimination of urate in man caused by the administration of pyrazinamide. Tubercle 1976; 57: 97-103.
- 328. Radal M, Jonville-Béra AP, Van-Egroo C, Carré P, Lemarié E, Autret E. Eruption après la première prise d'une chimiothérapie standard antituberculeuse. Penser au pyrazinamide. Rev Mal Resp 1998; 15: 305-6.
- 329. Olivier C, Radal M, Mazaud S, Jonville-Béra AP, Martel C, Autret E. Eruption après une première prise d'une quadrithérapie antituberculeuse: penser au pyrazinamide. Arch Pédiatr 1998; 5: 289-90.
- Layer P, Engelhard M. Tuberkulostatika-induzierter systemischer Lupus erythematodes. Dtsch Med Wschr 1986; 111: 1603-5.
- 331. Herlevsen P, Nielsen C, Thuesen Pedersen J. Convulsions after treatment with pyrazinamide. Tubercle 1987; 68: 145-6.
- 332. Choohakarn C, Janma J. Pyrazinamide-induced lichenoid photodermatitis. (Correspondence). J Am Acad Dermatol 1999; 40: 645-6.
- 333. Lacroix C, Guyonnaud C, Chaou M, Duwoos H, Lafont O. Interaction between allopurinol and pyrazinamide. Eur Respir J 1988; 1: 807-11.
- 334. Peloquin CA, Nitta AT, Burman WJ, Brudney KF, Miranda-Massari JR, McGuiness ME, Berning SE, Gerena G. Low antituberculosis drug concentrations in patients with AIDS. Ann Pharmacother 1996; 30: 919-25.
- 335. Fox IH, Stein HB, Gershon SL. Effects of vitamins on the renal handling of uric acid. Adv Exp Med Biol 1977; 76B: 30-5.
- Manuel MA, Steele TH. Pyrazinamide suppression of the uricosuric response to sodium chloride infusion. J Lab Clin Med 1967; 83: 417-27.
- 337. Wilkinson RG, Shepherd RG, Thomas JP, Baughn C. Stereospecificity in a new type of synthetic antituberculous agent. (Correspondence). J Am Chem Soc 1961; 83: 2212-3.
- 338. Thomas JP, Baughn CO, Wilkinson RG, Shepherd RG. A new synthetic compound with antituberclous activity in mice: ethambutol (dextro-2,2'-(ethylenediimino)-di-1-butanol). Am Rev Respir Dis 1961; 83: 891-3.
- Karlson AG. Therapeutic effect of ethambutol (dextro-2,2'-[ethylene-diimino]di-1-butanol) on experimental tuberculosis in guinea pigs. Am Rev Respir Dis 1961; 84: 902-4.
- 340. Karlson AG. The in vitro activity of ethambutol (dextro-2,2'-[ethylene-diimino]di-1-butanol) against tubercle bacilli and other microorganisms. Am Rev Respir Dis 1961; 84: 905-6.

- 341. Crowle AJ, Sbarbaro JA, Judson FN, May MH. The effect of ethambutol on tubercle bacilli within cultured human macrophages. Am Rev Respir Dis 1985; 132: 742-5.
- 342. Takayama K, Kilburn JO. Inhibition of synthesis of arabinogalactan by ethambutol in *Mycobacterium smegmatis*. Antimicrob Agents Chemother 1989; 33: 1493-9.
- 343. Deng L, Mikusová K, Robuck KG, Scherman M, Brennan PJ, McNeil MR. Recognition of multiple effects of ethambutol on metabolism of mycobacterial cell envelope. Antimicrob Agents Chemother 1995; 39: 694-701.
- 344. Mikusová K, Slayden RA, Besra GS, Brennan PJ. Biogenesis of the mycobacterial cell wall and the site of action of ethambutol. Antimicrob Agents Chemother 1995; 39: 2484-9.
- 345. David HL, Laszlo A, Rastogi N. Mode of action of antimycobacterial drugs. Acta Leprologica 1989; 7 (Suppl 1): 189-94.
- 346. Kilburn JO, Greenberg J. Effect of ethambutol on the viable cell count in *Mycobacterium smegmatis*. Antimicrob Agents Chemother 1977; 11: 534-40.
- 347. Hoffner SE, Svenson SB, Källenius G. Synergistic effects of antimycobacterial drug combinations on *Mycobacterium avium* complex determined radiometrically in liquid medium. Eur J Clin Microbiol 1987; 6: 530-5.
- 348. Rastogi N, David HL. Mode of action of antituberculous drugs and mechanisms of drug resistance in *Mycobacterium tuberculosis*. Res Microbiol 1993; 144: 133-43.
- 349. Place VA, Thomas JP. Clinical pharmacology of ethambutol. Am Rev Respir Dis 1963; 87:901-4.
- 350. Peets EA, Sweeney WM, Place VA, Buyske DA. The absorption, excretion, and metabolic fate of ethambutol in man. Am Rev Respir Dis 1965; 91: 51-8.
- 351. Peloquin CA. Pharmacology of the antimycobacterial drugs. Med Clin N Amer 1993; 77: 1253-62.
- 352. Kelly RG, Kaleita E, Eisner HJ. Tissue distribution of (¹⁴C)Ethambutol in mice. Am Rev Respir Dis 1981; 123: 689-90.
- 353. Liss RH, Letourneau J, Schepis JP. Distribution of ethambutol in primate tissues and cells. Am Rev Respir Dis 1981; 123: 529-32.
- 354. Varughese A, Brater DC, Benet LZ, Lee CSC. Ethambutol kinetics in patients with impaired renal function. Am Rev Respir Dis 1986; 134: 34-8.
- 355. Peloquin CA, Bulpitt AE, Jaresko GSJ, Jelliffe RW, Childs JM, Nix DE. Pharmacokinetics of ethambutol under fasting conditions, with food, and with antacids. Antimicrob Agents Chemother 1999; 43: 568-72.
- 356. American Thoracic Society, Centers for Disease Control, American Academy of Pediatrics. Treatment of tuberculosis and tuberculosis infection in adults and children. Am J Respir Crit Care Med 1994; 149: 1359-74.

- 357. Carr DE, Henkind P. Ocular manifestations of ethambutol. Toxic amblyopia after administration of an experimental antituberculosis drug. Arch Ophthalmol 1962; 55: 566-71.
- 358. Barron GJ, Tepper L, Iovine G. Ocular toxicity from ethambutol. Am J Ophthalmol 1974; 77: 256-60.
- 359. Tiburtius H. The undesired side-effects of myambutol. Antibiotica et Chemotherapia 1970; 16: 298-301.
- 360. Seth V, Khosla PK, Semwal OP, D'Monty V. Visual evoked responses in tuberculous children on ethambutol therapy. Ind Pediatr 1991; 28: 713-7.
- 361. Kahana LM. Ethambutol in tuberculosis. Biomed Pharmacother 1990; 44: 21-3.
- 362. Joubert PH, Strobele JG, Ogle CW, Van der Merwe CA. Subclinical impairment of colour vision in patients receiving ethambutol. Br J Clin Pharmac 1986; 21: 213-6.
- 363. Citron KM, Thomas GO. Ocular toxicity from ethambutol. (Editorial). Thorax 1986; 41: 737-9.
- 364. Salmon JF, Carmichael TR, Welsh NH. Use of contrast sensitivity measurement in the detection of subclinical ethambutol toxic optic neuropathy. Br J Ophthalmology 1987; 71: 192-6.
- 365. Polak BCP, Leys M, van Lith GHM. Blue-yellow colour vision changes as early symptoms of ethambutol oculotoxicity. Ophthalmologica Basel 1985; 191: 223-6.
- 366. Kahana LM. Ethambutol and the eye. (Correspondence). Lancet 1988; 2: 627-8.
- 367. Campbell IA, Ormerod LP. Ethambutol and the eye. (Correspondence). Lancet 1988; 2: 113-4.
- 368. Pau H. Myambutol (Ethambutol) bedingte Retinoneuritis. Klin Mbl Augenheilk 1985; 187: 25-9.
- Chatterjee VKK, Buchanan DR, Friedmann AI, Green M. Ocular toxicity following ethambutol in standard dosage. Br J Chest 1986; 80: 288-91.
- Russo PA, Chaglasian MA. Toxic optic neuropathy associated with ethambutol: implications for current therapy. J Am Optometric Ass 1994; 65: 332-8.
- 371. Kahana LM. Toxic ocular effects of ethambutol. Can Med Ass J 1987; 137: 213-6.
- 372. Leibold JE. The ocular toxicity of ethambutol and its relation to dose. Ann NY Acad Sci 1966; 135: 904-9.
- 373. Alvarez KL, Krop LC. Ethambutol-induced ocular toxicity revisited. (Correspondence). Ann Pharmacother 1993; 27: 102-3.
- 374. Vérin P, Pesme D, Yacoubi M, Morax S. Toxicité oculaire de l'ethambutol. Arch Ophthalmol 1971; 31: 669-86.
- 375. Woung LC, Jou JR, Liaw SL. Visual function in recovered ethambutol optic neuropathy. J Ocular Pharmacol Therap 1995; 11: 411-9.

- 376. Graham SM, Daley HM, Salaniponi FM, Harries AD. Ethambutol in tuberculosis: time to reconsider? Arch Dis Child 1998; 79: 274-8.
- 377. Cole A, May PM, Williams DR. Metal binding by pharmaceuticals. Part 1. Copper(II) and zinc(II) interactions following ethambutol administration. Agents Actions 1981; 11: 296-305.
- 378. Kozak SF, Inderlied CB, Hsu HY, Heller KB, Sadun AA. The role of copper on ethambutol's antimicrobial action and implications for ethambutol-induced optic neuropathy. Diagn Microbiol Infect Dis 1998; 30: 83-7.
- 379. Roberts SM. A review of the papers on the ocular toxicity of ethambutol hydrochloride myambutol, an anti-tuberculosis drug. Am J Optom Physiol Optics 1974; 51: 987-92.
- 380. Trébucq A. Should ethambutol be recommended for routine treatment of tuberculosis in children? A review of the literature. Int J Tuberc Lung Dis 1997; 1: 12-5.
- 381. Wong PC, Yew WW, Wong CF, Choi HY. Ethambutol-induced pulmonary infiltrates with eosinophilia and skin involvement. Eur Respir J 1995; 8: 866-8.
- 382. Dhamgaye T, Mohanty KC. Hypersensitivity to multiple drugs streptomycin, rifampicin and ethambutol: an unusual presentation. (Correspondence). Tubercle Lung Dis 1995; 76: 181.
- Rabinovitz M, Pitlik SD, Halevy J, Rosenfeld JB. Ethambutol-induced thrombocytopenia. Chest 2000; 81: 765-6.
- 384. Postlethwaite AE, Bartel AG, Kelley WN. Hyperuricemia due to ethambutol. N Engl J Med 1972; 286: 761-2.
- 385. Waksman SA, Curtis RE. The actinomyces of the soil. Soil Sci 1916; 1: 99-134.
- 386. Waksman SA. Streptomycin: background, isolation, properties, and utilization. Nobel Foundation 1952; 287-305.
- 387. Wallgren A. Physiology or medicine 1952. Presentation speech by Professor A. Wallgren, member of the Staff of Professors of the Royal Caroline Institute. Nobel Foundation 1952; 365-9.
- Waksman SA. Historical introduction. *In:* Waksman SA, Ed. Streptomycin. Nature and practical applications. Baltimore: The Williams & Wilkins Co., 1949; 1-10.
- 389. Schatz A, Bugie E, Waksman SA. Streptomycin, a substance exhibiting antibiotic activity against gram-positive and gram-negative bacteria. Proc Soc Experiment Biol Med 1944; 55: 66-9.
- 390. Waksman SA, Bugie E, Schatz A. Isolation of antibiotic substances from soil micro-organisms, with special reference to streptothricin and streptomycin. Mayo Clin Proc 1944; 19: 537-48.
- 391. Schatz A, Waksman SA. Effect of streptomycin and other antibiotic substances upon *Mycobacterium tuberculosis* and related organisms. Proc Soc Experiment Biol Med 1944; 57: 244-8.

- 392. Feldman WH, Hinshaw HC. Effects of streptomycin on experimental tuberculosis in guinea pigs: a preliminary report. Mayo Clin Proc 1944; 19: 593-9.
- 393. Hinshaw HC, Feldman WH. Streptomycin in treatment of clinical tuberculosis: a preliminary report. Mayo Clin Proc 1945; 20: 314-8.
- Feldman WH, Hinswaw C, Mann FC. Streptomycin in experimental tuberculosis. Am Rev Tuberc 1945; 52: 269-98.
- Hinshaw HC, Feldman WH, Pfuetze KH. Treatment of tuberculosis with streptomycin. JAMA 1946; 132: 778-82.
- 396. Crowle AJ, Sbarbaro JA, Judson FN, Douvas GS, May MH. Inhibition by streptomycin of tubercle bacilli within cultures human macrophages. Am Rev Respir Dis 1984; 130: 839-44.
- 397. Moazed D, Noller HF. Interaction of antibiotics with functional sites in 16S ribosomal RNA. Nature 1987; 327: 389-94.
- 398. Mitchison DA. The segregation of streptomycin-resistant variants of Mycobacterium tuberculosis into groups with characteristic levels of resistance. J Gen Microbiol 1951; 5: 596-604.
- 399. Rinder H, Mieskes KT, Löscher T. Heteroresistance in *Mycobacterium tuberculosis*. Int J Tuberc Lung Dis 2001; 5: 339-45.
- 400. Honoré N, Cole ST. Streptomycin resistance in mycobacteria. Antimicrob Agents Chemother 1994; 38: 238-42.
- 401. Holdiness MR. Chromatographic analysis of antituberculosis drugs in biological samples. J Chromatography 1985; 340: 321-59.
- 402. Douglas JG, McLeod MJ. Pharmacokinetic factors in the modern drug treatment of tuberculosis. Clin Pharmacokinetics 1999; 37: 127-46.
- 403. Johnston RN, Smith DH, Lockhart W, Ritchie RT. Optimal dose of streptomycin in pulmonary tuberculosis. BMJ 1961; 1: 105.
- 404. Morris JT, Cooper RH. Intravenous streptomycin: a useful route of administration. Clin Infect Dis 1994; 19: 1150-1.
- 405. Johnston RN, Smith DH, Ritchie RT, Lockhart W. Prolonged streptomycin and isoniazid for pulmonary tuberculosis. BMJ 1964; 1: 1679-83.
- 406. Pfaltz CR, Herzog H, Staub H, Wey W. Zur ototoxischen Wirkung hoher Streptomycindosen. Schweiz Med Wochenschr 1960; 90: 1472-8.
- 407. Jahrmärker H. Ueber Desensiblilisierung bei Streptomycin-Allergie des Pflegepersonals. Aerztl Wochenschrift 1955; 10: 873-6.
- 408. Holdiness MR. Neurological manifestations and toxicities of the antituberculosis drugs. A review. Med Toxicol 1987; 2: 33-51.
- 409. Paradelis AG, Triantaphyllidis CJ, Mironidou M, Crassaris LG, Karachalios DN, Giala MM. Interaction of aminoglycoside antibiotics and calcium channel blockers at the neuromuscular junction. Meth Find Exptl Clin Pharmacol 1988; 10: 687-90.

- 410. Paradelis AG, Triantaphyllidis C, Giala MM. Neuromuscular blocking activity of aminoglycoside antibiotics. Meth Find Exptl Clin Pharmacol 1980; 1: 45-51.
- 411. Ohtani I, Ohtsuki K, Omata T, Ouchi J, Saito T. Potentiation and its mechansims of cochlear damage resulting from furosemide and aminoglycoside antibiotics. J Otorhinlaryngol Relat Spec 1981; 40: 53-63.
- 412. Mathog RH, Capps MJ. Ototoxic interactions of ethacrynic acid and streptomycin. Ann Otol 1977; 86: 158-63.
- 413. Steinbereithner K. Synergistische Wirkung bestimmter Antibiotika mit Muskelrelaxantien vom Curaretyp. Bull Schweiz Med Wiss 1967; 23: 57-68.
- 414. Giala MM, Paradelis AG. Two cases of prolonged respiratory depression due to interaction of pancuronium with colistin and streptomycin. (Correspondence). J Antimicrob Chemother 1979; 5: 234-5.
- 415. Burkett L, Bikhazi GB, Thomas KC, Jr, Rosenthal DA, Wirta MG, Foldes FF. Mutual potentitiation of the neuromuscular effects of antibiotics and relaxants. Anesth Analg 1979; 58: 107-15.
- 416. Trubuhovich RV. Delayed reversal of diallyl-nortroxiferine after streptomycin. (Correspondence). Br J Anaesth 1966; 38: 843-4.
- 417. Fréour P, Nacef T, Fourcaud R, Belhassime T, Kissel M. Le prothionamide dans le traitement de la tuberculose pulmonaire. New York: Proceedings of the 20th Conference of IUATLD, 1969; 29-32.
- 418. Eule H. Les thioamides in vitro et en clinique. Résistance bactérienne et résistance croisée. Leur place actuelle dans le traitement de la tuberculose. New York: Proceedings of the 20th Conference of IUATLD, 1969; 25-8.
- 419. Domagk G. Investigations on the antituberculous activity of the thiosemicarbazones in vitro and in vivo. Am Rev Tuberc 1950; 61: 8-19.
- 420. Domagk G, Behnisch R, Mietzsch F, SChmidt H. Ueber eine neue, gegen Tuberkelbazillen in vitro wirksame Verbindungsklasse. Naturwissenschaften 1946; 33: 315.
- 421. Malluche H. Die Thiosemicarbazone-Therapie der Tuberkulose. Fortschr Tuberk Forsch 1952; 5: 152-254.
- 422. Grosset J, Benhassine M. La thiacétazone (TB1): données expérimentales et cliniques récentes. Adv Tuberc Res 1970; 17: 107-53.
- 423. Thomas KL, Joseph S, Subbaiah TV, Selkon JB. Identification of tubercle bacilli from Indian patients with pulmonary tuberculosis. Bull World Health Organ 1961; 25: 747-58.
- 424. Mitchison DA, Lloyd J. Comparison of the sensitivity to thiacetazone of tubercle bacilli from patients in Britain, East Africa, South India and Hong Kong. Tubercle 1964; 45: 360-9.
- 425. Rist N. Thiacetazone sensitivity and resistance: introductory remarks. Tubercle 1968; 49 (suppl): 36-8.

- 426. Mitchison DA. Natural sensitivity of *M. tuberculosis* to thiacetazone. Tubercle 1968; 49 (suppl): 38-46.
- 427. Grosset J, Rodrigues F, Benhassine M, Chaulet P, Larbaoui D. Sensitivity to thiacetazone of strains of *Mycobacterium tuberculosis* isolated in Algiers: practical deductions. Tubercle 1968; 49 (suppl): 46-8.
- 428. Gangadharam PRJ, Devaki V, Mohan K. Thiacetazone sensitivity of Indian tubercle bacilli. Tubercle 1968; 49 (suppl): 48-51.
- 429. Hamre D, Bernstein J, Donovick R. The chemotherapy of experimental tuberculosis. II. Thiosemicarbazones and analogues in experimental tuberculosis in the mouse. J Bacteriol 1950; 59: 675-80.
- 430. Protivinsky R. Chemotherapeutics with tuberculostatic action. Antibiotics Chemother 1971; 17: 101-21.
- 431. Liebermeister K. Zur Wirkung der tuberkulostatischen Chemotherapeutika. Deutsch Med Wschr 1950; 75: 621-2.
- 432. Wernitz W, Tornus H. Quantitative Contebenstudien. IV. Mitteilung. Contebenblutspiegel beim Menschen. Zeitschr Klin Med 1952; 150: 170-6.
- 433. Heilmeyer I, Heilmeyer L. Ueber Resorption und Ausscheidung von TBI 698 (Conteben) nach peroraler Belastung. Klin Wochenschr 1949; 27: 790-1.
- 434. Jenner PJ, Ellard GA, Swai OB. A study of thiacetazone blood levels and urinary excretion in man, using high performance liquid chromatography. Lepr Rev 1984; 55: 121-8.
- 435. Hinshaw HC, McDermott W. Thiosemicarbazone therapy of tuberculosis in humans. Am Rev Tuberc 1950; 61: 145-57.
- 436. Miller AB. Thiacetazone toxicity: a general review. Tubercle 1968; 49 (suppl): 54-6.
- 437. Aquinas M. Side effects and toxicity to thiacetazone and isoniazid: findings in a Hong Kong Tuberculosis Treatment Service / British Medical Research Council investigation. Tubercle 1968; 49 (suppl): 56-8.
- 438. Aquinas M. Side effects and toxicity in the combined regimen of thiacetazone and isoniazid in Morocco. Tubercle 1968; 49 (suppl): 58-9.
- 439. Miller AB, Fox W, Tall R. An international co-operative investigation into thiacetazone (thioacetazone) side effects. Tubercle 1966; 47: 33-74.
- 440. Stühmer A. Nebenerscheinungen bei der Behandlung von Lupuskranken mit Tb I 698. Med Klin 1949; 27: 864-6.
- 441. Raviglione MC, Dinan WA, Pablos-Mendez A, Palagiano A, Sabatini MT. Fatal toxic epidermal necrolysis during prophylaxis with pyrimethamine and sulfadoxine in a human immunodeficiency virus-infected person. Arch Intern Med 1988;148: 2683-4.
- 442. Senneville E, Lecocq P, Ajana F, Chidiac C, Mouton Y. Co-trimoxazole for toxic epidermal necrolysis in AIDS. (Correspondence). Lancet 1991; 337: 919.

- 443. Nunn P, Kibuga D, Gathua S, Brindle R, Imalingat A, Wasunna K, Lucas S, Gilks C, Omwega M, Were J, McAdam K. Cutaneous hypersensitivity reactions due to thiacetazone in HIV-1 seropositive patients treated for tuberculosis. Lancet 1991; 337: 627-30.
- 444. Ipuge YAI, Rieder HL, Enarson DA. Adverse cutaneous reactions to thiacetazone for tuberculosis treatment in Tanzania. Lancet 1995; 346: 657-60.
- 445. Kelly P, Buve A, Foster SD, McKenna M, Donnelly M, Sipatunyana G. Cutaneous reactions to thiacetazone in Zambia implications for tuberculosis treatment strategies. Trans R Soc Trop Med Hyg 1994; 88: 113-5.
- 446. Saiag P, Caumes E, Chosidow O, Revuz J, Roujeau JC. Drug-induced toxic epidermal necrolysis (Lyell syndrome) in patients infected with the human immunodeficiency virus. J Am Acad Dermatol 1992; 26: 567-74.
- 447. Dukes CS, Sugarman J, Cegielski JP, Lallinger GJ, Mwakyusa DH. Severe cutaneous hypersensitivity reactions during treatment of tuberculosis in patients with HIV infection in Tanzania. Trop Geogr Med 1992; 44: 308-11.
- 448. Roujeau JC. Toxidermies au cours de l'infection à VIH. Presse Méd 1994; 23: 111-2.
- 449. Nunn P, Porter J, Winstanley P. Thiacetazone avoid like poison or use with care? Trans R Soc Trop Med Hyg 1993; 87: 578-82.
- 450. van Gorkom J, Kibuga DK. Cost-effectiveness and total costs of three alternative strategies for the prevention and management of severe skin reactions attributable to thiacetazone in the treatment of human immunodeficiency virus positive patients with tuberculosis in Kenya. Tubercle Lung Dis 1996; 77: 30-6.
- 451. Sbarbaro J, Blomberg B, Chaulet P. Fixed-dose combination formulations for tuberculosis treatment. (Editorial). Int J Tuberc Lung Dis 1999; 3(suppl): S286-S288.
- 452. International Union Against Tuberculosis and Lung Disease. Assuring bioavailability of fixed-dose combinations of anti-tuberculosis medications. A joint statement of the International Union Against Tuberculosis and Lung Disease and the World Health Organization. As approved by the Executive Committee and Council of the IUATLD, Bangkok, November 1998. Int J Tuberc Lung Dis 1999; 3(suppl): S282-S283.
- 453. Anonymous. Quality assurance: protocol for assessing the rifampicin bioavailability of combined formulations in healthy volunteers. Int J Tuberc Lung Dis 1999; 3(suppl): S284-S285.
- 454. Chaulet P. Implementation of fixed-dose combinations in tuberculosis control: outline of responsibilities. Int J Tuberc Lung Dis 1999; 3(suppl): S353-S357.
- 455. Blomberg B, Kitler ME, Milstien J, Dellepiane N, Fanning A, Norval PY, Spinaci S. Availability of quality fixed-dose combinations for the treatment of tuberculosis: what can we learn from studying the World Health Organization's vaccine model? Int J Tuberc Lung Dis 1999; 3(suppl): S371-S380.

- 456. Mitchison DA. The action of antituberculosis drugs in short-course chemotherapy. Tubercle 1985; 66: 219-25.
- 457. Donald PR, Sirgel FA, Botha FJ, Seifart HI, Parkin DP, Vandenplas ML, Van de Wal BW, Maritz JS, Mitchison DA. The early bactericidal activity of isoniazid related to its dose size in pulmonary tuberculosis. Am J Respir Crit Care Med 1997; 156: 895-900.
- 458. Kamat SR, Dawson JJY, Devadatta S, Fox W, Janardhanam B, Radhakrishna S, Ramakrishnan CV, Somasundaram PR, Stott H, Velu S. A controlled study of the influence of segregation of tuberculous patients for one year on the attack rate of tuberculosis in a 5-year period of close family contacts in south India. Bull World Health Organ 1966; 34: 517-32.
- 459. Rouillon A, Perdrizet S, Parrot R. Transmission of tubercle bacilli: the effects of chemotherapy. Tubercle 1976; 57: 275-99.
- 460. Grosset J. New microbial aspects of the treatment of tuberculosis. *In:* Luvarà A, Ed. Rifampicin. TB today: from prevention of resistance to prevention of relapse. A symposium held at the Forlanini Institute, Rome June 19, 1997. Amsterdam: Excerpta Medica, 1977; 1-11.
- 461. Fox W. The current status of short-course chemotherapy. Tubercle 1979; 60: 177-90.
- 462. Dickinson JM, Mitchison DA. Experimental models to explain the high sterilizing activity of rifampin in the chemotherapy of tuberculosis. Am Rev Respir Dis 1981; 123: 367-71.
- 463. Mitchison DA. Treatment of tuberculosis. The Mitchell Lecture 1979. J Roy Coll Phys London 1980; 14: 91-9.
- 464. Mitchison DA. How drug resistance emerges as a result of poor compliance during short course chemotherapy of tuberculosis. Int J Tuberc Lung Dis 1998; 2: 10-5.
- 465. British Medical Research Council. Streptomycin treatment of pulmonary tuberculosis. A Medical Research Council investigation. BMJ 1948; 2: 769-83.
- 466. Canetti G. The J. Burns Amberson lecture. Present aspects of bacterial resistance in tuberculosis. Am Rev Respir Dis 1965; 92: 687-703.
- 467. Pyle MM. Relative numbers of resistant tubercle bacilli in sputa of patients before and during treatment with streptomycin. Proc Staff Meet Mayo Clin 1947; 22: 465-73.
- 468. Crofton J, Mitchison DA. Streptomycin resistance in pulmonary tuberculosis. BMJ 1948; 2: 1009-15.
- 469. Mitchison DA. Sensitivity testing. *In:* Heaf F, Rusby NL, Eds. Recent advances in respiratory tuberculosis. London: J & A Churchill Ltd, 1968; 160-182.
- 470. Orme IM. The latent tuberculosis bacillus (I'll let you know if I ever see one). (Counterpoint). Int J Tuberc Lung Dis 2001; 5: 589-93.

- 471. Mitchison DA. Basic mechanisms of chemotherapy. Chest 1979; 76: 771-81.
- 472. Mitchison DA, Dickinson JM. Laboratory aspects of intermittent drug therapy. Postgrad Med J 1971; 47: 737-41.
- 473. Hill AB. Memories of the British streptomycin trial in tuberculosis. The first randomized clinical trial. Contr Clin Trials 1990; 11: 77-9.
- 474. D'Arcy Hart P. A change in scientific approach: from alteration to randomised allocation in clinical trials in the 1940s. BMJ 1999; 319: 572-3.
- 475. Iseman MD, Sbarbaro JA. Short-course chemotherapy of tuberculosis. Hail Britannia (and friends). Editorial. Am Rev Respir Dis 1991; 143: 697-8.
- 476. O'Brien RJ, Vernon AA. New tuberculosis drug development. How can we do better? (Editorial). Am J Respir Crit Care Med 1998; 157: 1705-7.
- 477. Crofton J. "Sputum conversion" and the metabolism of isoniazid. (Correspondence). Am Rev Tuberc Pulm Dis 1958; 77: 869-71.
- 478. Crofton J. Chemotherapy of pulmonary tuberculosis. BMJ 1959; 1: 1610-4.
- 479. Ferebee SH, Theodore A, Mount FW. Long-term consequences of isoniazid alone as initial therapy. United States Public Health Service Tuberculosis Therapy Trials. Am Rev Respir Dis 1960; 82: 824-30.
- 480. Ferebee SH. The effect of streptomycin on the emergence of bacterial resistance to isoniazid. A United States Public Health Service Cooperative Investigation. Am Rev Tuberc 1953; 67: 553-67.
- 481. Mount FW, Ferebee SH. Control study of comparative efficacy of isoniazid, streptomycin - isoniazid, and streptomycin - para-aminosalicylic acid in pulmonary tuberculosis therapy. IV. Report on forty-week observations on 583 patients with streptomycin-susceptible infections. Am Rev Tuberc 1953; 68:264-9.
- 482. Mount FW, Ferebee SH. United States Public Health Service Cooperative Investigation of antimicrobial therapy of tuberculosis. V. Report of thirty-twoweek observations on combinations of isoniazid, streptomycin, and para-aminosalicylic acid. Am Rev Tuberc 1954; 70: 521-6.
- 483. Mount FW, Ferebee SH. Sequential use of paired combinations of isoniazid, streptomycin, para-aminosalicylic acid, and pyrazinamide. A United States Public Health Service tuberculosis therapy trial. Am Rev Respir Dis 1959; 80: 627-40.
- 484. Doster B, Murray FJ, Newman R, Woolpert SF. Ethambutol in the initial treatment of pulmonary tuberculosis. Am Rev Respir Dis 1973; 107: 177-90.
- 485. Doster B, Newman R. Ethambutol in re-treatment of pulmonary tuberculosis. United States Public Helath Service tuberculosis therapy trial. Am Rev Respir Dis 1968; 98: 825-36.
- 486. Newman R, Doster B, Murray FJ, Woolpert SF. Rifampin in initial treatment of pulmonary tuberculosis. A U.S. Public Health Service tuberculosis therapy trial. Am Rev Respir Dis 1974; 109: 216-32.

- 487. Newman R, Doster B, Murray FJ, Ferebee S. Rifampin in initial treatment of pulmonary tuberculosis. A U.S. Public Health Service tuberculosis therapy trial. Am Rev Respir Dis 1971; 103: 461-76.
- 488. Long MW, Snider DE, Farer LS. U.S. Public Health Service cooperative trial of three rifampin-isoniazid regimens in treatment of pulmonary tuberculosis. Am Rev Respir Dis 1979; 119: 879-93.
- 489. Snider DE, Long MW, Cross FS, Farer LS. Six-months isoniazid-rifampin therapy for pulmonary tuberculosis. Report of a United States Public Health Service cooperative trial. Am Rev Respir Dis 1984; 129: 573-9.
- 490. Geiter LJ, O'Brien RJ, Combs DL, Snider DE. United States Public Health Service tuberculosis therapy trial 21: preliminary results of an evaluation of a combination tablet of isoniazid, rifampin and pyrazinamide. Tubercle 1987; 68: 41-6.
- 491. Combs DL, O'Brien RJ, Geiter LJ. USPHS tuberculosis short-course chemotherapy trial 21: effectiveness, toxicity, and acceptability. Ann Intern Med 1990; 112: 397-406.
- 492. Long ER, Ferebee SH. A controlled investigation of streptomycin treatment in pulmonary tuberculosis. Publ Health Rep 1950; 65: 1421-51.
- 493. Tempel CW, Hughes FJ, Jr., Mardis RE, Towbin MN, Dye WE. Combined intermittent regimens employing streptomycin and para-aminosalicylic acid in the treatment of pulmonary tuberculosis. A comparison with daily and intermittent schedules. Am Rev Tuberc 1951; 63: 295-311.
- 494. British Medical Research Council. Treatment of pulmonary tuberculosis with streptomycin and para-aminosalicylic acid. A Medical Research Council investigation. BMJ 1950; 2: 1073-85.
- 495. Fox W, Sutherland I. A five-year assessment of patients in a controlled trial of streptomycin, para-aminosalicylic acid, and streptomycin plus para-aminosalicylic acid, in pulmonary tuberculosis. Report to the Tuberculosis Chemotherapy Trials Committee of the Medical Research Council. Quarterly J Med New Series 1956; 25: 221-43.
- 496. British Medical Research Council. Long-term chemotherapy in the treatment of chronic pulmonary tuberculosis with cavitation. A report to the Medical Research Council by their Tuberculosis Trials Committee. Tubercle 1962; 43: 201-67.
- 497. Springett VH. Ten-year results during the introduction of chemotherapy for tuberculosis. Tubercle 1971; 52: 73-87.
- 498. Fox W. The John Barnwell lecture. Changing concepts in the chemotherapy of pulmonary tuberculosis. Am Rev Respir Dis 1968; 97: 767-90.
- 499. Toman K. Tuberculosis case-finding and chemotherapy. Questions and answers. 1 ed. World Health Organization, Geneva, 1979; pp. 1-239.
- 500. The Committee on Treatment and the Committee on Bacteriology and Immunology of the International Union Against Tuberculosis. An international investigation of the efficacy of chemotherapy in previously untreated patients with pulmonary tuberculosis. Bull Int Union Tuberc 1964; 34: 80-191.

- 501. East African / British Medical Research Councils. Isoniazid with thiacetazone (thioacetazone) in the treatment of pulmonary tuberculosis in East Africa third investigation: the effect of an initial streptomycin supplement. Tubercle 1966; 47: 1-32.
- 502. Brouet G, Roussel G. Essai 6.9.12. Méthodologie globale et synthèse des résultats. Rev Fr Mal Respir 1977; 5 (suppl 1): 5-13.
- 503. Roussel G. Résultats lointains d'un essai de chimiothérapie de courte durée. L'enquête française 6.9.12. Rev Mal Resp 1983; 11: 847-57.
- 504. Fox W. Whither short-course chemotherapy? Br J Dis Chest 1981; 75: 331-57.
- 505. McCune RM, Jr., Tompsett R. Fate of *Mycobacterium tuberculosis* in mouse tissues as determined by the microbial enumeration technique. I. The persistence of drug-susceptible tubercle bacilli in the tissues despite prolonged antimicrobial therapy. J Exp Med 1956; 104: 737-62.
- 506. McCune RM, Jr., Tompsett R, McDermott W. The fate of *Mycobacterium tuber-culosis* in mouse tissues as determined by the microbial enumeration technique. II. The conversion of tuberculous infection to the latent state by the administration of pyrazinamide and a companion drug. J Exp Med 1956; 104: 763-802.
- 507. East African / British Medical Research Councils. Controlled clinical trial of four short-course (6-month) regimens of chemotherapy for treatment of pulmonary tuberculosis. Second report. Lancet 1973; 1: 1331-8.
- 508. East African / British Medical Research Councils. Controlled clinical trial of four short-course (6-month) regimens of chemothearpy for treatment of pulmonary tuberculosis. Second East African / British Medical Research Council study. Lancet 1974; 2: 1100-6.
- 509. Snider DE, Zierski M, Graczyk J, Bek E, Farer LS. Short-course tuberculosis chemotherapy studies conducted in Poland during the past decade. Eur J Respir Dis 1986; 68: 12-8.
- 510. Singapore Tuberculosis Service, British Medical Research Council. Clinical trial of six-month and four-month regimens of chemotherapy in the treatment of pulmonary tuberculosis. Am Rev Respir Dis 1979; 119: 579-85.
- 511. Singapore Tuberculosis Service, British Medical Research Council. Clinical trial of six-month and four-month regimens of chemotherapy in the treatment of pulmonary tuberculosis: the results up to 30 months. Tubercle 1981; 62: 95-102.
- 512. Singapore Tuberculosis Service, British Medical Research Council. Long-term follow-up of a clinical trial of six-month and four-month regimens of chemotherapy in the treatment of pulmonary tuberculosis. Am Rev Respir Dis 1986; 133: 779-83.
- 513. British Thoracic Association. A controlled trial of six months chemotherapy in pulmonary tuberculosis. First report: results during chemotherapy. Br J Dis Chest 1981; 75: 141-53.

- 514. British Thoracic Association. A controlled trial of six months chemotherapy in pulmonary tuberculosis. Second report: results during the 24 months after the end of chemotherapy. Am Rev Respir Dis 1982; 126: 460-2.
- 515. British Thoracic Society. A controlled trial of 6 months' chemotherapy in pulmonary tuberculosis. Final report: results during the 36 months after the end of chemotherapy and beyond. Br J Dis Chest 1984; 78: 330-6.
- 516. Mehrotra ML, Gautam KD, Chaube CK. Shortest possible acceptable, effective ambulatory chemotherapy in pulmonary tuberculosis: preliminary report I. Am Rev Respir Dis 1981; 124: 239-44.
- 517. Mehrotra ML, Gautam KD, Chaube CK. Shortest possible acceptable effective chemotherapy in ambulatory patients with pulmonary tuberculosis. Part II. Results during the 24 months after the end of chemotherapy. Am Rev Respir Dis 1984; 129: 1016-7.
- 518. Joint Tuberculosis Committee of the British Thoracic Society. Chemotherapy and management of tuberculosis in the United Kingdom: recommendations 1998. Thorax 1998; 53: 536-48.
- 519. East African Medical Research Council, British Medical Research Council. Controlled clinical trial of four short-course regimens of chemotherapy for two durations in the treatment of pulmonary tuberculosis. First report. Third East African/British Medical Research Councils Study. Am Rev Respir Dis 1978; 118: 39-48.
- 520. East African Medical Research Council, British Medical Research Council. Controlled clinical trial of four short-course regimens of chemotherapy for two durations in the treatment of pulmonary tuberculosis. Second report. Third East African/British Medical Research Council Study. Tubercle 1980; 61: 59-69.
- 521. Tuberculosis Research Centre Chennai. A controlled clinical trial of oral shortcourse regimens in the treatment of sputum-positive pulmonary tuberculosis. Int J Tuberc Lung Dis 1997; 1: 509-17.
- 522. Mathew R, Santha T. The treatment of WHO Category 1 tuberculosis with 2HRZE/6EH is indeed defensible. (Counterpoint). Int J Tuberc Lung Dis 2000; 4: 795.
- 523. Tuberculosis Chemotherapy Centre Madras. A concurrent comparison of intermittent (twice-weekly) isoniazid plus streptomycin and daily isoniazid plus PAS in the domiciliary treatment of pulmonary tuberculosis. Bull World Health Organ 1964; 31: 247-71.
- 524. Cohn DL, Catlin BJ, Peterson KL, Judson FN, Sbarbaro JA. A 62-dose, 6-month therapy for pulmonary and extrapulmonary tuberculosis. A twice-weekly, directly observed, and cost-effective regimen. Ann Intern Med 1990; 112: 407-15.
- 525. Snider DE, Rogowski J, Zierski M, Bek E, Long MW. Successful intermittent treatment of smear-positive pulmonary tuberculosis in six months: a cooperative study in Poland. Am Rev Respir Dis 1982; 125: 265-7.

- 526. East African / British Medical Research Council. A pilot study of two regimens of intermittent thiacetazone plus isoniazid in the treatment of pulmonary tuberculosis in East Africa. Tubercle 1974; 55: 211-21.
- 527. Harries AD, Gausi FK, Kwanjana JH, Nyirenda TE, Salaniponi FML. Is oral intermittent initial phase anti-tuberculosis treatment associated with higher mortality in high-prevalent areas in sub-Saharan Africa? Int J Tuberc Lung Dis 2001; 5: 483-5.
- 528. Parthasarathy R, Prabhakar R, Somasundaram PR. Efficacy of 3-month regimen in pulmonary tuberculosis. (Correspondence). Am Rev Respir Dis 1985; 131: 801-2.
- 529. Hong Kong Chest Service, Tuberculosis Research Centre Madras, British Medical Research Council. A controlled trial of 3-month, 4-month, and 6-month regimens of chemotherpy for sputum-smear-negative pulmonary tuberculosis. Results at 5 years. Am Rev Respir Dis 1989; 139: 871-6.
- 530. Rieder HL, Snider DE, Jr., Cauthen GM. Extrapulmonary tuberculosis in the United States. Am Rev Respir Dis 1990; 141: 347-51.
- 531. Powell DA. Tuberculous lymphadenitis. *In:* Schlossberg D, Ed. Tuberculosis and nontuberculous mycobacterial infections. Philadelphia: W.B. Saunders Company, 1999; 186-194.
- 532. Thompson BC. The pathogenesis of tuberculosis of peripheral lymph nodes. A clinical study of 324 cases. Tubercle 1940; 21: 217-35.
- 533. Thompson BC. The pathogenesis of tuberculosis of peripheral lymph nodes. A clinical study of 324 cases. (Continued from p. 235). Tubercle 1940; 21: 260-8.
- 534. Iles PB, Emerson PA. Tuberculous lymphadenitis. BMJ 1974; 1: 143-5.
- 535. Campbell IA, McGavin CR, Friend JAR, Greenwood RM, Jenkins PA, Somner AR. Short course chemotherapy for tuberculosis of lymph nodes: a controlled trial. BMJ 1985; 290: 1106-8.
- 536. Campbell IA, McGavin CR, Friend JAR, Greenwood RM, Jenkins PA, Somner AR. Short course chemotherapy for lymph node tuberculosis: final report at 5 years. Br J Dis Chest 1988; 82: 282-4.
- 537. Campbell IA, Ormerod LP, Friend JAR, Jenkins PA, Prescott RJ. Six months *versus* nine months chemotherapy for tuberculosis of lymph nodes: final results. Respir Med 1993; 87: 621-3.
- 538. van Loenhout-Rooyackers JH, Laheij RJR, Richter C, Verbeek ALM. Shortening the duration of treatment for cervical tuberculous lymphadenitis. Eur Respir J 2000; 15: 192-5.
- 539. Griffiths DL. Symposium on surgical and medical treatment of tuberculosis in developing countries. The treatment of tuberculosis of bone and joint. Trans R Soc Trop Med Hyg 1978; 72: 559-63.
- 540. Leong JCY. Tuberculosis of the spine. J Bone Joint Surg 1993; 75-B: 173-5.

- 541. Konstam PG, Konstam ST. Spinal tuberculosis in southern Nigeria. With special reference to ambulant treatment of thoracolumbar disease. J Bone Joint Surg 1958; 40-B: 26-32.
- 542. Konstam PG, Blesovsky A. The ambulant treatment of spinal tuberculosis. Br J Surg 1962; 50: 26-38.
- 543. Hodgson AR, Stock FE. Anterior spinal fusion. A preliminary communication on the radical treatment of Pott's disease and Pott's paraplegia. Br J Surg 1956; 44: 266-75.
- 544. Hodgson AR, Stock FE. Anterior spine fusion for the treatment of tuberculosis of the spine. The operative findings and results of treatment in the first one hundred cases. J Bone Joint Surg 1960; 42-A: 295-310.
- 545. Hodgson AR, Yau A, Kwon JS, Kim D. A clinical study of 100 consecutive cases of Pott's paraplegia. Clin Orthop Rel Res 1964; 36: 128-50.
- 546. Hodgson AR, Skinsnes OK, Leong CY. The pathogenesis of Pott's paraplegia. J Bone Joint Surg 1967; 49-A: 1147-56.
- 547. Anonymous. Tuberculosis of the spine. BMJ 1974; 2: 613-4.
- 548. Anonymous. Tuberculosis of the spine. Lancet 1974; 2: 137-8.
- 549. Medical Research Council. A controlled trial of ambulant out-patient treatment and in-patient rest in bed in the management of tuberculosis of the spine in young Korean patients on standard chemotherapy. A study in Masan, Korea. First Report of the Medical Research Council Working Party on Tuberculosis of the Spine. J Bone Joint Surg 1973; 55-B: 678-97.
- 550. Medical Research Council. A controlled trial of plaster-of-Paris jackets in the management of ambulant outpatient treatment of tuberculosis of the spine in children on standard chemotherapy: a study in Pusan, Korea. Second Report of the Medical Research Council Working Party on Tuberculosis of the Spine. Tubercle 1973; 54: 262-82.
- 551. Medical Research Council. A controlled trial of débridement and ambulatory treatment in the management of tuberculosis of the spine in patients on standard chemotherapy. A study in Bulawayo, Rhodesia. Third Report of the Medical Research Council Working Party on Tuberculosis of the Spine. J Trop Med Hyg 1974; 77: 72-92.
- 552. Medical Research Council. A controlled trial of anterior spinal fusion and débridement in the surgical managment of tuberculosis of the spine in patients on standard chemotherapy: a study in Hong Kong. Fourth Report of the Medical Research Council Working Party on Tuberculosis of the Spine. Br J Surg 1974; 61: 853-66.
- 553. Medical Research Council. A five-year assessment of controlled trials of inpatient and out-patient treatment and of plaster-of-Paris jackets for tuberculosis of the spine in children on standard chemotherapy. Fifth Report of the Medical Research Council Working Party on Tuberculosis of the Spine. J Bone Joint Surg (Br) 1976; 58-B: 399-411.

- 554. Medical Research Council. Five-year assessments of controlled trials of ambulatory treatment, debridement and anterior spinal fusion in the management of tuberculosis of the spine. Studies in Bulawayo (Rhodesia) and in Hong Kong. Sixth Report of the Medical Research Council Working Party on Tuberculosis of the Spine. J Bone Joint Surg 1978; 60-B: 163-77.
- 555. Medical Research Council. A controlled trial of anterior spinal fusion and débridement in the surgical management of tuberculosis of the spine in patients on standard chemotherapy: a study in two centres in South Africa. Seventh Report of the Medical Research Council Working Party on Tuberculosis of the Spine. Tubercle 1978; 59: 79-105.
- 556. Medical Research Council. A 10-year assessment of a controlled trial comparing debridement and anterior spinal fusion in the management of tuberculosis of the spine in patients on standard chemotherapy in Hong Kong. Eighth Report of the Medical Research Council Working Party on Tuberculosis of the Spine. J Bone Joint Surg 1982; 64-B: 393-8.
- 557. Medical Research Council. A 10-year assessment of controlled trials of inpatient and outpatient treatment and of plaster-of-Paris jackets for tuberculosis of the spine in children on standard chemotherapy. Ninth Report of the Medical Research Council Working Party on Tuberculosis of the Spine. J Bone Joint Surg (Br) 1985; 67-B: 103-10.
- 558. Medical Research Council. A controlled trial of six-month and nine-month regimens of chemotherapy in patients undergoing radical surgery for tuberculosis of the spine in Hong Kong. Tenth Report of the Medical Research Council Working Party on Tuberculosis of the Spine. Tubercle 1986; 67: 243-59.
- 559. Indian Council of Medical Research, British Medical Research Council. A controlled trial of short-course chemotherapy in patients receiving ambulatory treatment or undergoing radical surgery for tuberculosis of the spine. Ind J Tuberc 1989; 36(Suppl): 1-21.
- 560. Medical Research Council. Controlled trial of short-course regimens of chemotherapy in the ambulatory treatment of spinal tuberculosis. Results at three years of a study in Korea. Twelfth Report of the Medical Research Council Working Party on Tuberculosis of the Spine. J Bone Joint Surg (Br) 1993; 75-B: 240-8.
- 561. Medical Research Council. A 15-year assessment of controlled trials of the management of tuberculosis of the spine in Korea and Hongkong. Thirteenth Report of the Medical Research Council Working Party on Tuberculosis of the Spine. J Bone Joint Surg (Br) 1998; 80-B: 456-62.
- 562. Medical Research Council. Five-year assessment of controlled trials of shortcourse chemotherapy regimens of 6, 9 or 18 months' duration for spinal tuberculosis in patients ambulatory from the start or undergoing radical surgery. Fourteenth Report of the Medical Research Council Working Party on Tuberculosis of the Spine. Intern Orthopaedics 1999; 23: 73-81.

- 563. Ellard GA, Humphries MJ, Allen BW. Cerebrospinal fluid drug concentrations and the treatment of tuberculous meningitis. Am Rev Respir Dis 1993; 148: 650-5.
- 564. D'Oliveira JJG. Cerebrospinal fluid concentrations of rifampin in meningeal tuberculosis. Am Rev Respir Dis 1972; 106: 432-7.
- 565. Holdiness MR. Cerebrospinal fluid pharmacokinetics of the antituberculosis drugs. Clin Pharmacokinetics 1985; 10: 532-4.
- 566. Place VA, Pyle MM, de la Huerga J. Ethambutol in tuberculous meningitis. Am Rev Respir Dis 1969; 99: 783-5.
- 567. Bobrowitz ID. Ethambutol in tuberculous meningitis. Chest 1972; 61: 629-32.
- 568. Holdiness MR. Management of tuberculosis meningitis. Drugs 1990; 39: 224-33.
- 569. Hughes IE, Smith H, Kane PO. Ethionamide: its passage into the cerebrospinal fluid in man. Lancet 1962; 1: 616-2.
- 570. Donald PR, Seifart HI. Cerebrospinal fluid concentrations of ethionamide in children with tuberculous meningitis. J Pediatr 1989; 115: 483-6.
- 571. Humphries M. The management of tuberculous meningitis. (Editorial). Thorax 1992; 47: 577-81.
- 572. Visudhiphan P, Chiemchanya S. Evaluation of rifampicin in the treatment of tuberculous meningitis in children. J Pediatr 1975; 87: 983-6.
- 573. Donald PR, Schoeman JF, Van Zyl LE, de Villiers JN, Pretorius M, Springer P. Intensive short course chemotherapy in the management of tuberculous meningitis. Int J Tuberc Lung Dis 1998; 2: 704-11.
- 574. Kole HM. Thiacetazone-induced hypersensitivity. (Correspondence). Lancet 1991; 338: 583-4.
- 575. Pozniak AL, MacLeod GA, Mahari M, Legg W, Weinberg J. The influence of HIV status on single and multiple drug reactions to antituberculous therapy in Africa. AIDS 1992; 6: 809-14.
- 576. Grosset JH. Treatment of tuberculosis in HIV infection. Tubercle Lung Dis 1992; 73: 384-7.
- 577. Okwera A, Johnson JL, Vjecha MJ, Wolski K, Whalen CC, Hom D, Huebner R, Mugerwa RD, Ellner JJ. Risk factors for adverse drug reactions during thiacetazone treatment of pulmonary tuberculosis in human immunodeficiency virus infected adults. Int J Tuberc Lung Dis 1997; 1: 441-5.
- 578. Chaisson RE, Schecter GF, Theuer CP, Rutherford GW, Echenberg DF, Hopewell PC. Tuberculosis in patients with the acquired immunodeficiency syndrome. Clinical features, response to therapy, and survival. Am Rev Respir Dis 1987; 136: 570-4.
- 579. Jones BE, Otaya M, Antoniskis D, Sian S, Wang F, Mercado A, Davidson PT, Barnes PF. A prospective evaluation of antituberculosis therapy in patients with human immunodeficiency virus infection. Am J Respir Crit Care Med 1994; 150: 1499-502.

- 580. Nolan CM. Failure of therapy for tuberculosis in human immunodeficiency virus infection. Am J Med Sci 1992; 304: 168-73.
- 581. Peloquin CA, MacPhee AA, Berning SE. Malabsorption of antimycobacterial medications. (Correspondence). N Engl J Med 1993; 329: 1122-3.
- 582. Berning SE, Huitt GA, Iseman MD, Peloquin CA. Malabsorption of antituberculosis medications by a patient with AIDS. N Engl J Med 1992; 327: 1817-8.
- 583. Patel KB, Belmonte R, Crowe HM. Drug malabsorption and resistant tuberculosis in HIV-infected patients. (Correspondence). N Engl J Med 1995; 332: 336-7.
- 584. Choudhri SH, Hawken M, Gathua S, Minyiri GO, Watkins W, Sahai J, Sitar DS, Aoki FY, Long R. Pharmacokinetics of antimycobacterial drugs in patients with tuberculosis, AIDS, and diarrhea. Clin Infect Dis 1997; 25: 104-11.
- 585. Taylor B, Smith PJ. Does AIDS impair the absorption of anti-tuberculosis agents? Int J Tuberc Lung Dis 1998; 2: 670-5.
- 586. Brindle RJ, Nunn PP, Githui W, Allen BW, Gathua S, Waiyaki P. Quantitative bacillary response to treatment in HIV-associated pulmonary tuberculosis. Am Rev Respir Dis 1993; 147: 958-61.
- 587. Iseman MD. Is standard chemotherapy adequate in tuberculosis patients infected with the HIV? Am Rev Respir Dis 1987; 136: 1326.
- 588. Small PM, Schecter GF, Goodman PC, Sande MA, Chaisson RE, Hopewell PC. Treatment of tuberculosis in patients with advanced human immunodeficiency virus infection. N Engl J Med 1991; 324: 289-94.
- 589. Schürmann D, Bergmann F, Jautzke G, Fehrenbach FJ, Mauch H, Ruf B. Acute and long-term efficacy of antituberculous treatment in HIV-seropositive patients with tuberculosis: a study of 36 cases. J Infect 1993; 26: 45-54.
- 590. Banda H, Kang'ombe C, Harries AD, Nyangulu DS, Whitty CJM, Wirima JJ, Salaniponi FM, Maher D, Nunn P. Mortality rates and recurrent rates of tuberculosis in patients with smear-negative pulmonary tuberculosis and tuberculous pleural effusion who have completed treatment. Int J Tuberc Lung Dis 2000; 4: 968-74.
- 591. Perriëns JH, Colebunders RL, Karahunga C, Willame JC, Jeugmans J, Kaboto M, Mukadi Y, Puwels P, Ryder RW, Prignot J, Piot P. Increased mortality and tuberculosis treatment failure rate among human immunodeficiency virus (HIV) seropositive compared with HIV seronegative patients with pulmonary tuberculosis treated with "standard" chemotherapy in Kinshasa, Zaire. Am Rev Respir Dis 1991; 144: 750-5.
- 592. Githui W, Nunn P, Juma E, Karimi F, Brindle R, Kamunyi R, Gathua S, Gicheha C, Morris J, Omwega M. Cohort study of HIV-positive and HIV-negative tuberculosis, Nairobi, Kenya: comparison of bacteriological results. Tubercle Lung Dis 1992; 73: 203-9.

- 593. Hawken M, Nunn P, Gathua S, Brindle R, Godfrey-Faussett P, Githui W, Odhiambo J, Batchelor B, Gilks C, Morris J, McAdam K. Increased recurrence of tuberculosis in HIV-1-infected patients in Kenya. Lancet 1993; 342: 332-7.
- 594. Perriens JH, Louis ME, Mukadi YB, Brown C, Prignot J, Pouthier F, Portaels F, Willame JC, Mandala JK, Kaboto M, Ryder RW, Roscigno G, Piot P. Pulmonary tuberculosis in HIV-infected patients in Zaire. A controlled trial of treatment for either 6 or 12 months. N Engl J Med 1995; 332: 779-84.
- 595. Narita M, Ashkin D, Hollender ES, Pitchenik AE. Paradoxical worsening of tuberculosis following antiretroviral therapy in patients with AIDS. Am J Respir Crit Care Med 1998; 158: 157-61.
- 596. Centers for Disease Control and Prevention. Prevention and treatment of tuberculosis among patients infected with human immunodeficiency virus: principles of therapy and revised recommendations. Morb Mortal Wkly Rep 1998; 47(No. RR-20): 1-58.
- 597. Vernon A, Burman W, Benator D, Khan A, Bozeman L. Acquired rifamycin monoresistance in patients with HIV-related tuberculosis treated with once-weekly rifapentine and isoniazid. Lancet 1999; 353: 1843-7.
- 598. Rieder HL, Arnadottir T, Trébucq A, Enarson DA. Tuberculosis treatment: dangerous regimens? (Counterpoint). Int J Tuberc Lung Dis 2001; 5: 1-3.
- 599. Fitzgerald DW, Desvarieux M, Svere P, Joseph P, Johnson WD, Jr, Pape JW. Effect of post-treatment isoniazid on prevention of recurrent tuberculosis in HIV-1-infected individuals: a randomised trial. Lancet 2000; 356: 1470-4.
- 600. World Health Organization. Anti-tuberculosis drug resistance in the world. Report No. 2. WHO/CDS/TB/2000.278: Geneva: WHO, 2000: 1-117.
- 601. Tanzania Medical Research Council, British Medical Research Council. Controlled clinical trial of two 6-month regimens of chemotherapy in the treatment of pulmonary tuberculosis. Am Rev Respir Dis 1985; 131: 727-31.
- 602. Swai OB, Aluoch JA, Githui WA, Thiong'o R, Edwards EA, Darbyshire JH, Nunn AJ. Controlled clinical trial of a regimen of two durations for the treatment of isoniazid resistant pulmonary tuberculosis. Tubercle 2000; 69: 5-14.
- 603. Mitchison DA, Nunn AJ. Influence of initial drug resistance on the response to short-course chemotherapy of pulmonary tuberculosis. Am Rev Respir Dis 1986; 133: 423-30.
- 604. Enarson DA. Principles of IUATLD Collaborative Tuberculosis Programmes. Bull Int Union Tuberc Lung Dis 1991; 66: 195-200.
- 605. Espinal MA, Kim SJ, Suarez PG, Kam KM, Khomenko AG, Migliori GB, Baéz J, Kochi A, Dye C, Raviglione MC. Standard short-course chemotherapy for drugresistant tuberculosis. Treatment outcomes in 6 countries. JAMA 2000; 283: 2537-45.

- 606. Crofton J, Chaulet P, Maher D, Grosset J, Harris W, Horne N, Iseman M, Watt B. Guidelines for the management of drug-resistant tuberculosis. Geneva: World Health Organization 1996; 96.210(Rev. 1): 1-40.
- 607. World Health Organization. Coordination of DOTS-plus pilot projects for the management of MDR-TB. WHO/CDS/CDC/TB/99.262: Geneva: WHO, 1999; 1-16.
- 608. World Health Organization. Guidelines for establishing DOTS-Plus projects for the managment of multidrug-resistant tuberculosis (MDR-TB). WHO/CDS/TB/ 2000.279: Geneva: WHO, 2000; 1-87.
- 609. World Health Organization. Report. Multidrug resistant tuberculosis (MDR TB). Basis for the development of an evidence-based case-management strategy for MDR TB within the WHO's DOTS strategy. WHO/TB/99.260: Geneva: WHO, 1999.
- 610. Farmer P, Kim JY. Community based approaches to the control of multidrug resistant tuberculosis: introducing "DOTS-plus". BMJ 1998; 317: 671-4.
- 611. Farmer P, Furin J, Bayona J, Becerra M, Henry C, Hiatt H, Kim JY, Mitnick C, Nardell E, Shin S. Management of MDR-TB in resource-poor countries. (Counter counterpoint). Int J Tuberc Lung Dis 1999; 3: 643-5.
- 612. Van Deun A, Hamid Salim A, Rigouts L, Rahman M, Fissette K, Portaels F. Evaluation of tuberculosis control by periodic or routine susceptibility testing in previously treated cases. Int J Tuberc Lung Dis 2001; 5: 329-38.
- 613. Murray CJL, De Jonghe E, Chum HJ, Nyangulu DS, Salomao A, Styblo K. Cost effectiveness of chemotherapy for pulmonary tuberculosis in three sub-Saharan African countries. Lancet 1991; 338: 1305-8.
- 614. Chum HJ, Ilmolelian G, Rieder HL, Msangi J, Mwinyi N, Zwahlen M, Enarson DA, Ipuge YA. Impact of the change from an injectable to a fully oral regimen on patient adherence to ambulatory treatment in Dar es Salaam, Tanzania. Tubercle Lung Dis 1995; 76: 286-9.
- 615. Boisier P, Rabarijaona L, Rakotomanana F, Ratsitorahina M, Ratsirahonana O, Roux J, Aurégan G. Comparaison de protocoles thérapeutiques utilisés en routine à Madagascar dans le traitement des tuberculoses pulmonaires à microscopie positive. (Résultats préliminaires). Arch Inst Pasteur Madagascar 1995; 62: 72-6.
- 616. Gninafon M, Lambregts-van Weezenbeek CSB, Tawo L, Trebucq A. Ethambutol versus streptomycin during the hospitalized intensive phase of tuberculosis treatment in Benin. (Correspondence). Tubercle Lung Dis 1995; 76: 373-4.
- 617. Singapore Tuberculosis Service, British Medical Research Council. Clinical trial of three 6-month regimens of chemotherapy given intermittently in the continuation phase in the treatment of pulmonary tuberculosis. Am Rev Respir Dis 1985; 132: 374-8.
- 618. Okwera A, Whalen C, Byekwaso F, Vjecha M, Johnson J, Huebner R, Mugerwa R, Ellner J. Randomised trial of thiacetazone and rifampicin-containing regimens for pulmonary tuberculosis in HIV-infected Ugandans. Lancet 1994; 344: 1323-8.

- 619. World Health Organization. The WHO/IUATLD Global Project on Anti-Tuberculosis Drug Resistance Surveillance. Anti-tuberculosis drug resistance in the world. Report No. 2 - prevalence and trends. WHO/CDC/TB/2000.278: Geneva: WHO, 2000; 1-253.
- 620. Lan NTN, Iademarco MF, Binkin NJ, Quy HT, Cô NV. A case series: initial outcome of persons with multidrug-resistant tuberculosis after treatment with the WHO standard retreatment regimen in Ho Chi Minh City, Vietnam. Int J Tuberc Lung Dis 2001; 5: 575-8.
- 621. Pablos-Méndez A, Raviglione MC, Laszlo A, Binkin N, Rieder HL, Bustreo F, Cohn DL, Lambregts-van Weezenbeek CSB, Kim SJ, Chaulet P, Nunn P. Global surveillance for antituberculosis-drug resistance, 1994-1997. N Engl J Med 1998; 338: 1641-9.
- 622. Espinal MA, Laszlo A, Simonsen L, Boulahbal F, Kim SJ, Reniero A, Hoffner S, Rieder HL, Binkin N, Dye C, Williams R, Raviglione MC. Global trends in resistance to antituberculosis drugs. N Engl J Med 2001; 344: 1294-303.
- 623. Pfyffer GE, Bonato DA, Ebrahimzadeh A, Gross W, Hotaling J, Kornblum J, Laszlo A, Roberts G, Salfinger M, Wittwer F, Siddiqi S. Multicenter laboratory validation of susceptibility testing of *Mycobacterium tuberculosis* against classical second-line and newer antimicrobial drugs using the radiometric BACTEC 460 technique and the proportion method with solid media. J Clin Microbiol 1999; 37: 3179-86.
- 624. Bastian I, Rigouts L, Van Deun A, Portaels F. Directly observed treatment, short-course strategy and multidrug-resistant tuberculosis: are any modifications required? Bull World Health Organ 2000; 78: 238-51.
- 625. Grimaldo ER, Tupasi TE, Rivera AB, Quelapio MID, Cardaño RC, Derilo JO, Velen VA. Increased resistance to ciprofloxacin and ofloxacin in multidrug-resistant *Mycobacterium tuberculosis* isolates from patients seen at a tertiary hospital in the Philippines. Int J Tuberc Lung Dis 2001; 5: 546-50.
- 626. Bechan S, Connolly C, Short GM, Standing E, Wilkinson D. Directly observed therapy for tuberculosis given twice weekly in the workplace in urban South Africa. Trans R Soc Trop Med Hyg 1997; 91: 704-7.
- 627. Sbarbaro JA. Compliance: inducements and enforcements. Chest 1979; 76(suppl): 750-6.
- 628. Bayer R, Wilkinson D. Directly observed therapy for tuberculosis: history of an idea. Lancet 1995; 345: 1545-8.
- 629. Snider DE, Hutton MD. Improving patient compliance in tuberculosis treatment programs. U.S. Public Health Service, 1986.
- 630. Walley JD, Khan MA, Newell JN, Khan MH. Effectiveness of the direct observation components of DOTS for tuberculosis: a randomised controlled trial in Pakistan. Lancet 2001; 357: 664-9.

- 631. Weis SE, Slocum PC, Blais FX, King B, Nunn M, Matney GB, Gomez E, Foresman BH. The effect of directly observed therapy on the rates of drug resistance and relapse in tuberculosis. N Engl J Med 1994; 330: 1179-84.
- 632. Trébucq A, Anagonou S, Gninafon M, Lambregts K, Boulahbal F. Prevalence of primary and acquired resistance of *Mycobacterium tuberculosis* to antituberculosis drugs in Benin after 12 years of short-course chemotherapy. Int J Tuberc Lung Dis 1999; 3: 466-70.
- 633. Kenyon TA, Mwasekaga MJ, Huebner R, Rumisha D, Binkin N, Maganu E. Low levels of drug resistance amidst rapidly increasing tuberculosis and human immunodeficiency virus co-epidemics in Botswana. Int J Tuberc Lung Dis 1999; 3: 4-11.
- 634. Snider DE, Jr., Kelly GD, Cauthen GM, Thompson NJ, Kilburn JO. Infection and disease among contacts of tuberculosis cases with drug-resistant and drug-susceptible bacilli. Am Rev Respir Dis 1985; 132: 125-32.
- 635. Teixeira L, Perkins MD, Johnson JL, Palaci M, do Valle Dettoni V, Canedo Rocha LM, Debanne S, Talbot E, Dietze R. Infection and disease among house-hold contacts of patients with multidrug-resistant tuberculosis. Int J Tuberc Lung Dis 2001; 5: 321-8.
- 636. Wilson TM, De Lisle GW, Collins DM. Effect of *inhA* and *katG* on isoniazid resistance and virulence of *Mycobacterium bovis*. Mol Microbiol 1995; 15: 1009-15.
- 637. Manca D, Paul S, Barry CE, III, Freedman VH, Kaplan G. *Mycobacterium tuberculosis* catalase and peroxidase activities and resistance to oxidative killing in human monocytes in vitro. Infect Immun 1999; 67: 74-9.
- 638. van Soolingen D, De Haas PEW, van Doorn HR, Kuijper E, Rinder H, Borgdorff MW. Mutations at amino acid position 315 of the *katG* gene are associated with high-level resistance to isoniazid, other drug resistance, and successful transmission of *Mycobacterium tuberculosis* in The Netherlands. J Infect Dis 2000; 182: 1788-90.
- 639. Hong YP, Kim SJ, Bai JY, Lew WJ, Lee EG. Twenty-year trend of chronic excretors of tubercle bacilli based on the nationwide tuberculosis prevalence surveys in Korea, 1975-1995. Int J Tuberc Lung Dis 2000; 4: 911-9.
- 640. Boulahbal F, Khaled S, Tazir M. The interest of follow-up of resistance of the tubercle bacillus in the evaluation of a programme. Bull Int Union Tuberc Lung Dis 1989; 64: 23-5.
- 641. Ferebee SH. Controlled chemoprophylaxis trials in tuberculosis. A general review. Adv Tuberc Res 1969; 17: 28-106.
- 642. Singh J, Garg PK, Tandon RK. Hepatotoxicity due to antituberculosis therapy. Clinical profile and reintroduction of therapy. J Clin Gastroenterol 1996; 22: 211-4.
- 643. Pech O, May A, Henrich R, Mayer G. Orale Schnelldesensibilisierung mit Rifampicin. Deutsch Med Wschr 2001; 126: 16.

- 644. Sandman L, Schluger NW, Davidow AL, Bonk S. Risk factors for rifampinmonoresistant tuberculosis. A case-control study. Am J Respir Crit Care Med 1999; 159: 468-72.
- 645. Zorini AO. Sul nuovo metodo di chemioprofilassi antitubercolare mediante isoniazide. Rivista Tuberc Malattie Appar Resp 1956; 4: 403-37.
- 646. Zorini AO. Antituberculous chemoprophylaxis. (Correspondence). Am Rev Respir Dis 1982; 127: 943-4.
- 647. von Behring E. Tuberkulose. Einleitung. Beitr Experiment Ther 1902; 5: V-XVIII.
- 648. von Behring E, Römer P, Ruppel WG. Tuberkulose. Beitr Experiment Ther 1902; 5: 1-90.
- 649. Webb GB, Williams WW. Immunity in tuberculosis. Its production in monkeys and children. JAMA 1911; 57: 1431-5.
- 650. Smith T. Certain aspects of natural and acquired resistance to tuberculosis and their bearing on preventive measures. JAMA 1917; 68: 669-74/-764-9.
- 651. Collins DM. New tuberculosis vaccines based on attenuated strains of the *Mycobacterium tuberculosis* complex. Immunology Cell Biol 2000; 78: 342-8.
- 652. Friedmann FF. Immunisierung gegen Tuberkulose. Deutsche Med Wschr 1903; 29: 953-4.
- 653. Bock V. Die Friedmann-Methode. Referat erstattet im Auftrag des staatlichen Ausschusses zur Prüfung des Friedmannschen Heil- und Schutzmittels gegen Tuberkulose. 1 ed. Leipzig: Verlag von S. Hirzel, 1922; pp. 1-157.
- 654. Kruse W. Die Friedmannsche Heil- und Schutzimpfung gegen Tuberkulose. Deutsche Med Wschr 1918; 44: 147-8.
- 655. Calmette A. Preventive vaccination against tuberculosis with BCG. Proc Roy Soc Med 1931; 24: 85-94.
- 656. Sakula A. BCG: who were Calmette and Guérin? Thorax 1983; 38: 806-12.
- 657. Calmette A, Guérin C. Vaccination des bovidés contre la tuberculose et méthode nouvelle de prophylaxie de la tuberculose bovine. Ann Inst Pasteur 1924; 38: 371-98.
- 658. Calmette A, Guérin C, Breton M. Contribution à l'étude de la tuberculose expérimentale du cobaye. (Infection et essais de vaccination par la voie digestive). Ann Inst Pasteur 1907; 21: 401-16.
- 659. Calmette A, Guérin C. Nouvelles recherches expérimentales sur la vaccination des bovidés contre la tuberculose. Ann Inst Pasteur 1920; 34: 553-60.
- 660. Osborn TW. Changes in BCG strains. Tubercle 1983; 64: 1.
- 661. Grange JM, Gibson J, Osborn TW, Collins CH, Yates MD. What is BCG? Tubercle 1983; 64: 129-39.

- 662. Griffith AS. A study of the BCG strain of tubercle bacillus. With an account of two immunity experiments and a preliminary report on the cultivation of tubercle bacilli on bile media. Lancet 1932; 1: 361-3.
- 663. Weill-Hallé B, Turpin R. Premiers essais de vaccination antituberculeuse de l'enfant par le bacille Calmette-Guérin (BCG). Bull Soc Méd Hôpitaux (France) 1925; 49: 1589-601.
- 664. Calmette A, Guérin C, Nègre L, Boquet A. Prémunition des nouveau-nés contre la tuberculose par le vaccin BCG (1921 à 1926). Extrait Ann Inst Pasteur 1926; 40: 1-45.
- 665. Calmette A. La vaccination préventive de la tuberculose par le BCG dans les familles de médecins 1924-1932. Ann Inst Pasteur 1932; 49(suppl): 1-62.
- 666. Institut Pasteur. Vaccination préventive de la tuberculose de l'homme et des animaux par le BCG. 1 ed. Paris: Masson et Cie, 1932; pp. 1-366.
- 667. Calmette A. L'infection bacillaire et la tuberculose chez l'homme et chez les animaux. Processus d'infection et de défense, étude biologique et expérimentale, vaccination préventive. 3 ed. Paris: Masson et Cie, 1928; pp. 1-828.
- 668. Heimbeck J. Sur la vaccination préventive de la tuberculose par injection souscutanée de BCG chez les élèves-infirmières de l'hôpital Ulleval, à Oslo (Norvège). Ann Inst Pasteur 1929; 43: 1229-32.
- 669. Heimbeck J. Tuberculosis in hospital nurses. Tubercle 1936; 18: 97-9.
- 670. Heimbeck J. BCG vaccination in nurses. Tubercle 1948; 29: 84-8.
- 671. Kereszturi C, Park WH, Levine M, Vogel P, Sackett M. Clinical study of BCG vaccination. N Y State J Med 1933; 33: 375-81.
- 672. Baudouin JA. Vaccination against tuberculosis with the BCG vaccine. Can J Public Health 1936; 27: 20-6.
- 673. Petroff SA. A new analysis of the value and safety of protective immunization with BCG (Bacillus Calmette-Guérin). Am Rev Tuberc 1929; 20: 275-96.
- 674. Petroff SA, Branch A, Steenken W, Jr. A study of Bacillus Calmette-Guérin (BCG). I. Biological characteristics, cultural "dissociation" and animal experimentation. Am Rev Tuberc 1929; 19: 9-46.
- 675. Heimbeck J. Immunity to tuberculosis. Arch Intern Med 1928; 41: 336-42.
- 676. Lange B. Untersuchungen zur Klärung der Ursachen der im Anschluss an die Calmette-Impfung aufgetretenen Säuglingserkrankungen in Lübeck. Zeitschr Tuberkulose 1930; 59: 1-18.
- 677. Calmette A. Epilogue de la catastrophe de Lubeck. Presse Méd 1931; 2: 17-8.
- 678. Moegling A. Die "Epidemiologie" der Lübecker Säuglingstuberkulose. Arbeiten a d Reichsges-Amt 1935; 69: 1-24.
- 679. Dormandy T. The white death. 1 ed. London and Rio Grande: The Hambledon Press, 1999; pp. 1-433.

- Lange L. Zu den Tuberkuloseschutzimpfungen in L
 übeck. Zeitschr Tuberkulose 1930; 57: 305-10.
- 681. Schürmann P, Kleinschmidt H. Pathologie und Klinik der Lübecker Säuglingstuberkuloseerkrankungen. Arbeiten ad Reichsges-Amt 1935; 69: 25-204.
- 682. Lange L, Pescator H. Bakteriologische Untersuchungen zur Lübecker Säuglingstuberkulose. Arbeiten a d Reichsges-Amt 1935; 69: 205-305.
- 683. Hashimoto T. The vaccination, theory and practice. BCG. Tokyo: International Medical Foundation Japan, 1975.
- 684. Gheorghiu M, Augier J, Lagrange PH. Maintenance and control of the French BCG strain 1173-P₂ (primary and secondary seed-lots). Bull Inst Pasteur 1983; 81: 281-8.
- 685. Behr MA, Small PM. A historical and molecular phylogeny of BCG strains. Vaccine 1999; 17: 915-22.
- 686. Oettinger T, Jørgensen M, Ladefoged A, Hasløv K, Andersen P. Development of the *Mycobacterium bovis* BCG vaccine: review of the historical and biochemical evidence for a genealogical tree. Tubercle Lung Dis 1999; 79: 243-50.
- 687. Behr MA, Schroeder BG, Brinkman JB, Slayden RA, Barry CE, III. A point mutation in the *mma3* gene is responsible for impaired methoxymycolic acid production in *Mycobacterium bovis* BCG strains obtained after 1927. J Bacteriol 2000; 182: 3394-9.
- 688. Lotte A, Wasz-Höckert O, Poisson N, Dumitrescu N, Verron M, Couvet E. BCG complications. Estimates of the risks among vaccinated subjects and statistical analysis of their main characteristics. Adv Tuberc Res 1984; 21: 107-93.
- 689. Lotte A, Wasz-Höckert O, Poisson N, Dumitrescu N, Verron M, Couvet E. A bibliography of the complications of BCG vaccination. A comprehensive list of the World Literature since the introduction of BCG up to July 1982, supplemented by over 100 personal communications. Adv Tuberc Res 1984; 21: 194-245.
- 690. FitzGerald JM. Management of adverse reactions to Bacille Calmette-Guérin vaccine. Clin Infect Dis 2000; 31(suppl): S75-S76.
- 691. Lotte A, Wasz-Höckert O, Poisson N, Engbaek H, Landmann H, Quast U, Andrasofszky B, Lugosi L, Vadasz I, Mihailescu P, Sudic D, Pal D. Second IUATLD study on complications induced by intradermal BCG-vaccination. Bull Int Union Tuberc Lung Dis 1988; 63(2): 47-59.
- 692. Böttiger M, Romanus V, De Verdier C, Boman G. Osteitis and other complications caused by generalized BCG-itis. Experiences in Sweden. Acta Paediatr Scand 1982; 71: 471-8.
- 693. Schopfer K, Matter L, Brunner C, Pagon S, Stanisic M, Baerlocher K. BCG osteomyelitis. Case report and review. Helv Paediat Acta 1982; 37: 73-81.
- 694. Kröger L, Korppi M, Brander E, Kröger H, Wasz-Höckert O, Bacman A, Rapola J, Launiala K, Katila ML. Osteitis caused by Bacille Calmette-Guérin vaccination: a retrospective analysis of 222 cases. J Infect Dis 1995; 172: 574-6.

- 695. Horwitz O, Meyer J. The safety record of BCG vaccination and untoward reactions observed after vaccination. Adv Tuberc Res 1957; 8: 245-71.
- 696. Tardieu M, Truffot-Pernot C, Carrière JP, Dupic Y, Landrieu P. Tuberculous meningitis due to BCG in two previously healthy children. Lancet 1988; 1: 440-1.
- 697. Abramowsky C, Gonzalez B, Sorensen RU. Disseminated Bacillus Calmette-Guérin infections in patients with primary immunodeficiencies. Am J Clin Pathol 1993; 100: 52-6.
- 698. Stone MM, Vannier AM, Storch SK, Nitta AT, Zhang Y. Brief report: meningitis due to iatrogenic BCG infection in two immunocompromised children. N Engl J Med 1995; 333: 561-3.
- 699. Gonzalez B, Moreno S, Budach R, Valenzuela MT, Henriquez A, Ramos MI, Sorensen RU. Clinical presentation of Bacillus Calmette-Guérin infections in patients with immunodeficiency syndromes. Pediatr Infect Dis J 1989; 8: 201-6.
- 700. Jouanguy E, Altare F, Lamhamedi S, Revy P, Emile JF, Newport M, Levin M, Blanche S, Fischer A. Interferon-gamma-receptor deficiency in an infant with fatal Bacille Calmette-Guérin infection. N Engl J Med 1996; 335: 1956-61.
- 701. Casanova JL, Blanche S, Emile JF, Jouanguy E, Lamhamedi S, Altare F, Stéphan JL, Bernaudin F, Bordigioni P, Turck D, Lachaux A, Albertini M, Bourrillon A, Dommergues JP, Pocidalo MA, Le Deist F, Gaillard JL, Griscelli C, Fischer A. Idiopathic disseminated Bacillus Calmette-Guérin infection: a French national retrospective study. Pediatrics 1996; 98: 774-8.
- 702. Talbot EA, Perkins MD, Fagundes M Silva S, Frothingham R. Disseminated Bacille Calmette-Guérin disease after vaccination: case report and review. Clin Infect Dis 1997; 24: 1139-46.
- 703. Romanus V. The impact of BCG vaccination on mycobacterial disease among children born in Sweden between 1969 and 1993. Smittskyddsinstitutet, Stockholm; 1995.
- 704. Jeena PM, Chhagan MK, Topley J, Coovadia HM. Safety of the intradermal Copenhagen 1331 BCG vaccine in neonates in Durban, South Africa. Bull World Health Organ 2001; 79: 337-43.
- 705. Nousbaum JB, Garre M, Boles JM, Garo B, Larzul JJ. Deux manifestations inhabituelles d'une infection par le virus LAV-HTLV III: BCGite et varicelle pulmonaire. Rev Pneumol Clin 1986; 42: 310-1.
- 706. von Reyn CF, Mann JM, Clements CJ. Human immunodeficiency virus infection and routine childhood immunisation. Lancet 1987; 2: 669-71.
- 707. Weltman AC, Rose DN. The safety of bacille Calmette-Guérin vaccination in HIV infection and AIDS. AIDS 1993; 7: 149-57.
- 708. Houde C, Dery P. *Mycobacterium bovis* sepsis in an infant with human immunodeficiency virus infection. Pediatr Infect Dis 1988; 7: 810-1.
- 709. Boudes P, Sobel A, Deforges L, Leblic E. Disseminated *Mycobacterium bovis* infection from BCG vaccination and HIV infection. (Correspondence). JAMA 1989; 262: 2386.

- 710. Ninane J, Grymonprez A, Burtonboy G, François A, Cornu G. Disseminated BCG in HIV infection. Arch Dis Child 1988; 63: 1268-9.
- 711. Lallemant-Le Coeur S, Lallemant M, Cheynier D, Nzingoula S, Drucker J, Larouzé B. Bacillus Calmette-Guérin immunization in infants born to HIV-1-seropositive mothers. AIDS 1991; 5: 195-9.
- 712. Besnard M, Sauvion S, Offredo C, Gaudelus J, Gaillard JL, Veber F, Blanche S. *Bacillus Calmette-Guérin* infection after vaccination of human immunodeficiency virus-infected children. Pediatr Infect Dis 1993; 12: 993-7.
- 713. Rosenfeldt V, Pærregaard A, Valerius NH. Disseminated infection with Bacillus Calmette-Guerin in a child with advanced HIV disease. Scand J Infect Dis 1997; 29: 526-7.
- 714. Romanus V, Fasth A, Tordai P, Wiholm BE. Adverse reactions in healthy and immunocompromised children under six years of age vaccinated with the Danish BCG vaccine, strain Copenhagen 1331: implications for the vaccination policy in Sweden. Acta Paediatr 1993; 82: 1043-52.
- 715. van Deutekom H, Smulders YM, Roozendaal KJ, van Soolingen D. Bacille Calmette-Guérin (BCG) meningitis in an AIDS patient 12 years after vaccination with BCG. (Correspondence). Clin Infect Dis 1996; 22: 870-1.
- 716. O'Brien KL, Ruff AJ, Louis MA, Desormeaux J, Joseph DJ, McBrien M, Coberly J, Boulos R, Halsey NA. Bacillus Calmette-Guérin complications in children born to HIV-1-infected women with a review of the literature. Pediatrics 1995; 95: 414-8.
- 717. Reynes J, Perez C, Lamaury I, Janbon F, Bertrand A. Bacille Calmette-Guérin adenitis 30 years after immunization in a patient with AIDS. (Correspondence). J Infect Dis 1989; 160: 727.
- 718. Armbruster C, Junker W, Vetter N, Jaksch G. Disseminated Bacille Calmette-Guérin infection in an AIDS patient 30 years after BCG vaccination. (Correspondence). J Infect Dis 1990; 162: 1216.
- 719. Reichman LB. Why hasn't BCG proved dangerous in HIV-infected patients? JAMA 1989; 261: 3246.
- 720. Waddell RD, Lishimpi K, Fordham von Reyn C, Chintu C, Baboo KS, Kreiswirth B, Talbot EA, Karagas MR. Bacteremia due to *Mycobacterium tuber-culosis* or *M. bovis*, Bacille Calmette-Guérin (BCG) among HIV-positive children and adults in Zambia. AIDS 2001; 15: 55-60.
- 721. World Health Organization. Special Programme on AIDS and Expanded Programme on Immunization Joint Statement. Consultation on human immunodeficiency virus (HIV) and routine childhood immunization. WHO Wkly Epidem Rec 1987; 62: 297-9.
- 722. Milstien JB, Gibson JJ. Quality control of BCG vaccine by WHO: a review of factors that may influence vaccine effectiveness and safety. Bull World Health Organ 1990; 68: 93-108.

- 723. World Health Organization. BCG in immunization programmes. Wkly Epidem Rec 2001; 76: 33-9.
- 724. Close GC, Nasiiro R. Management of BCG adenitis in infancy. (Correspondence). J Trop Pediatr 1985; 31: 286.
- 725. Hanley SP, Gumb J, Macfarlane JT. Comparison of erythromycin and isoniazid in treatment of adverse reactions to BCG vaccination. BMJ 1985; 290: 970.
- 726. Oguz F, Müjgan S, Alper G, Alev F, Neyzi O. Treatment of *Bacille Calmette-Guérin*-associated lymphadenitis. Pediatr Infect Dis J 1992; 11: 887-8.
- 727. Victoria MS, Shah BR. Bacillus Calmette-Guérin lymphadenitis: a case report and review of the literature. Pediatr Infect Dis 1985; 4: 295-6.
- 728. Banani SA, Alborzi A. Needle aspiration for suppurative post-BCG adenitis. Arch Dis Child 1994; 71: 446-7.
- 729. Boman G, Sjögren I, Dahlström G. A follow-up study of BCG-induced osteoarticular lesions in children. Bull Int Union Tuberc Lung Dis 1984; 59: 198-200.
- 730. Last JM. A dictionary of epidemiology. 3 ed. New York: Oxford University Press, 1995; pp. 1-180.
- 731. Rothman KJ, Greenland S. Modern epidemiology. 2 ed. Philadelphia: Lippincott - Raven Publishers, 1998; pp. 1-738.
- 732. Orenstein WA, Bernier RH, Dondero TJ, Hinman AR, Marks JS, Bart KJ, Sirotkin B. Field evaluation of vaccine efficacy. Bull World Health Organ 1985; 63: 1055-68.
- 733. Smith PG. Retrospective assessment of the effectiveness of BCG vaccination against tuberculosis using the case-control method. Tubercle 1982; 63: 23-35.
- 734. Schlesselman JJ. Case-control studies. Design, conduct, analysis. 1 ed. New York: Oxford University Press, 1982; pp. 1-354.
- 735. Rodrigues LC, Smith PG. Use of the case-control approach in vaccine evaluation: efficacy and adverse effects. Epidemiol Rev 1999; 21: 56-72.
- 736. Aronson JD, Dannenberg AM. Effect of vaccination with BCG on tuberculosis in infancy and in childhood. Correlation of reactions to tuberculin tests, roentgenologic diagnosis and mortality. Am J Dis Child 1935; 50: 1117-30.
- 737. Feldberg GD. Disease and class. Tuberculosis and the shaping of modern North American society. 1 ed. New Jersey: Rutgers University Press, 1995; pp. 1-214.
- 738. Aronson JD, Palmer CE. BCG vaccination among American Indians. Publ Health Rep 1946; 61: 802-20.
- 739. Townsend JG, Aronson JD, Saylor R, Parr I. Tuberculosis control among the North American Indians. Am Rev Tuberc 1942; 45: 41-2.
- 740. Aronson JD, Aronson CF, Taylor HC. A twenty-year appraisal of BCG vaccination in the control of tuberculosis. Arch Intern Med 1958; 101: 881-93.

- 741. Aronson JD. Protective vaccination against tuberculosis with special reference to BCG vaccination. Am Rev Tuberc 1948; 58: 255-81.
- 742. Rosenthal SR, Loewinsohn E, Graham ML, Liveright D, Thorne MG, Johnson V. BCG vaccination against tuberculosis in Chicago. A twenty-year study statistically analyzed. Pediatrics 1961; 28: 624-41.
- 743. Ferguson RG, Simes AB. BCG vaccination of Indian infants in Saskatchewan. Tubercle 1949; 30: 5-11.
- 744. Levine MI, Sackett MF. Results of BCG immunization in New York City. Am Rev Tuberc 1946; 53: 517-32.
- 745. Wünsch Filho V, de Castilho EA, Rodrigues LC, Huttly SRA. Effectiveness of BCG vaccination against tuberculous meningitis: a case-control study in São Paulo, Brazil. Bull World Health Organ 1990; 68: 69-74.
- 746. Wünsch-Filho V, Moncau JEC, Nakao N. Methodological considerations in casecontrol studies to evaluate BCG vaccine effectiveness. Int J Epidemiol 1993; 22: 149-55.
- 747. Miceli I, De Kantor IN, Colaiácovo D, Peluffo G, Cutillo I, Gorra R, Botta R, Hom S, ten Dam H. Evaluation of the effectiveness of BCG vaccination using the case-control method in Buenos Aires, Argentina. Int J Epidemiol 1988; 17: 629-34.
- 748. Murtagh K. Efficacy of BCG. (Correspondence). Lancet 1980; 1: 423.
- 749. Zodpey SP, Maldhure BR, Dehankar AG, Shrikhande SN. Effectiveness of Bacillus Calmette Guerin (BCG) vaccination against extra-pulmonary tuberculosis: a case-control study. J Commun Dis 1996; 28: 77-84.
- 750. Chavalittamrong B, Chearskul S, Tuchinda M. Protective value of BCG vaccination in children in Bangkok, Thailand. Pediatr Pulmonol 1986; 2: 202-5.
- 751. Sharma RS, Srivastava DK, Asunkanta Singh A, Kumaraswamy DN, Mullick DN, Rungsung N, Datta AK, Bhuiya GC, Datta KK. Epidemiological evaluation of BCG vaccine efficacy in Delhi - 1989. J Com Dis 1989; 21: 200-6.
- 752. Camargos PAM, Guimaraes MDC, Antunes CMF. Risk assessment for acquiring meningitis tuberculosis among children not vaccinated with BCG: a case-control study. Int J Epidemiol 1988; 17: 193-7.
- 753. Myint TT, Win H, Aye HH, Kyaw-Mint TO. Case-control study on evaluation of BCG vaccination of newborn in Rangoon, Burma. Ann Trop Pediatr 1987; 7: 159-66.
- 754. Rosenthal SR, Loewinsohn E, Graham ML, Liveright D, Thorne MG, Johnson V. BCG vaccination in tuberculous households. Am Rev Respir Dis 1961; 84: 690-704.
- 755. Sirinavin S, Chotpitayasunondh T, Suwanjutha S, Sunakorn P, Chantarojanasiri T. Protective efficacy of neonatal Bacillus Calmette-Guérin vaccination against tuberculosis. Pediatr Infect Dis 1991; 10: 359-65.

- 756. Al-Kassimi FA, Al-Hajjaj MS, Al-Orainey IO, Bamgboye EA. Does the protective effect of neonatal BCG correlate with vaccine-induced tuberculin reaction? Am J Respir Crit Care Med 1995; 152: 1575-8.
- 757. Lanckriet C, Lévy-Bruhl D, Bingono E, Siopathis RM, Guérin N. Efficacy of BCG vaccination of the newborn: evaluation by a follow-up study of contacts in Bangui. Int J Epidemiol 1995; 24: 1042-9.
- 758. Packe GE, Innes JA. Protective effect of BCG vaccination in infant Asians: a case-control study. Arch Dis Child 1988; 63: 277-81.
- 759. Young TK, Hershfield ES. A case-control study to evaluate the effectiveness of mass neonatal BCG vaccination among Canadian Indians. Am J Public Health 1986; 76: 783-6.
- 760. Bhat GJ, Diwan VK, Chintu C, Kabika M, Masona J. HIV, BCG and TB in children: a case control study in Lusaka, Zambia. J Trop Pediatr 1993; 39: 219-23.
- 761. Rodrigues LC, Gill ON, Smith PG. BCG vaccination in the first year of life protects children of Indian subcontinent ethnic origin against tuberculosis in England. J Epidemiol Comm Health 1991; 45: 78-80.
- 762. Smith PG. Evaluating interventions against tropical diseases. Int J Epidemiol 1987; 16: 159-66.
- 763. Tuberculosis Prevention Trial. Trial of BCG vaccines in south India for tuberculosis prevention: first report. Bull World Health Organ 1979; 57: 819-27.
- 764. Baily GVJ, Narain R, Mayurnath S, Vallishayee RS, Guld J. Trial of BCG vaccines in south India for tuberculosis prevention. Tuberculosis Prevention Trial, Madras. Indian J Med Res 1980; 72(suppl): 1-74.
- 765. Tuberculosis Research Centre (ICMR) Chennai. Fifteen year follow up of trial of BCG vaccines in south India for tuberculosis prevention. Indian J Med Res 1999; 110: 56-69.
- 766. Comstock GW, Livesay VT, Woolpert SF. Evaluation of BCG vaccination among Puerto Rican children. Am J Public Health 1974; 64: 283-91.
- 767. Frimodt-Moller J, Thomas J, Parthasarathy R. Observations on the protective effect of BCG vaccination in a South Indian rural population. Bull World Health Organ 1964; 30: 545-74.
- 768. Frimodt-Møller J, Acharyulu GS, Pillai KK. Observations on the protective effect of BCG vaccination in a South Indian rural population: fourth report. Bull Int Union Tuberc 1973; 48: 40-9.
- 769. Shapiro C, Cook N, Evans D, Willett W, Fajardo I, Koch-Weser D, Bergonzoli G, Bolanos O, Guerrero R, Hennekens CH. A case-control study of BCG and childhood tuberculosis in Cali, Colombia. Int J Epidemiol 1985; 14: 441-6.
- 770. Capewell S, Leitch AG. The value of contact procedures for tuberculosis in Edinburgh. Br J Dis Chest 1984; 78: 317-29.

- 771. D'Arcy Hart P, Pollock TM, Sutherland I. C. Assessment of the first results of the Medical Research Council's trial of tuberculosis vaccines in adolescents in Great Britain. Adv Tuberc Res 1957; 8: 171-89.
- 772. British Medical Association. BCG and vole bacillus vaccines in the prevention of tuberculosis in adolescents. First (progress) report to the Medical Research Council by their Tuberculosis Vaccines Clinical Trials Committee. BMJ 1956; 1: 1-15.
- 773. British Medical Association. BCG and vole bacillus vaccines in the prevention of tuberculosis in adolescents. Second report to the Medical Research Council by their Tuberculosis Vaccines Clinical Trials Committee. BMJ 1959; 2: 379-96.
- 774. British Medical Association. BCG and vole bacillus vaccines in the prevention of tuberculosis in adolescence and early adult life. Third report to the Medical Research Council by their Tuberculosis Vaccines Clinical Trials Committee. BMJ 1963; 1: 973-8.
- 775. Tuberculosis Vaccines Clinical Trials Committee. BCG and vole bacillus vaccines in the prevention of tuberculosis in adolescence and early adult life. Fourth report to the Medical Research Council by its Tuberculosis Vaccines Clinical Trials Committee. Bull World Health Organ 1972; 46: 371-85.
- 776. D'Arcy Hart P, Sutherland I. BCG and vole bacillus vaccines in the prevention of tuberculosis in adolescence and early adult life. Final report to the Medical Research Council. BMJ 1977; 2: 293-5.
- 777. Karonga Prevention Trial Group. Randomised controlled trial of single BCG, repeated BCG, or combined BCG and killed *Mycobacterium leprae* vaccine for prevention of leprosy and tuberculosis in Malawi. Lancet 1996; 348: 17-24.
- 778. Coetzee AM, Berjak J. B.C.G. in the prevention of tuberculosis in an adult population. Proc Mine Med Off Assoc 1968; 48: 41-53.
- 779. Sepulveda RL, Parcha C, Sorensen RU. Case-control study of the efficacy of BCG immunization against pulmonary tuberculosis in young adults in Santiago, Chile. Tuber Lung Dis 1992; 73: 372-7.
- 780. Houston S, Fanning A, Soskolne CL, Fraser N. The effectiveness of bacillus Calmette-Guérin (BCG) vaccination against tuberculosis. A case-control study in treaty Indians, Alberta, Canada. Am J Epidemiol 1990; 131: 340-8.
- 781. Palmer CE, Shaw LW, Comstock GW. Community trials of BCG vaccination. Am Rev Tuberc Pulm Dis 1958; 77: 877-907.
- 782. Comstock GW, Palmer CE. Long-term results of BCG vaccination in the southern United States. Am Rev Respir Dis 1966; 93: 171-83.
- 783. Comstock GW, Shaw LW. Controlled trial of BCG vaccination in a school population. Publ Health Rep 1960; 75: 583-94.
- 784. Comstock GW, Woolpert SF, Livesay VT. Tuberculosis studies in Muscogee County, Georgia. Twenty-year evaluation of a community trial of BCG vaccination. Publ Health Rep 1976; 91: 276-80.

- 785. Comstock GW, Webster RG. Tuberculosis studies in Muscogee County, Georgia. VII. A twenty-year evaluation of BCG vaccination in a school population. Am Rev Respir Dis 1969; 100: 839-45.
- 786. Bettag OL, Kaluzny AA, Morse D, Radner DB. BCG study at a state school for mentally retarded. Dis Chest 1964; 45: 503-7.
- 787. Vandiviere HM, Dworski M, Melvin IG, Watson KA, Begley J. Efficacy of Bacille Calmette-Guérin and isoniazid-resistant Bacille Calmette-Guérin with and without isoniazid chemoprophylaxis from day of vaccination. II. Field trial in man. Am Rev Respir Dis 1973; 108: 301-13.
- 788. Corrah T, Byass P, Jaffar S, Thomas V, Bouchier V, Stanford JL, Whittle HC. Prior BCG vaccination improves survival of Gambian patients treated for pulmonary tuberculosis. Trop Med Intern Health 2000; 5: 413-7.
- 789. Smith PG. BCG vaccination. *In:* Davies PDO, Ed. Clinical tuberculosis. London: Chapman & Hall Medical, 1994; 297-310.
- 790. Smith PG, Fine PEM. BCG vaccination. *In:* Davies PDO, Ed. Clinical tuberculosis. London: Chapman & Hall Medical, 1998; 417-431.
- 791. Colditz GA, Brewer TF, Berkey CS, Wilson ME, Burdick E, Fineberg HV, Mosteller F. Efficacy of BCG vaccine in the prevention of tuberculosis. Metaanalysis of the published literature. JAMA 1994; 271: 698-702.
- 792. Colditz GA, Berkey CS, Mosteller F, Brewer TF, Wilson ME, Burdick E, Fineberg HV. The efficacy of Bacillus Calmette-Guérin vaccination of newborns and infants in the prevention of tuberculosis: meta-analyses of the published literature. Pediatrics 1995; 96: 29-35.
- 793. World Health Organization. Vaccination against tuberculosis. Report of an ICMR/WHO Scientific Group. Tech Rep Ser 1980; 651: 1-21.
- 794. Comstock GW. Identification of an effective vaccine against tuberculosis. Am Rev Respir Dis 1988; 138: 479-80.
- 795. Dannenberg AM, Jr., Bishai WR, Parrish WR, Parrish N, Ruiz R, Johnson W, Zook BC, Boles JW, Pitt LM. Efficacies of BCG and vole bacillus (*Mycobacterium microti*) vaccines in preventing clinically apparent pulmonary tuberculosis in rabbits: a preliminary report. Vaccine 2001; 19: 796-800.
- 796. Sutrisna B, Utomo P, Komalarini S, Swatrinai S. Penelitan efectifitas vaksin BCG can beberapa faktor lainnya pada anak yang menderita TBC berat di 3 rumah sakit di Jakarta 1981-182. Medika 1983; 9: 143-50.
- 797. Bøe J. Variations in the virulence of BCG. Acta Tuberc Scand 1947; 21: 123-33.
- 798. Mitchison DA, Wallace JG, Bhatia AL, Selkon JB, Subbaiah TV, Lancaster MC. A comparison of the virulence in guinea-pigs of South Indian and British tubercle bacilli. Tubercle 1960; 41: 1-22.
- 799. Dickinson JM, Lefford MJ, Lloyd J, Mitchison DA. The virulence in the guineapig of tubercle bacilli from patients with pulmonary tuberculosis in Hong Kong. Tubercle 1963; 44: 446-51.

- 800. Middlebrook G, Cohn ML. Some observations on the pathogenicity of isoniazid-resistant variants of tubercle bacilli. Science 1953; 118: 297-9.
- 801. Ordway DJ, Sonnenberg MG, Donahue SA, Belisle JT, Orme IM. Drug-resistant strains of *Mycobacterium tuberculosis* exhibit a range of virulence for mice. Infect Immun 1995; 63: 741-3.
- 802. Cohn ML, Davis CI. Infectivity and pathogenicity of drug-resistant strains of tubercle bacilli studied by aerogenic infection of guinea pigs. Am Rev Respir Dis 1970; 102: 97-100.
- 803. Sutherland I, Lindgren I. The protective effect of BCG vaccination as indicated by autopsy studies. Tubercle 1979; 60: 225-31.
- 804. Raleigh JW, Wichelhausen R. Exogenous reinfection with *Mycobacterium tuberculosis* confirmed by phage typing. Am Rev Respir Dis 1973; 108: 639-42.
- Romeyn JA. Exogenous reinfection in tuberculosis. Am Rev Respir Dis 1970; 101: 923-7.
- 806. Small PM, Shafer RW, Hopewell PC, Singh SP, Murphy MJ, Desmond E, Sierra MF, Schoolnik GK. Exogenous reinfection with multidrug-resistant *Mycobacterium tuberculosis* in patients with advanced HIV infection. N Engl J Med 1993; 328: 1137-44.
- 807. Nardell E, McInnis B, Thomas B, Weidhaas S. Exogenous reinfection with tuberculosis in a shelter for the homeless. N Engl J Med 1986; 315: 1570-5.
- 808. Godfrey-Faussett P, Githui W, Batchelor B, Brindle R, Paul J, Hawken M, Gathua S, Odhiambo J, Ojoo JC, Gilks C, McAdam K, Stoker N. Recurrence of HIV-related tuberculosis in an endemic area may be due to relapse or reinfection. Tubercle Lung Dis 1994; 75: 199-202.
- Nolan CM. Reinfection with multidrug-resistant tuberculosis. (Correspondence). N Engl J Med 1993; 329: 811.
- Vynnycky E, Fine PEM. The natural history of tuberculosis: the implications of age-dependent risks of disease and the role of reinfection. Epidemiol Infect 1997; 119: 183-201.
- 811. Sutherland I, Svandová E, Radhakrishna S. The development of clinical tuberculosis following infection with tubercle bacilli. 1. A theoretical model of clinical tuberculosis following infection, linking data on the risk of tuberculous infection and the incidence of clinical tuberculosis in the Netherlands. Tubercle 1982; 63: 255-68.
- ten Dam HG, Pio A. Pathogenesis of tuberculosis and effectiveness of BCG vaccination. Tubercle 1982; 63: 226-33.
- 813. Smith D, Wiegeshaus E, Balasubramanian V. An analysis of some hypotheses to the Chingleput Bacille Calmette-Guérin trial. Clin Infect Dis 2000; 31(suppl 3): S77-S80.

- 814. Vynnycky E, Fine PEM. The annual risk of infection with *Mycobacterium tuberculosis* in England and Wales since 1901. Int J Tuberc Lung Dis 1997; 1: 389-96.
- 815. Abel L, Cua VV, Oberti J, Lap VD, Due LK, Grosset J, Lagrange PH. Leprosy and BCG in southern Vietnam. (Correspondence). Lancet 1990; 335: 1536.
- 816. Brown JAK, Stone MM, Sutherland I. BCG vaccination of children against leprosy in Uganda: results at end of second follow-up. BMJ 1968; 1: 24-7.
- 817. Fine PEM, Maine N, Ponnighaus JM, Clarkson JA, Bliss L. Protective efficacy of BCG against leprosy in northern Malawi. Lancet 1986; 2: 499-502.
- 818. Lwin K, Sundaresan T, Mg Gyi MG, Bechelli LM, Tamondong C, Gallego Garbajosa P, Sansarricq H, Noordeen SK. BCG vaccination of children against leprosy: fourteen-year findings of the trial in Burma. Bull World Health Organ 1985; 63: 1069-78.
- Orege PA, Fine PEM, Lucas SB, Obura M, Okelo C, Okuku P. Case-control study of BCG vaccination as a risk factor for leprosy and tuberculosis in Western Kenya. Int J Leprosy 1993; 61: 542-9.
- Sutherland I. Research into the control of tuberculosis and leprosy in the community. Br Med Bull 1988; 44: 665-78.
- 821. Zodpey SP, Bansod BS, Shrikhande SN, Maldhure BR, Kulkarni SW. Protective effect of Bacillus Calmette Guerin (BCG) against leprosy: a population-based case-control study in Nagpur, India. Lepr Rev 1999; 70: 287-94.
- 822. Pönnighaus JM, Fine PEM, Bliss L, Gruer PJK, Kapira-Mwamondwe B, Msosa E, Rees RJW, Clayton D, Pike MC, Sterne JAC, Oxborrow SM. The Karonga prevention trial: a leprosy and tuberculosis vaccine trial in Northern Malawi. I. Methods of the vaccination phase. Lepr Rev 1993; 64: 338-56.
- 823. Pönnighaus JM, Fine PEM, Sterne JAC, Wilson RJ, Msosa E, Gruer PJK, Jenkins PA, Lucas SB, Liomba NG, Bliss L. Efficacy of BCG vaccine against leprosy and tuberculosis in northern Malawi. Lancet 1992; 339: 636-9.
- 824. Black GF, Dockrell HM, Crampin AC, Floyd S, Weir RE, Bliss L, Sichali L, Mwaungulu L, Kanyongoloka H, Ngwira B, Warndorff DK, Fine PEM. Patterns and implications of naturally acquired immune responses to environmental and tuberculous mycobacterial antigens in Northern Malawi. J Infect Dis 2001; 184: 322-9.
- 825. Stanford JL, Shield MJ, Rook GAW. How environmental mycobacteria may predetermine the protective efficacy of BCG. Tubercle 1981; 62: 55-67.
- 826. Palmer CE, Long MW. Effects of infection with atypical mycobacteria on BCG vaccination and tuberculosis. Am Rev Respir Dis 1966; 94: 553-68.
- 827. Edwards ML, Goodrich JM, Muller D, Pollack A, Ziegler JE, Smith DW. Infection with *Mycobacterium avium-intracellulare* and the protective effects of Bacille Calmette-Guérin. J Infect Dis 1982; 145: 733-41.

- 828. Orme I, Collins FM. Efficacy of *Mycobacterium bovis* BCG vaccination in mice undergoing prior pulmonary infection with atypical mycobacteria. Infect Immun 1984; 44: 28-32.
- 829. Brown CA, Brown IN, Swinburne S. The effect of oral *Mycobacterium vaccae* on subsequent responses of mice to BCG sensitization. Tubercle 1985; 66: 251-60.
- 830. Comstock GW, Edwards PQ. An American view of BCG vaccination, illustrated by results of a controlled trial in Puerto Rico. Scand J Respir Dis 1972; 53: 207-17.
- 831. Fine PEM, Sterne JAC, Ponnighaus JM, Rees RJW. Delayed-type hypersensitivity, mycobacterial vaccines and protective immunity. Lancet 1994; 344: 1245-9.
- Fine PEM. Variation in protection by BCG: implications of and for heterologous immunity. [Published erratum appears in Lancet 1996; 347: 340]. Lancet 1995; 346: 1339-45.
- 833. Romanus V, Hallander HO, Whälén P, Olinder-Nielsen AM, Magnusson PHW, Juhlin I. Atypical mycobacteria in extrapulmonary disease among children. Incidence in Sweden from 1969 to 1990, related to changing BCG vaccination coverage. Tubercle Lung Dis 1995; 76: 300-10.
- 834. Tala E, Romanus V, Tala-Heikkilä M. Bacille Calmette-Guérin vaccination in the 21st century. Eur Respir Mon 1997; 4: 327-53.
- 835. Trnka L, Dankova D, Svandová E. Six years' experience with the discontinuation of BCG vaccination. 4. Protective effect of BCG vaccination against the *Myco-bacterium avium intracellulare* complex. Tubercle Lung Dis 1994; 75: 348-52.
- 836. Elias D, Wolday D, Akuffo H, Petros B, Bronner B, Britton S. Effect of deworming on human T cell responses to mycobacterial antigens in helminth-exposed individuals before and after bacille Calmette-Guérin (BCG) vaccination. Clin Exp Immunol 2001; 123: 219-25.
- 837. Tala-Heikkilä M. Evaluation of the Finnish BCG-revaccination programme in schoolchildren. Ann Univ Turkuensis 1993; 119: 5-65.
- 838. Tala-Heikkilä M, Tuominen JE, Tala EOJ. Bacillus Calmette-Guérin revaccination questionable with low tuberculosis incidence. Am J Respir Crit Care Med 1998; 157: 1324-7.
- 839. Fine PEM. BCG vaccination against tuberculosis and leprosy. Br Med Bull 1988; 44: 691-703.
- 840. Smith PG, Revill WDL, Lukwago E, Rykushin YP. The protective effect of BCG against *Mycobacterium ulcerans* disease: a controlled trial in an endemic area of Uganda. Trans R Soc Trop Med Hyg 1976; 70: 449-57.
- Anonymous. Topical BCG for recurrent superficial bladder cancer. Lancet 1991; 337: 821-2.
- 842. Melekos MD, Chionis H, Pantazakos A, Fokaefs E, Paranychianakis G, Dauaher H. Intravesical Bacillus Calmette-Guérin immunoprophylaxis of superficial bladder cancer: results of a controlled prospective trial with modified treatment schedule. J Urology 1993; 149: 744-8.

- 843. Talic RF, Hargreve TB, Bishop MC, Kirk D, Prescott S. Intravesical Evans Bacille Calmette-Guérin for carcinoma *in situ* of the urinary bladder. Br J Urology 1994; 73: 645-8.
- 844. Wishahi MM, Ismail IMH, El-Sherbini M. Immunotherapy with bacille Calmette-Guérin in patients with superficial transitional cell carcinoma of the bladder associated with bilharziasis. Br J Urology 1994; 73: 649-54.
- 845. Fellows GJ, Parmar MKB, Grigor KM, Hall RR, Heal MR, Wallace DMA. Marker tumour response to Evans and Pasteur Bacille Calmette-Guérin in multiple recurrent pTa/pTI bladder tumours: report from the Medical Research Council Subgroup on Superficial Bladder Cancer (Urological Cancer Working Party). Br J Urology 1994; 73: 639-44.
- 846. Rogerson JW. Intravesical bacille Calmette-Guérin in the treatment of superficial transitional cell carcinoma of the bladder. Br J Urology 1994; 73: 655-8.
- 847. Mack D, Frick J. Five-year results of a phase II study with low-dose Bacille Calmette-Guérin therapy in high-risk superficial bladder cancer. Urology 1995; 45: 958-61.
- 848. Witjes JA, van den Meijden APM, Collette L, Sylvester R, Debruyne FMJ, van Aubel A, Witjes WPJ. Long-term follow-up of an EORTC randomized prospective trial comparing intravesical Bacille Calmette-Guérin-RIVM and mitomycin C in superficial bladder cancer. Urology 1998; 52: 403-10.
- 849. Alexandroff AB, Jackson AM, O'Donnell MA, James K. BCG immunotherapy of bladder cancer: 20 years on. Lancet 1999; 353: 1689-94.
- 850. Malström PU, Wijkström H, Lundholm C, Wester K, Bush C, Norlén BJ. 5year follow-up of a randomized prospective study comparing mitomycin C and Bacillus Calmette-Guérin in patients with superficial bladder carcinoma. J Urology 1999; 161: 1124-7.
- 851. Czarnetzki BM, Macher E, Suciu S, Thomas D, Steerenberg PA, Rümke P. Longterm adjuvant immunotherapy in stage I high risk malignant melanoma, comparing two BCG preparations versus non-treatment in a randomised multicentre study. (EORTC Protocol 18781). Eur J Cancer 1993; 29A: 1237-42.
- 852. Shirakawa T, Enomoto T, Shimazu S, Hopkin JM. The inverse association between tuberculin responses and atopic disorders. Science 1997; 275: 77-9.
- 853. Alm JS, Lilja G, Scheynus A. Early BCG vaccination and development of atopy. Lancet 1997; 350: 400-3.
- 854. Aaby P, Shaheen SO, Heyes CB, Goudiaby A, Hall AJ, Shiell AW, Jensen H, Marchant A. Early BCG vaccination and reduction in atopy in Guinea-Bissau. Clin Experiment Allergy 2000; 30: 644-50.
- 855. Barreto ML, Rodrigues LC, Silva PCR, Assis AMO, Reis MG, Santos CAST, Blanton RE. Lower hookworm incidence, prevalence, and intensitiy of infection in children with a Bacillus Calmette-Guérin vaccination scar. J Infect Dis 2000; 182: 1800-3.

- 856. Elliott AM, Nakiyingi J, Quigley MA, French N, Gilks CF, Whitworth JAG. Inverse association between BCG immunisation and intestinal nematode infestation among HIV-1-positive individuals in Uganda. Lancet 1999; 354: 1000-1.
- 857. Odent M. Future of BCG. (Correspondence). Lancet 1999; 354: 2170.
- 858. Styblo K, Meijer J. Impact of BCG vaccination programmes in children and young adults on the tuberculosis problem. Tubercle 1976; 57: 17-43.
- 859. Rouillon A, Waaler H. BCG vaccination and epidemiological situation. Adv Tuberc Res 1976; 19: 64-126.
- World Health Organization. BCG vaccination policies. Report of a WHO Study Group. Tech Rep Ser 1980; 652: 1-17.
- 861. International Union Against Tuberculosis and Lung Disease. Criteria for discontinuation of vaccination programmes using Bacille Calmette-Guerin (BCG) in countries with a low prevalence of tuberculosis. A statement of the International Union Against Tuberculosis and Lung Disease. Tubercle Lung Dis 1994; 75: 179-80.
- 862. World Health Organization. Global Tuberculosis Programme and Global Programme on Vaccines. Statement on BCG revaccination for the prevention of tuberculosis. WHO Wkly Epidem Rec 1995; 70: 229-31.
- 863. Arbeláez MP, Nelson KE, Muñoz A. BCG vaccine effectiveness on preventing tuberculosis and its interaction with human immunodeficiency virus infection. Int J Epidemiol 2000; 29: 1085-91.
- 864. Centers for Disease Control and Prevention. The role of BCG vaccine in the prevention and control of tuberculosis in the United States: a joint statement by the Advisory Council for the Elimination of Tuberculosis and the Advisory Committee on Immunization Practices. Morb Mortal Wkly Rep 1996; 45(No.RR 4): 1-18.
- 865. Fine PEM, Carneiro IAM, Milstien JB, Clements CJ. Issues relating to the use of BCG in immunization programmes. A discussion document. WHO/V&B/-99.23: Geneva: WHO, 1999; 1-42.
- 866. Greenberg PD, Lax KG, Schechter CB. Tuberculosis in house staff. A decision analysis comparing the tuberculin screening strategy with the BCG vaccination. Am Rev Respir Dis 1991; 143: 490-5.
- Reichman LB, Jordan TJ, Greenberg PD. Decision analysis comparing the tuberculin screening strategy with BCG vaccine. (Correspondence). Am Rev Respir Dis 1992; 145: 732-3.
- 868. Lincoln EM. The effect of antimicrobial therapy on the prognosis of primary tuberculosis in children. Am Rev Tuberc 1954; 69: 682-9.
- Ferebee SH, Mount FW, Anastasiades AA. Prophylactic effects of isoniazid on primary tuberculosis in children. Am Rev Respir Dis 1957; 76: 942-63.
- 870. Mount FW, Ferebee SH. Preventive effects of isoniazid in the treatment of primary tuberculosis in children. N Engl J Med 1961; 265: 713-21.

- 871. Pamra SP, Mathur GP. Effects of chemoprophylaxis on minimal pulmonary tuberculosis lesions of doubtful activity. Bull World Health Organ 1971; 45: 593-602.
- 872. Ferebee SH, Mount FW, Murray FJ, Livesay VT. A controlled trial of isoniazid prophylaxis in mental institutions. Am Rev Respir Dis 1963; 88: 161-75.
- Comstock GW. Isoniazid prophylaxis in an undeveloped area. Am Rev Respir Dis 1962; 86: 810-22.
- 874. Mount FW, Ferebee SH. The effect of isoniazid prophylaxis on tuberculosis morbidity among household contacts of previously known cases of tuberculosis. Am Rev Respir Dis 1962; 85: 821-7.
- 875. Horwitz O, Payne PG, Wilbek E. Epidemiological basis of tuberculosis eradication. 4. The isoniazid trial in Greenland. Bull World Health Organ 1966; 35: 509-29.
- 876. Comstock GW, Ferebee SH, Hammes LM. A controlled trial of communitywide isoniazid prophylaxis in Alaska. Am Rev Respir Dis 1967; 95: 935-43.
- 877. Groth-Petersen E, Østergaard F. Mass chemoprophylaxis of tuberculosis. The acceptability and untoward side effects of isoniazid in a control study in Greenland. Am Rev Respir Dis 1960; 81: 643-52.
- 878. Veening GJJ. Long term isoniazid prophylaxis. Controlled trial on INH prophylaxis after recent tuberculin conversion in young adults. Bull Int Union Tuberc 1968; 41: 169-71.
- 879. Ferebee SH, Mount FW. Tuberculosis morbidity in a controlled trial of the prophylactic use of isoniazid among household contacts. Am Rev Respir Dis 1962; 85: 490-521.
- 880. Egsmose T, Ang'Awa JOW, Poti SJ. The use of isoniazid among household contacts of open cases of pulmonary tuberculosis. Bull World Health Organ 1965; 33: 419-33.
- 881. Bush OB, Jr, Sugimoto M, Fuji Y, Brown FA, Jr. Isoniazid prophylaxis in contacts of persons with known tuberculosis. Second report. Am Rev Respir Dis 1965; 92: 732-40.
- 882. Goletti D, Weissman D, Jackson RW, Collins F, Kinter A, Fauci AS. The in vitro induction of human immunodeficiency virus (HIV) replication in purified protein derivative-positive HIV-infected persons by recall antigen response to *Mycobacterium tuberculosis* is the result of a balance of the effects of endogenous interleukin-2 and proinflammatory cytokines. J Infect Dis 1998; 177: 1332-8.
- 883. Del Amo J, Malin AS, Pozniak A, De Cock KM. Does tuberculosis accelerate the progression of HIV disease? Evidence from basic science and epidemiology. AIDS 1999; 13: 1151-8.
- 884. Pape JW, Jean SS, Ho JL, Hafner A, Johnson WD, Jr. Effect of isoniazid prophylaxis on incidence of active tuberculosis and progression of HIV infection. Lancet 1993; 342: 268-72.

- 885. Mwinga A, Hosp M, Godfrey-Faussett P, Quigley M, Mwaba P, Mugala BN, Nyirenda O, Luo N, Pobee J, Elliott AM, McAdam KPWJ, Porter JDH. Twice weekly tuberculosis preventive therapy in HIV infection in Zambia. AIDS 1998; 12: 2447-57.
- 886. Whalen CC, Johnson JL, Okwera A, Hom DL, Huebner R, Mugyenyi P, Mugerwa RD, Ellner JJ. A trial of three regimens to prevent tuberculosis in Ugandan adults infected with the human immunodeficiency virus. N Engl J Med 1997; 337: 801-8.
- 887. Gordin FM, Matts JP, Miller C, Brown LS, Hafner R, John SL, Klein M, Vaughn A, Besch CL, Perez G, Szabo S, El-Sadr W. A controlled trial of isoniazid in persons with anergy and human immunodeficiency virus infection who are at high risk for tuberculosis. N Engl J Med 1997; 337: 315-20.
- 888. Hawken MP, Meme HK, Chakaya JM, Morris JS, Githui WA, Juma ES, Odhiambo JA, Thiong'o LN, Kimari JN, Ngugi EN, Bwayo JJ, Gilks CF, Plummer FA, Porter JDH, Nunn PP, McAdam KPWJ. Isoniazid preventive therapy for tuberculosis in HIV-1 infected adults: results of a randomized controlled trial. AIDS 1997; 11: 875-82.
- 889. Krebs A, Farer LS, Snider DE, Thompson NJ. Five years of follow-up of the IUAT trial of isoniazid prophylaxis in fibrotic lesions. Bull Int Union Tuberc Lung Dis 1979; 54: 65-9.
- 890. Katz J, Kunofsky S, Damijonaitis V, Lafleur A, Caron T. Effect of isoniazid upon the reactivation of inactive tuberculosis. Preliminary report. Am Rev Respir Dis 1962; 86: 8-15.
- 891. Katz J, Kunofsky S, Damijonaitis V, Lafleur A, Caron T. Effect of isoniazid upon the reactivation of inactive tuberculosis. Final report. Am Rev Respir Dis 1965; 91: 345-50.
- 892. Hong Kong Chest Service/Tuberculosis Research Centre, Madras Medical Research Council, British Medical Research Council. A double-blind placebo-controlled clinical trial of three antituberculosis chemoprophylaxis regimens in patients with silicosis in Hong Kong. Am Rev Respir Dis 1992; 145: 36-41.
- 893. John GT, Thomas PP, Thomas M, Jeyaseelan L, Jacob CK, Shastry JCM. A double-blind randomized controlled trial of primary isoniazid prophylaxis in dialysis and transplant patients. Transplantation 1994; 57: 1683-4.
- 894. Comstock GW, Baum C, Snider DE. Isoniazid prophylaxis among Alaskan Eskimos: a final report of the Bethel isoniazid studies. Am Rev Respir Dis 1979; 119: 827-30.
- 895. Snider DE, Caras GJ, Koplan JP. Preventive therapy with isoniazid. Cost-effectiveness of different durations of therapy. JAMA 1988; 255: 1579-83.
- 896. Comstock GW. How much isoniazid is needed for prevention of tuberculosis among immunocompetent adults? (Counterpoint). Int J Tuberc Lung Dis 1999; 3: 847-50.

- 897. American Thoracic Society, Centers for Disease Control and Prevention. Targeted tuberculin testing and treatment of latent tuberculosis infection. Am J Respir Crit Care Med 2000; 161 (Suppl): S221-S247.
- 898. Citron KM. Control and prevention of tuberculosis: a code of practice. BMJ 1983; 287: 1118-21.
- 899. Subcommittee of the Joint Tuberculosis Committee of the British Thoracic Society. Guidelines on the management of tuberculosis and HIV infection in the United Kingdom. BMJ 1992; 304: 1231-3.
- 900. Lecoeur HF, Truffot-Pernot C, Grosset JH. Experimental short-course preventive therapy of tuberculosis with rifampin and pyrazinamide. Am Rev Respir Dis 1989; 140: 1189-93.
- 901. Halsey NA, Coberly JS, Desormeaux J, Losikoff P, Atkinson J, Moulton LH, Contave M, Johnson M, Davis H, Geiter L, Johnson E, Huebner R, Boulos R, Chaisson RE. Randomised trial of isoniazid versus rifampicin and pyrazinamide for prevention of tuberculosis in HIV-1 infection. Lancet 1998; 351: 786-92.
- 902. Gordin F, Chaisson RE, Matts JP, de Lourdes Garcia M, Hafner R, Valdespino JL, Coberly J, Schecter M, Klukowicz AJ, Barry MA, O'Brien RJ. Rifampin and pyrazinamide vs isoniazid for prevention of tuberculosis in HIV-infected patients. An international randomized trial. JAMA 2000; 283: 1445-50.
- 903. Quigley MA, Mwinga A, Hosp M, Lisse I, Fuchs D, Porter JDH, Godfrey-Faussett P. Long-term effect of preventive therapy for tuberculosis in a cohort of HIV-infected Zambian adults. AIDS 2001; 15: 215-22.
- 904. Centers for Disease Control and Prevention. Fatal and severe hepatitis associated with rifampin and pyrazinamide for the treatment of latent tuberculosis infection
 New York and Georgia, 2000. Morb Mortal Wkly Rep 2001; 50: 289-91.
- 905. Aisu T, Raviglione MC, Van Praag E, Eriki P, Narain JP, Barugahare L, Tembo G, McFarland D, Engwau FA. Preventive chemotherapy for HIV-associated tuberculosis in Uganda: an operational assessment at a voluntary counselling and testing centre. AIDS 1995; 9: 267-73.
- 906. Kochi A. The global tuberculosis situation and the new control strategy of the World Health Organization. (Leading article). Tubercle 1991; 72: 1-6.
- 907. World Health Organization, UNAIDS. Policy statement on preventive therapy against tuberculosis in people living with HIV. Report of a meeting held in Geneva 18 - 20 February 1998. WHO/TB/98.255: Geneva: WHO, 1998; 1-26.
- 908. Dooley DP, Carpenter JL, Rademacher S. Adjunctive corticosteroid therapy for tuberculosis: a critical reappraisal of the literature. Clin Infect Dis 1997; 25: 872-87.
- 909. Bell WJ, Brown PP, Horn DW. Prednisolone in the treatment of acute extensive pulmonary tuberculosis in West Africans. Tubercle 1960; 41: 341-51.

- 910. Research Committee of the Tuberculosis Society of Scotland. Prednisolone in the treatment of pulmonary tuberculosis: a controlled trial. A preliminary report by the Research Committee of the Tuberculosis Society of Scotland. BMJ 1957; 2: 1131-4.
- 911. Weinstein HJ, Koler JJ. Adrenocorticosteroids in the treatment of tuberculosis. N Engl J Med 1959; 260: 412-7.
- 912. Angel JH, Chu LS, Lyons HA. Corticotropin in the treatment of tuberculosis. A controlled study. Arch Intern Med 1961; 108: 353-69.
- 913. British Tuberculosis Association. A trial of corticotrophin and prednisone with chemotherapy in pulmonary tuberculosis. A report from the Research Committee of the British Tuberculosis Association. Tubercle 1961; 42: 391-412.
- 914. British Tuberculosis Association. Trial of corticotropin and prednisone with chemotherapy in pulmonary tuberculosis: a two-year follow-up. A report from the Research Committee of the British Tuberculosis Association. Tubercle 1963; 44: 484-6.
- 915. McLean RL. The role of adrenocorticotrophic and adrenocortico-steroid hormones in the treatment of tuberculosis. Ann N Y Acad Sci 1963; 106: 130-47.
- 916. Marcus H, Yoo OH, Akyol T, Williams MH, Jr. A randomized study of the effects of corticosteroid therapy on healing of pulmonary tuberculosis as judged by clinical, roentgenographic, and physiologic measures. Am Rev Respir Dis 1963; 88: 55-64.
- 917. Johnson JR, Taylor BC, Morrissey JF, Jenne JW, MacDonald FM. Corticosteroids in pulmonary tuberculosis. 1. Overall results in Madison-Minneapolis Veterans Administration Hospitals Steroid Study. Am Rev Respir Dis 1965; 92: 376-91.
- 918. Doster BE. Prednisolone in the treatment of pulmonary tuberculosis. A United States Public Health Service tuberculosis therapy trial. Am Rev Respir Dis 1965; 91: 329-38.
- 919. Malik SK, Martin CJ. Tuberculosis, corticosteroid therapy, and pulmonary function. Am Rev Respir Dis 1969; 100: 13-8.
- 920. Tuberculosis Research Centre Madras. Study of chemotherapy regimens of 5 and 7 months' duration and the role of corticosteroids in the treatment of sputum-positive patients with pulmonary tuberculosis in South India. Tubercle 1983; 64: 73-9.
- 921. Johnson JR, Davey WN. Cortisone, corticotropine, and antimicrobial therapy in tuberculosis in animals and man. A review. Am Rev Tuberc 1954; 70: 623-36.
- 922. Karlson AG, Gainer JH. The influence of cortisone on experimental tuberculosis of guinea pigs. Dis Chest 1951; 20: 469-81.
- 923. Cummings MM, Hudgins PC, Whorton MC, Sheldon WH. The influence of cortisone and streptomycin on experimental tuberculosis in the albino rat. Am Rev Tuberc 1952; 65: 596-602.

- 924. Elliott AM, Halwiindi B, Bagshawe A, Hayes RJ, Luo N, Pobee J, McAdam KPWJ. Use of prednisolone in the treatment of HIV-positive tuberculosis patients. Quarterly J Med 1992; 85: 307-8.
- 925. Heap BJ. Corticosteroids and tuberculosis. (Correspondence). BMJ 1991; 303: 1204.
- 926. Ratcliffe GE. Amoebic disease precipitated by corticosteroids prescribed for tuberculous pleural effusions. Tubercle 1988; 69: 219-21.
- 927. Allen MB, Cooke NJ. Corticosteroids and tuberculosis. (Editorial). BMJ 1991; 303: 871-2.
- 928. Aspin J, O'Hara H. Steroid-treated tuberculous pleural effusions. Br J Tuberc Dis Chest 1958; 52: 81-3.
- 929. Fleishman SJ, Coetzee AM, Mindel S, Berjak J, Lichter AI, Kerrich JE. Antituberculous therapy combined with adrenal steroids in the treatment of pleural effusions. A controlled clinical trial. Lancet 1960; 1: 199-201.
- 930. Mathur KS, Prasad R, Mathur JS. Intrapleural hydrocortisone in tuberculous pleural effusion. Tubercle 1960; 41: 358-62.
- 931. Menon NK. Steroid therapy in tuberculous pleural effusion. Tubercle 1964; 45: 17-20.
- 932. Tani P, Poppius H, Mäkipaja J. Cortisone therapy for exudative tuberculous pleurisy in the light of a follow-up study. Acta Tuberc Scand 1964; 44: 303-9.
- 933. Lee CH, Wang WJ, Lan RS, Tsai YH, Chiang YC. Corticosteroids in the treatment of tuberculous pleurisy. A double-blind, placebo-controlled, randomised study. Chest 1988; 94: 1256-9.
- 934. Paley SS, Milhaly JP, Mais EL, Gittens SA, Lupini B. Prednisone in the treatment of tuberculous pleural effusions. Am Rev Tuberc Pulm Dis 1959; 79: 307-14.
- 935. Filler J, Porter M. Physiologic studies of the sequelae of tuberculous pleural effusion in children treated with antimicrobial drugs and prednisone. Am Rev Respir Dis 1963; 88: 181-8.
- 936. Galarza I, Cañete C, Granados A, Estopà R, Manresa F. Randomised trial of corticosteroids in the treatment of tuberculous pleurisy. Thorax 1995; 50: 1305-7.
- 937. Wyser C, Walzl G, Smedema JP, Swart F, van Schalkwyk EM, Van de Wal BW. Corticosteroids in the treatment of tuberculous pleurisy. A double-blind, placebocontrolled, randomized study. Chest 1996; 110: 333-8.
- 938. Schrire V. Experience with pericarditis at Groote Schuur Hospital, Cape Town. An analysis of one hundred and sixty cases studied over a six-year period. S Afr Med J 1959; 33: 810-7.
- 939. Long RL, Younes M, Patton N, Hershfield E. Tuberculous pericarditis: longterm outcome in patients who received medical therapy alone. Am Heart Journal 1989; 117: 1133-9.

- 940. Rooney JJ, Crocco JA, Lyons HA. Tuberculous pericarditis. Ann Intern Med 1970; 72: 73-8.
- 941. Strang JIG, Gibson DG, Mitchison DA, Girling DJ, Kakaza HHS, Allen BW, Evans DJ, Nunn AJ. Controlled clinical trial of complete open surgical drainage and of prednisolone in treatment of tuberculous pericardial effusion in Transkei. Lancet 1988; 2: 759-64.
- 942. Strang JIG, Gibson DG, Nunn AJ, Kakaza HHS, Girling DJ, Fox W. Controlled trial of prednisolone as adjuvant in treatment of tuberculous constrictve pericarditis in Transkei. Lancet 1987; 2: 1418-22.
- 943. Strang JIG. Tracing patients in rural Africa. Lancet 1996; 348: 1083-4.
- 944. Strang JIG. Tuberculous pericarditis. J Infect 1997; 35: 215-9.
- 945. Hakim JG, Ternouth I, Mushangi E, Siziya S, Robertson V, Malin A. Double blind randomised placebo controlled trial of adjunctive prednisolone in the treatment of effusive tuberculous pericarditis in HIV seropositive patients. Heart 2000; 84: 183-8.
- 946. Haas DW. Editorial response: is adjunctive corticosteroid therapy indicated during tuberculous peritonitis? (Editorial). Clin Infect Dis 1998; 27: 57-8.
- 947. Alrajhi AA, Halim MA, Al-Hokail A, Alrabiah F, Al-Omran K. Corticosteroid treatment of peritoneal tuberculosis. Clin Infect Dis 1998; 27: 52-6.
- 948. Singh MM, Bhargava AN, Jain KP. Tuberculous peritonitis. An evaluation of pathogenetic mechanisms, diagnostic procedures and therapeutic measures. N Engl J Med 1969; 281: 1091-4.
- 949. Bulkeley WCM. Tuberculous meningitis treated with A.C.T.H. and isoniazid. A comparison with intrathecal streptomycin. BMJ 1953; 2: 1127-9.
- 950. Ashby MA, Grant H. Tuberculous meningitis treated with cortisone. Lancet 1955; 1: 65-6.
- 951. Shane SJ, Riley C. Tuberculous meningitis. Combined therapy with cortisone and antimicrobial agents. N Engl J Med 1953; 249: 829-34.
- 952. Choremis C, Papadatos C, Gargoulas A, Drosos C. Intrathecal hydrocortisone in the treatment of tuberculous meningitis. J Pediatr 1957; 50: 138-44.
- 953. Johnson JR, Rurstenberg NA, Patterson R, Schoch HK, Davey WN. Corticotropin and adrenal steroids as adjuncts to the treatment of tuberculous meningitis. Ann Intern Med 1957; 46: 316-31.
- 954. Voljavec BF, Corpe RF. The influence of corticosteroid hormones in the treatment of tuberculous meningitis in Negroes. Am Rev Respir Dis 1960; 81: 539-45.
- 955. Lepper MH, Spies HW. The present status of the treatment of tuberculosis of the central nervous system. Ann N Y Acad Sci 1963; 1963: 106-23.
- 956. O'Toole RD, Thornton GF, Mukherjee MK, Nath RL. Dexamethasone in tuberculous meningitis. Relationship of cerebrospinal fluid effects to therapeutic efficacy. Am Int Med 1969; 70: 39-48.

- 957. Escobar JA, Belsey MA, Dueñas A, Medina P. Mortality from tuberculous meningitis reduced by steroid therapy. Pediatrics 1975; 56: 1050-5.
- 958. Hockaday JM, Smith HMV. Corticosteroids as an adjuvant to the chemotherapy of tuberculous meningitis. Tubercle 1966; 47: 75-91.
- 959. Girgis NI, Farid LS, Hanna LS, Yassin MW, Wallace CK. The use of dexamethasone in preventing ocular complications in tuberculous meningitis. Trans R Soc Trop Med Hyg 1983; 77: 658-9.
- 960. Girgis NI, Farid Z, Kilpatrick ME, Sultan Y, Mikhail IA. Dexamethasone adjunctive treatment for tuberculous meningitis. Pediatr Infect Dis 1991; 10: 179-83.
- 961. Jacobs RF, Sunakorn P, Chotpitayasunondh T, Pope S, Kelleher K. Intensive short course chemotherapy for tuberculous meningitis. Pediatr Infect Dis 1992; 11: 194-8.
- 962. Yechoor VK, Shandera WX, Rodriguez P, Cate TR. Tuberculous meningitis among adults with and without HIV infection. Experience in an urban public health hospital. Arch Intern Med 1996; 156: 1710-6.
- 963. Kumarvelu S, Prasad K, Khosla A, Behari M, Ahuja GK. Randomized controlled trial of dexamethasone in tuberculous meningitis. Tubercle Lung Dis 1994; 75: 203-7.
- 964. Schoeman JF, Van Zyl LE, Laubscher JA, Donald PR. Effect of corticosteroids on intracranial pressure, computed tomographic findings, and clinical outcome in young children with tuberculous meningitis. Pediatrics 1997; 99: 226-31.
- 965. Nemir RL, Cardona J, Lacoius A, David M. Prednisone therapy as an adjunct in the treatment of lymph node-bronchial tuberculosis in childhood. A doubleblind study. Am Rev Respir Dis 1963; 88: 189-98.
- 966. Ip MSM, Lam WK, Mok CK. Endobronchial tuberculosis revisited. Chest 1986; 89: 727-30.
- 967. Toppet M, Malfroot A, Derde MP, Toppet V, Spehl M, Dab I. Corticosteroids in primary tuberculosis with bronchial obstruction. Arch Dis Child 1990; 65: 1222-6.
- 968. Stanford JL, Grange JM. New concepts for the control of tuberculosis in the twenty first century. J Roy Coll Phys London 1993; 27: 218-23.
- 969. Hofstetter FL. Die Behandlung von Stenosen und Blasenveränderungen bei Nierentuberkulose mit Kortikoiden. Prax Klin Pneumol 1980; 34: 469-73.
- 970. Keers RY. Pulmonary tuberculosis. A journey down the centuries. 1 ed. London: Ballière Tyndall, 1978; pp. 1-265.
- 971. Naef AP. De la tuberculose à la greffe du coeur. 1940-1990 parcours d'un chirurgien. 1 ed. Corcelles: Editions Médecine et Hygiène, 1995; pp. 1-101.
- 972. Liebig S. Indikationen zur chirurgischen Behandlung der Lungentuberkulose in der Aera der Kurzzeitchemotherapie. Oeff Gesundh -Wes 1986; 48: 42-8.

- 973. Iseman MD, Madsen L, Goble M, Pomerantz M. Surgical intervention in the treatment of pulmonary disease caused by drug-resistant *Mycobacterium tuber-culosis*. Am Rev Respir Dis 1990; 141: 623-5.
- 974. Pomerantz M, Madsen L, Goble M, Iseman M. Surgical management of resistant mycobacterial tuberculosis and other mycobacterial pulmonary infections. Ann Thoracic Surg 1991; 52: 1108-12.
- 975. Nitta AT, Iseman MD, Newell JD, Madsen LA, Goble M. Ten-year experience with artificial pneumoperitoneum for end-stage, drug-resistant pulmonary tuber-culosis. Clin Infect Dis 1993; 16: 219-22.
- 976. Veen J. Drug resistant tuberculosis: back to sanatoria, surgery and cod-liver oil? (Editorial). Eur Respir J 1995; 8: 1073-5.
- 977. Agarwal SK, Roy DC, Jha N. Empyema thoracis: a review of 70 cases. Ind J Chest Dis All Sci 1985; 27: 17-22.
- 978. Blanco-Perez J, Bordón J, Piñeiro-Amigo L, Roca-Serrano R, Izquierdo R, Abal-Arca J. Pneumothorax in active pulmonary tuberculosis: resurgence of an old complication? Respir Med 1998; 92: 1269-73.
- 979. Elliott AM, Berning SE, Iseman MD, Peloquin CA. Failure of drug penetration and acquisition of drug resistance in chronic tuberculous empyema. Tubercle Lung Dis 1995; 76: 463-7.
- 980. Janssens JP, de Haller R. Spinal tuberculosis in a developed country. A review of 26 cases with special emphasis on abscesses and neurologic complications. Clin Orthop Rel Res 1990; 257: 67-75.
- 981. Heymann SJ, Brewer TF, Wilson ME, Fineberg HV. The need for global action against multidrug-resistant tuberculosis. (Commentary). JAMA 1999; 281: 2138-40.
- 982. Horsburgh CR, Jr. The global problem of muldtidrug-resistant tuberculosis. The genie is out of the bottle. (Editorial). JAMA 2000; 283: 2575-6.
- 983. Iseman MD. Tuberculosis control strategies and utilitarianism. (Editorial). Int J Tuberc Lung Dis 2000; 4: 95.
- 984. Kawaguchi H. Discovery, chemistry, and activity of amikacin. J Infect Dis 1976; 134 (Suppl): S242-S248.
- 985. Kawaguchi H, Naito T, Nakagawa S, Fujisawa K. BB-K 8, a new semisynthetic aminoglycoside antibiotic. J Antibiotics 1972; 25: 695-708.
- 986. Edson RS, Terrell CL. The aminoglycosides. Mayo Clin Proc 1999; 74: 519-28.
- 987. Garcia Rodriguez JA, Martin Luengo F, Saenz Gonzalez MC. Activity of amikacin against *Mycobacterium tuberculosis*. (Correspondence). J Antimicrob Chemother 1978; 4: 293-4.
- 988. Allen BW, Mitchison DA, Chan YC, Yew WW, Allan WGL, Girling DJ. Amikacin in the treatment of pulmonary tuberculosis. Tubercle 1983; 64: 111-8.

- 989. Hoffner SE, Källenius G. Susceptibility of streptomycin-resistant *Mycobacterium tuberculosis* strains to amikacin. Eur J Clin Microbiol 1988; 7: 188-90.
- 990. Donald PR, Sirgel FA, Venter A, Smit E, Parkin DP, Van de Wal BW, Mitchison DA. The early bactericidal activity of amikacin in pulmonary tuberculosis. Int J Tuberc Lung Dis 2001; 5: 533-8.
- 991. Singh YN, Marshall IG, Harvey AL. Some effects of the aminoglycoside amikacin on neuromuscular and autonomic transmission. Br J Anaesth 1978; 50: 109-17.
- 992. Dehpour AR, Samadian T, Roushanzamir F. Interaction of aminoglycoside antibiotics and lithium at the neuromuscular junction. Drugs Exptl Clin Res 1992; 18: 383-7.
- 993. Zarfin Y, Koren G, Maresky D, Perlman M, MacLeod S. Possible indomethacinaminoglycoside interaction in preterm infants. J Pediatr 1985; 106: 511-2.
- 994. Umezawa H, Ueda M, Maeda K, Yagishita K, Kondo S, Okami Y, Utahara R, Osato Y, Nitta K, Takeguchi T. Production and isolation of a new antibiotic, kanamycin. J Antibiotics Japan Ser A 1957; 10: 181-8.
- 995. Umezawa S, Tatsuta K, Koto S. The total synthesis of kanamycin A. J Antibiotics 1968; 21: 367-8.
- 996. Umezawa S, Koto S, Tatsuta K, Hineno H, Nishimura Y, Tsumura T. The total synthesis of kanamycin B. J Antibiotics 1968; 21: 424-5.
- 997. Umezawa S, Koto S, Tatsuta K, Tsumura T. The total synthesis of kanamycin C. J Antibiotics 1968; 21: 162-3.
- 998. Bunn PA. Kanamycin. Med Clin N Am 1970; 54: 1245-57.
- 999. Rempt E. Gehörschäden bei Kanamycinlangzeittherapie. Zeitschr Laryngol Rhinol Otol 1970; 49: 504-9.
- 1000. Alberghina M, Nicoletti G, Torrisi A. Genetic determinants of aminoglycoside resistance in strains of *Mycobacterium tuberculosis*. Chemotherapy 1973; 19: 148-60.
- 1001. Herr JB, Jr, Haney ME, Pittenger GE, Higgins CE. Isolation and characterization of a new peptide antibiotic. Proc Ind Acad Sci 1959; 69: 134.
- 1002. Ho YII, Chan CY, Cheng AFB. In-vitro activities of aminoglyoside-aminocyclitols against mycobacteria. J Antimicrob Chemother 1997; 40: 27-32.
- 1003. Heifets L, Lindholm-Levy P. Comparison of bactericidal activities of streptomycin, amikacin, kanamycin, and capreomycin against *Mycobacterium avium* and *M. tuberculosis* Antimicrob Agents Chemother 1989; 33: 1298-301.
- 1004. Aquinas M, Citron KM. Rifampicin, ethambutol and capreomycin in pulmonary tuberculosis, previously treated with both first and second line drugs: the results of 2 years chemotherapy. Tubercle 1972; 53: 153-65.
- 1005. McClatchy JK, Kanes W, Davidson PT, Moulding TS. Cross-resistance in *M. tuberculosis* to kanamycin, capreomycin and viomycin. Tubercle 1977; 58: 29-34.

- 1006. Freerksen E, Krüger-Thiemer E, Rosenfeld M. Cycloserin (D-4-Amino-isoxazolidin-3-on). Antibiotica et Chemotherapia 1959; 6: 303-96.
- 1007. Shoji JI. Study on orientomycin, identified with D-4-amino-3-isoxazolidone. Study on actinomyces antibiotics. XXXVII. J Antibiotics Japan Ser A 1965; 9: 164-7.
- 1008. Mitui S, Imaizumi S. Study on reduction (eighth report). Identification of orientomycin and cycloserine (oxamycin, substance PA-94). (In Japanese). J Chem Soc Japan Chem Sect 1957; 78: 812-4.
- 1009. Hidy PH, Hodge EB, Young VV, Harned RL, Brewer GA, Philips WF, Runge WF, Stavely HE, Pohland A, Boaz H, Sullivan HR. Structure and reactions of cycloserine. Am Chem Soc 1955; 77: 2345-6.
- 1010. Benda R, Cans MR, Franchel F, Nataf R. Sur l'emploi d'un nouvel antibiotique (oxamycine) dans 12 cas de tuberculose pulmonaire. Rev Tuberc 1956; 20: 568-73.
- 1011. Kuehl FA, Jr., Wolf FJ, Peck RL, Buhs P, Howe E, Putter I, Hunnewell BD, Ormond R, Downing G, Lyons JE, Newstead E, Chaiet L, Folkers K. D-4amino-3-isoxazolidone, a new antibiotic. (Correspondence). J Am Chem Soc 1955; 77: 2344-5.
- 1012. Shull GM, Sardinas JL. P-94, an antibiotic identical with D-4-amino-3-isoxazolidinone (cycloserine, oxamycin). Antibiotics Chemother 1955; 5: 398-9.
- 1013. Ramaswami S, Musser JM. Molecular genetic basis of antimicrobial agent resistance in *Mycobacterium tuberculosis*: 1998 update. Tubercle Lung Dis 1998; 79: 3-29.
- 1014. David HL, Goldman DS, Takayama K. Inhibition of the synthesis of wax D peptidoglycolipid of *Mycobacterium tuberculosis* by D-cycloserine. Infect Immun 1970; 1: 74-7.
- 1015. Iseman MD. Management of multidrug-resistant tuberculosis. Chemotherapy 1999; 45(suppl 2): 3-11.
- 1016. Walker WC, Murdock JM. Cycloserine in the treatment of pulmonary tuberculosis. A report on toxicity. Tubercle 1957; 38: 297-302.
- 1017. Köster E. Klinische Erfahrungen mit Cycloserin bei Lungentuberkulose. Beitr Klin Tuberk 1957; 117: 317-26.
- 1018. Vallade L, Hudonenq H, Jude JP. La neuro-toxicité de la cyclosérine. Mise au point de ses manifestations cliniques et électro-encéphalographiques d'après 30 publications françaises. Presse Méd 1959; 67: 138-40.
- 1019. Isebarth R, Wiedemann O. D-Cycloserin bei Lungentuberkulose. Bericht über eine gemeinschaftliche Untersuchung an neun Kliniken. Tuberkulosearzt 1960; 14: 144-60.
- 1020. Bucco T, Meligrana G, De Luca V. Neurotoxic effects of cycloserine therapy in pulmonary tuberculosis of adolescents and young adults. Scand J Respir Dis 1970; 71 (Suppl): 259-65.

- 1021. Helmy B. Side effects of cycloserine. Scand J Respir Dis 1970; 71 (Suppl): 220-5.
- 1022. Pasargiklian M, Biondi L. Neurologic and behavioural reactions of tuberculous patients treated with cycloserine. Scand J Respir Dis 1970; 71 (Suppl): 201-8.
- 1023. Schultka H. D-Cycloserin bei Tuberkulösen mit gleichzeitig bestehendem psychiatrisch-neurologischem Krankheitsbild. Tuberkulosearzt 1961; 15: 251-4.
- 1024. Vítek V, Rysánek K. Interaction of D-cycloserine with the action of some monoamine oxidase inhibitors. Biochem Pharmacol 1965; 14: 1417-23.
- 1025. Akula SK, Aruna AS, Johnson JE, Anderson DS. Cycloserine-induced Stevens-Johnson syndrome in an AIDS patient with multidrug-resistant tuberculosis. Int J Tuberc Lung Dis 1997; 1: 187-90.
- 1026. Glass F, Mallach HJ, Simsch A. Beobachtungen und Untersuchungen über die gemeinsame Wirkung von Alkohol und D-Cycloserin. Drug Res 1965; 15: 684-8.
- 1027. Bernheim F. The effect of salicylate on the oxygen uptake of the tubercle bacillus. Science 1940; 92: 204.
- 1028. Lehmann J. *Para*-aminosalicylic acid in the treatment of tuberculosis. Lancet 1946; 1: 15-6.
- 1029. Lehmann J. Twenty years afterward. Historical notes on the discovery of the antituberculosis effect of para-aminosalicylic acid (PAS) and the first clinical trials. (Editorial). Am Rev Respir Dis 1964; 90: 953-6.
- 1030. Dubovsky H. The history of para-aminosalicylic acid (pas), the first tuberculosis anti-microbial agent, and streptomycin (sm): a comparative study. Adler Museum Bulletin 1988; 14: 7-11.
- 1031. Dubovsky H. Correspondence with a pioneer, Jürgen Lehmann (1898-1989), producer of the first effective antituberculosis specific. S Afr Med J 1991; 79: 48-50.
- 1032. Peloquin CA, Berning SE, Huitt GA, Childs JM, Singleton MD, James GT. Once-daily and twice-daily dosing of p-aminosalicylic acid granules. Am J Respir Crit Care Med 1999; 159: 932-4.
- 1033. Fodor T, Pataki G, Schrettner M. PAS infusion in treatment of multidrug-resistant tuberculosis. (Correspondence). Int J Tuberc Lung Dis 2000; 4: 187-8.
- 1034. Anonymous. PAS. (Leading article). Tubercle 1973; 54: 165-7.
- 1035. British Medical Research Council. Co-operative controlled trial of a standard regimen of streptomycin, PAS and isoniazid and three alternative regimens of chemotherapy in Britain. Tubercle 1973; 54: 99-129.
- 1036. Huang KL, Beutler SM, Wang C. Hypothyroidism in a patient receiving treatment for multidrug-resistant tuberculosis. Clin Infect Dis 1998; 27: 910.
- 1037. Soumakis SA, Berg D, Harris HW. Hypothyroidism in a patient receiving treatment for multidrug-resistant tuberculosis. Clin Infect Dis 1998; 27: 910-1.

- 1038. Akhtar AJ, Crompton GK, Schonell ME. Para-aminosalicylic acid as a cause of intestinal malabsorption. Tubercle 1968; 49: 328-31.
- 1039. Longstreth GF, Newcomer AD, Westbrook PR. Para-aminosalicylic acid-induced malabsorption. Am J Dig Dis 1972; 17: 731-4.
- 1040. Wurzel HA, Mayock RL. Thrombocytopenia induced by sodium para-aminosalicylic acid. Report of a case. JAMA 1953; 153: 1094-5.
- 1041. Eisner EV, Kasper K. Immune thrombocytopenia due to a metabolite of paraaminosalicylic acid. Am J Med 1972; 53: 790-6.
- 1042. Feigin RD, Zarkowsky HF, Shearer W, Anderson DC. Thrombocytopenia following administration of para-aminosalicylic acid. (Correspondence). J Pediatr 1973; 83: 502-3.
- 1043. Kreukniet J, Blom van Assendelft PM, Mouton RP, Tassman A, Bangma PJ. The influence of para-aminosalicylic acid on isonicotinic acid hydrazide blood level after oral and intravenous administration. Scand J Respir Dis 1967; 47: 236-43.
- 1044. Hanngren Å, Borgå, Sjöqvist F. Inactivation of isoniazid (INH) in Swedish tuberculosis patients before and during treatment with para-aminosalicylic acid (PAS). Scand J Respir Dis 1970; 51: 61-9.
- 1045. Tuberculosis Chemotherapy Centre Madras. A controlled comparison of two fully supervised once-weekly regimens in the treatment of newly diagnosed pulmonary tuberculosis. Tubercle 1973; 54: 23-45.
- 1046. Dandona P, Greenbury E, Becket AG. *Para*-aminosalicylic acid-induced hypoglycemia in a patient with diabetic nephropathy. Postgrad Med 1980; 56: 135-6.
- 1047. Lalonde RG, Barkun J. Prolonged ciprofloxacin therapy fails to prevent reactivation tuberculosis. Clin Infect Dis 1998; 27: 913-4.
- 1048. Zhao BY, Pine R, Domagala J, Drlica K. Fluoroquinolone action against clinical isolates of *Mycobacterium tuberculosis*: effects of a C-8 methoxyl group on survival in liquid media and in human macrophages. Antimicrob Agents Chemother 1999; 43: 661-6.
- 1049. Mitchison DA. Early bactericidal activity and sterilizing activity of ciprofloxycin in pulmonary tuberculosis. (Correspondence). Am J Respir Crit Care Med 1995; 151: 921.
- 1050. Gillespie SH, Kennedy N. Early bactericidal activity and sterilizing activity of ciprofloxacin in pulmonary tuberculosis. (Correspondence). Am J Respir Crit Care Med 1995; 151: 921-2.
- 1051. Yew WW, Piddock LJV, Li MSK, Lyon D, Chan CY, Cheng AFB. In-vitro activity of quinolones and macrolides against mycobacteria. J Antimicrob Chemother 1994; 34: 343-51.
- 1052. Honeybourne D, Wise R, Andrews JM. Ciprofloxacin penetration into lungs. (Correspondence). Lancet 1987; 1: 1040.

- 1053. Thomas L, Naumann P, Crea A. In-vitro-Aktivität von Ciprofloxacin und Ofloxacin gegen Mycobacterium tuberculosis, M. avium, M. africum, M. kansasii und BCG-Stämme. Immun Infekt 1986; 14: 203-7.
- 1054. Heifets LB, Lindholm-Levy PJ. Bacteriostatic and bactericidal activity of ciprofloxacin and ofloxacin against *Mycobacterium tuberculosis* and *Mycobacterium avium* complex. Tubercle 1987; 68: 267-76.
- 1055. Young LS, Berlin OGW, Inderlied CB. Activity of ciprofloxacin and other fluorinated quinolones against mycobacteria. Am J Med 1967; 82: 23-6.
- 1056. Sirgel FA, Botha FJ, Parkin DP, Van de Wal BW, Schall R, Donald PR, Mitchison DA. The early bactericidal activity of ciprofloxacin in patients with pulmonary tuberculosis. Am J Respir Crit Care Med 1997; 156: 901-5.
- 1057. Hohl P, Salfinger M, Kafader FM. In vitro activity of the new quinolone RO 23-6240 (AM-833) and the new cephalosporins RO 15-8074 and RO 19-5247 (T-2525) against *Mycobacterium fortuitum* and *Mycobacterium chelonae*. Eur J Clin Microbiol 1987; 6: 487-8.
- 1058. Tomioka H, Sato K, Akaki T, Kajitani H, Kawahara S, Sakatani M. Comparative in vitro antimicrobial activities of the newly synthesized quinolone HSR-903, sitafloxacin (DU-6859a), gatifloxacin (AM-1155), and levofloxacin against *Mycobacterium tuberculosis* and *Mycobacterium avium* complex. Antimicrob Agents Chemother 1999; 43: 3001-4.
- 1059. Peloquin CA, Berning SE, Huitt GA, Iseman MD. Levofloxacin for drug-resistant *Mycobacterium tuberculosis*. (Correspondence). Ann Pharmacother 1998; 32: 268.
- 1060. Ji B, Lounis N, Truffot-Pernot C, Grosset J. In vitro and in vivo activities of levofloxacin against *Mycobacterium tuberculosis*. Antimicrob Agents Chemother 1995; 39: 1341-4.
- 1061. Ji B, Lounis N, Maslo C, Truffot-Pernot C, Bonnafous P, Grosset J. In vitro and in vivo activities of moxifloxacin and clinafloxacin against *Mycobacterium tuberculosis*. Antimicrob Agents Chemother 1998; 42: 2066-9.
- 1062. Piersimoni C, Morbiducci V, Bornigia S, De Sio G, Scalise G. *In vitro* activity of the new quinolone lomefloxacin against *Mycobacterium tuberculosis*. Am Rev Respir Dis 1992; 146: 1445-7.
- 1063. Gillespie SH, Billington O. Activity of moxifloxacin against mycobacteria. J Antimicrob Chemother 1999; 44: 393-5.
- 1064. Tsukamura M. Antituberculosis activity of ofloxacin (DL 8280) on experimenal tuberculosis in mice. Am Rev Respir Dis 1985; 132: 915.
- 1065. Mangunnegoro H, Hudoyo A. Efficacy of low-dose ofloxacin in the treatment of multidrug-resistant tuberculosis in Indonesia. Chemotherapy 1999; 45(suppl 2): 19-25.
- 1066. Alegre J, Fernandez de Sevilla T, Falcò V, Martinez Vazquez JM. Ofloxacin in miliary tuberculosis. Eur Respir J 1990; 3: 238-9.

- 1067. Kohno S, Koga H, Kaku M, Maesaki S, Hara K. Prospective comparative study of ofloxacin or ethambutol for the treatment of pulmonary tuberculosis. Chest 1992; 102: 1815-8.
- 1068. Casal M, Ruiz P, Herreras A. Study of the in vitro susceptibility of *M. tuberculosis* to ofloxacin in Spain. Int J Tuberc Lung Dis 2000; 4: 588-91.
- 1069. Global Alliance for TB Drug Development. Scientific blueprint for tuberculosis drug development. Tuberculosis 2001; 81(suppl 1): 1-52.
- 1070. Burkhardt JE, Walterspiel JN, Schaad UB. Quinolone arthropathy in animals versus children. Clin Infect Dis 1997; 25: 1196-204.
- 1071. Post FA, Wood R. Tuberculous pleural effusions in HIV-positive patients. (Correspondence). Int J Tuberc Lung Dis 1998; 2: 941.
- 1072. Gugler R, Allgayer H. Effects of antacids on the clinical pharmacokinetics of drugs. An update. Clin Pharmacokinetics 1990; 18: 210-9.
- 1073. Alangaden GJ, Manavathu EK, Vakulenko SB, Zvonok NM, Lerner SA. Characterization of fluoroquinolone-resistant mutant strains of *Mycobacterium tuberculosis* selected in the laboratory and isolated from patients. Antimicrob Agents Chemother 1995; 39: 1700-3.
- 1074. Cambau E, Jarlier V. Resistance to quinolones in mycobacteria. Res Microbiol 1996; 147: 52-9.
- 1075. Berning SE. The role of fluoroquinolones in tuberculosis today. Drugs 2001; 61: 9-18.
- 1076. Brogden RN, Fitton A. Rifabutin. A review of its antimicrobial activity, pharmacokinetic properties and therapeutic efficacy. Drugs 1994; 47: 983-1009.
- 1077. Marsili L, Pasqualucci CR, Vigevani A, Gioia B, Schioppacassi G, Oronzo G. New rifamycins modified at positions 3 and 4. Synthesis, structure and biological evaluation. J Antibiotics 1981; 24: 1033-8.
- 1078. O'Brien RJ, Lyle MA, Snider DE. Rifabutin (ansamycin LM 427): a new rifamycin-S derivative for the treatment of mycobacterial diseases. Rev Infect Dis 1987; 9: 519-30.
- 1079. Kunin CM. Antimicrobial activity of rifabutin. Clin Infect Dis 1996; 22 (Suppl): S3-S14.
- 1080. Heifets LB, Iseman MD. Determination of *in vitro* susceptibility of mycobacteria to ansamycin. Am Rev Respir Dis 1985; 132: 710-1.
- 1081. Woodley CL, Kilburn JO. In vitro susceptibility of *Mycobacterium avium* complex and *Mycobacterium tuberculosis* strains to a spiro-piperidyl rifamycin. Am Rev Respir Dis 1982; 126: 586-7.
- 1082. Gangadharam PRJ, Perumal VK, Jairam BT, Rao PN, Nguyen AK, Farhi DC, Iseman MD. Activity of rifabutin alone or in combination with clofazimine or ethambutol or both against acute and chronic experimental *Mycobacterium intracellulare* infections. Am Rev Respir Dis 1987; 136: 329-33.

- 1083. Perumal VK, Gangadharam PRJ, Iseman MD. Effect of rifabutin on the phagocytosis and intracellular growth of *Mycobacterium intracellulare* in mouse resident and activated peritoneal and alveolar macrophages. Am Rev Respir Dis 1987; 136: 334-7.
- 1084. Perumal VK, Gangadharam PRJ, Heifets LB, Iseman MD. Dynamic aspects of the *in vitro* chemotherapeutic activity of ansamycin (rifabutine) on *Mycobacterium intracellulare*. Am Rev Respir Dis 1985; 132: 1278-80.
- 1085. O'Brien RJ, Geiter LJ, Lyle MA. Rifabutin (ansamycin LM427) for the treatment of pulmonary *Mycobacterium avium* complex. Am Rev Respir Dis 1990; 141: 821-6.
- 1086. Hong Kong Chest Service, British Medical Research Council. A controlled study of rifabutin and an uncontrolled study of ofloxacin in the retreatment of patients with pulmonary tuberculosis resistant to isoniazid, streptomycin and rifampicin. Tubercle Lung Dis 1992; 73: 59-67.
- 1087. McGregor MM, Olliaro P, Wolmarans L, Mabuza B, Bredell M, Felten MK, Fourie PB. Efficacy and safety of rifabutin in the treatment of patients with newly diagnosed pulmonary tuberculosis. Am J Respir Crit Care Med 1996; 154: 1462-7.
- 1088. Gonzalez Montaner LJ, Natal S, Yongchaiyud P, Olliaro P. Rifabutin for the treatment of newly-diagnosed pulmonary tuberculosis: a multinational, randomized, comparative study versus rifampicin. Tubercle Lung Dis 1994; 75: 341-7.
- 1089. Schwander S, Rüsch-Gerdes S, Mateega A, Lutalo T, Tugume S, Kityo C, Rubaramira R, Mugyenyi P, Okwera A, Mugerwa R, Aisu T, Moser R, Ochen K, M'Bonye B, Dietrich M. A pilot study of antituberculosis combinations comparing rifabutin with rifampicin in the treatment of HIV-1 associated tuberculosis. Tubercle Lung Dis 1995; 76: 210-8.
- 1090. Chan SL, Yew WW, Ma WK, Girling DJ, Aber VR, Felmingham D, Allen BW, Mitchison DA. The early bactericidal activity of rifabutin measured by sputum viable counts in Hong Kong patients with pulmonary tuberculosis. Tubercle Lung Dis 1992; 73: 33-8.
- 1091. Blaschke TF, Skinner MH. The clinical pharmacokinetics of rifabutin. Clin Infect Dis 1996; 22(suppl 1): S15-S22.
- 1092. Bendetti MS. Inducing properties of rifabutin, and effects on the pharmacokinetics and metabolism of concomitant drugs. Pharmacol Res 1995; 32: 177-87.
- 1093. Narita M, Stambaugh JL, Hollender ES, Jones D, Pitchenik AE, Ashkin D. Use of rifabutin with protease inhibitors for human immunodeficiency virus-infected patients with tuberculosis. Clin Infect Dis 2000; 30: 779-83.
- 1094. Mancini P, Pasqua F, Mazzei L, Olliaro P. Rifabutin treatment for tuberculosis patients with liver function abnormalities. (Correspondence). J Antimicrob Chemother 1992; 30: 242.

- 1095. Yang B, Koga H, Ohno H, Ogawa K, Fukuda M, Hirakata Y, Maesaki S, Tomono K, Tashiro T, Kohno S. Relationship between antimycobacterial activities of rifampicin, rifabutin and KRM-1648 and *rpoB* mutations of *Mycobacterium tuberculosis*. J Antimicrob Chemother 1998; 42: 621-8.
- 1096. Sintchenko V, Chew WK, Jelfs PJ, Gilbert GL. Mutations in the *rpoB* gene and rifabutin susceptibility of multidrug-resistant *Mycobacterium tuberculosis* strains isolated in Australia. Pathology 1999; 31: 257-60.
- 1097. Arioli V, Berti M, Carniti G, Randisi E, Rossi E, Scotti R. Antibacterial activity of DL 473, a new semisynthetic rifamycin derivative. J Antibiotics 1981; 24: 1026-32.
- 1098. Dickinson JM, Mitchison DA. In vitro properties of rifapentine (MDL 473) relevant to its use in intermittent chemotherapy of tuberculosis. Tubercle 1987; 68: 113-8.
- 1099. Jarvis B, Lamb HM. Rifapentine. Drugs 1998; 56: 607-16.
- 1100. Miyazaki E, Chaisson RE, Bishai WR. Analysis of rifapentine for preventive therapy in the Cornell mouse model of latent tuberculosis. Antimicrob Agents Chemother 1999; 43: 2126-30.
- 1101. Grosset J, Lounis N, Truffot-Pernot C, O'Brien RJ, Raviglione MC, Ji B. Onceweekly rifapentine-containing regimens for treatment of tuberculosis in mice. Am J Respir Crit Care Med 1998; 157: 1436-40.
- 1102. Keung ACF, Owens RC, Jr., Eller MG, Weir SJ, Nicolau DP, Nightingale CH. Pharmacokinetics of rifapentine in subjects seropositive for the human immunodeficiency virus: a phase I study. Antimicrob Agents Chemother 1999; 43: 1230-3.
- 1103. Reith K, Keung A, Toren PC, Cheng L, Eller MG, Weir SJ. Disposition and metabolism of ¹⁴C-rifapentine in health volunteers. Drug Metabolism Disp 1998; 26: 732-8.
- 1104. Marshall JD, Abdel-Rahman S, Johnson K, Kauffman RE, Kearns GL. Rifapentine pharmacokinetics in adolescents. Pediatr Infect Dis J 1999; 18: 882-8.
- 1105. Conte JE, Jr., Golden JA, McQuitty M, Kipps J, Lin ET, Zurlinden E. Singledose intrapulmonary pharmacokinetics of rifapentine in normal subjects. Antimicrob Agents Chemother 2000; 44: 985-90.
- 1106. Gardner TS, Wenis E, Lee J. The synthesis of compounds for the chemotherapy of tuberculosis. IV. The amide function. J Org Chemistry 1954; 19: 753-7.
- 1107. Libermann D, Moyeux M, Rist N, Grumbach F. Sur la préparation de nouveaux thioamides pyridiniques actifs dans la tuberculose expérimentale. C R Acad Sci Paris 1956; 242: 2409-12.
- 1108. Rist N, Grumbach F, Libermann D. Experiments on the antituberculous activity of alpha-ethyl-thioisonicotinamide. Am Rev Respir Dis 1959; 79: 1-5.

- 1109. Lucchesi M. L'éthionamide activité, résistance, résistance croisée. New York: Proceedings of the 20th Conference of IUATLD, 1969; 58-60.
- 1110. Franz H, Urbanczik R, Stoll K, Müller U. Prothionamid-Blutspiegel nach oraler Verabreichung von Prothionamid allein oder kombiniert mit Isoniazid und/oder mit Diamino-Difenylsulfon. Prax Pneumol 1974; 28: 605-12.
- 1111. Cooperative Study Unit on Chemotherapy of Tuberculosis of the National Sanatoria in Japan. Comparison of the clinical usefulness of ethionamide and prothionamide in initial treatment of tuberculosis: tenth series of controlled trials. Tubercle 1968; 49: 281-90.
- 1112. Brouet G, Marche J, Rist N, Chevallier J, LeMeur G. Observations on the antituberculous effectiveness of alpha-ethyl-thiosonicotinamide in tuberculosis in humans. Am Rev Respir Dis 1959; 79: 6-18.
- 1113. Lees AW. Toxicity in newly diagnosed cases of pulmonary tuberculosis treated with ethionamide. Am Rev Respir Dis 1963; 88: 347-54.
- 1114. Pernod J. Hepatic tolerance of ethionamide. Am Rev Respir Dis 1965; 92: 39-42.
- 1115. Phillips S, Trevathan R. Serum glutamic oxaloacetic transaminase elevation and possible hepatotoxicity accompanying the administration of ethionamide. Am Rev Respir Dis 1962; 86: 268-9.
- 1116. Phillips S, Tashman H. Ethionamide jaundice. Am Rev Respir Dis 1963; 87: 896-8.
- 1117. Conn HO, Binder HJ, Orr HD. Ethionamide-induced hepatitis. Am Rev Respir Dis 1964; 90: 542-52.
- 1118. Gupta DK. Acceptability of thioamides II. prothionamide. J Postgraduate Medicine 1977; 23: 181-5.
- 1119. Research Committee of the British Tuberculosis Association. A comparison of the toxicity of prothionamide and ethionamide. Tubercle 1968; 49: 125-34.
- 1120. Schütz I, Bartmann K, Radenbach KL, Siegler W. Vergleich der Verträglichkeit von Protionamid und Ethionamid im Doppelblindversuch. Klin Erforsch Tuberkul Lungenkrankheiten 1969; 140: 296-303.
- 1121. O'Brien RJ, Nunn PP. The need for new drugs against tuberculosis. Obstacles, opportunities, and next steps. Am J Respir Crit Care Med 2001; 162: 1055-8.
- 1122. Jones PB, Parrish NM, Houston TA, Stapon A, Bansal NP, Dick JD, Townsend CA. A new class of antituberculosis agents. J Med Chem 2000; 43: 3304-14.
- 1123. Kasik JE. The nature of mycobacterial penicillinase. Am Rev Respir Dis 1965; 91: 117-9.
- 1124. Segura C, Salvadó M, Collado I, Chaves J, Coira A. Contribution of β -lactamases to β -lactam susceptibilities of susceptible and multidrug-resistant *Mycobacterium tuberculosis* clinical isolates. Antimicrob Agents Chemother 1998; 42: 1524-6.

- 1125. Kwon HH, Tomioka H, Saito H. Distribution and characterization of β -lactamases of mycobacteria and related organisms. Tubercle Lung Dis 1995; 76: 141-8.
- 1126. Parenti F. New experimental drugs for the treatment of tuberculosis. Rev Infect Dis 1989; 11: 479-83.
- 1127. Chambers HF, Moreau D, Yajko D, Miick C, Wagner C, Hackbarth C, Kocagöz S, Rosenberg E, Hadley WK, Nikaido H. Can penicillins and other beta-lactam antibiotics be used to treat tuberculosis? Antimicrob Agents Chemother 1995; 39: 2620-4.
- 1128. Acred P, Hunter PA, Mizen L, Rolinson GN. α-amino-p-hydroxybenzylpenicillin (BRL 2333), a new broad-spectrum semisynthetic penicillin: in vivo evaluation. Antimicrob Agents Chemother 1970; 10: 416-22.
- 1129. Long AAW, Nayler JHC, Smith H, Taylor T, Ward N. Derivatives of 6aminopenicillanic acid. Part XI. α-amino-p-hydroxy-benzylpenicillin. J Chem Soc (C) 1971; 10: 1920-2.
- 1130. Tamás F. Use of amikacin and amoxicillin-clavulanic acid against *Mycobacterium tuberculosis*. (Correspondence). Chest 1993; 104: 328.
- 1131. Cynamon MH, Palmer GS. In vitro activity of amoxicillin in combination with clavulanic acid against *Mycobacterium tuberculosis*. Antimicrob Agents Chemother 1983; 24: 429-31.
- 1132. Bergmann JS, Woods GL. In vitro activity of antimicrobial combinations against clinical isolates of susceptible and resistant *Mycobacterium tuberculosis*. Int J Tuberc Lung Dis 1998; 2: 621-6.
- 1133. Abate G, Miörner H. Susceptibility of multidrug-resistant strains of *Mycobacterium tuberculosis* to amoxycillin in combination with clavulanic acid and ethambutol. J Antimicrob Chemother 1998; 42: 735-40.
- 1134. Nadler JP, Berger J, Nord JA, Cofsky R, Saxena M. Amoxicillin-clavulanic acid for treating drug-resistant *Mycobacterium tuberculosis*. Chest 1991; 99: 1025-6.
- 1135. Yew WW, Wong CF, Lee J, Wong PC, Chau CH. Do β-lactam-β-lactamase inhibitor combinations have a place in the treatment of multidrug-resistant pulmonary tuberculosis? (Correspondence). Tubercle Lung Dis 1995; 76: 90-1.
- 1136. Chambers HF, Kocagöz T, Sipit T, Turner J, Hopewell PC. Activity of amoxicillin/clavulanate in patients with tuberculosis. Clin Infect Dis 1998; 26: 874-7.
- 1137. Schraufnagel DE. Tuberculosis treatment for the beginning of the next century. Int J Tuberc Lung Dis 1999; 3: 651-62.
- 1138. Saxon A, Beall GN, Rohr AS, Adelman DC. Immediate hypersensitivity reactions to beta-lactam antibiotics. Ann Intern Med 1987; 107: 204-15.
- 1139. Neu HC. New macrolide antibiotics: azithromycin and clarithromycin. (Editorial). Ann Intern Med 1992; 116: 515-7.
- 1140. Rodvold KA. Clinical pharmacokinetics of clarithromycin. Clin Pharmacokinetics 1999; 37: 385-98.

- 1141. Ashtekar DR, Costa-Perira R, Nagrajan K, Vishvanathan N, Bhatt AD, Rittel W. In vitro and in vivo activities of the nitroimidazole CGI 17341 against *Mycobacterium tuberculosis*. Antimicrob Agents Chemother 1993; 37: 183-6.
- 1142. Berlin OGW, Young LS, Floyd-Reising SA, Bruckner DA. Comparative in vitro activity of the new macrolide A-56268 against mycobacteria. Eur J Clin Microbiol 1987; 6: 486-7.
- 1143. Heifets LB, Lindholm-Levy PJ, Comstock RD. Clarithromycin minimal inhibitory and bactericidal concentrations against *Mycobacterium avium*. Am Rev Respir Dis 1992; 145: 856-8.
- 1144. Luna-Herrera J, Reddy VM, Daneluzzi D, Gangadharam PRJ. Antituberculosis activity of clarithromycin. Antimicrob Agents Chemother 1995; 39: 2692-5.
- 1145. Mor N, Esfandiari A. Synergistic activities of clarithromycin and pyrazinamide against *Mycobacterium tuberculosis* in human macrophages. Antimicrob Agents Chemother 1997; 41: 2035-6.
- 1146. Brown BA, Wallace RJ, Onyl GO, De Rosas V. Activities of four macrolides, including clarithromycin, against *Mycobacterium fortuitum*, *Mycobacterium chelonae*, and *M. chelonae*-like organisms. Antimicrob Agents Chemother 1992; 36: 180-4.
- 1147. Burman WJ, Reves RR, Rietmeijer CA, Cohn DL. A retrospective comparison of clarithromycin versus rifampin in combination treatment for disseminated *Mycobacterium avium* complex disease in AIDS: clarithromycin decreases transfusion requirements. Int J Tuberc Lung Dis 1997; 1: 163-9.
- 1148. Pierce M, Crampton S, Henry D, Heifets L, LaMarca A, Montecalvo M, Wormser G, Jablonowski H, Jemsek J, Cynamon M, Yangco BG, Notario G, Craft JC. A randomized trial of clarithromycin as prophylaxis against disseminated *Mycobacterium avium* complex infection in patients with advanced acquired immunodeficiency syndrome. N Engl J Med 1996; 335: 384-91.
- 1149. Shafran SD, Singer J, Zarowny DP, Phillips P, Salit I, Walmsley SL, Fong IW, Gill MJ, Rachlis AR, Lalonde RG, Fanning MM, Tsoukas CM. A comparison of two regimens for the treatment of *Mycobacterium avium* complex bacteremia in AIDS: rifabutin, ethambutol, and clarithromycin versus rifampin, ethambutol, clofazimine, and ciprofloxacin. N Engl J Med 1996; 335: 377-83.
- 1150. Dautzenberg B, Truffot-Pernot C, Hazebroucq J, Legris S, Guérin C, Begelman C, Guermonprez G, Fievet MH, Chastang C, Grosset J. A randomized comparison of two clarithromycin doses for treatment of *Mycobacterium avium* complex infections. Infection 1997; 25: 16-21.
- 1151. Dubé MP, Sattler FR, Torriani FJ, See D, Havlir DV, Kemper CA, Dezfuli MG, Bozzette SA, Bartok AE, Leedom JM, Tilles JG, McCutchan JA. A randomized evaluation of ethambutol for prevention of relapse and drug resistance during treatment of *Mycobacterium avium* complex bacteremia with clarithromycinbased combination therapy. J Infect Dis 1997; 176: 1225-32.

- 1152. Suzuki K, Tsuyuguchi K, Matsumoto H, Yamamoto T, Hashimoto T, Tanaka T, Amitani R, Kuze F. Activity of KRM 1648 or rifabutin alone or in combination with clarithromycin against *Mycobacterium avium* complex in human alveolar macrophages. Int J Tuberc Lung Dis 1997; 1: 460-7.
- 1153. Roussel G, Igual J. Clarithromycin with minocyclin and clofazimine for *Mycobacterium avium intracellulare* complex lung disease in patients without the acquired immunodeficiency syndrome. Int J Tuberc Lung Dis 1998; 2: 462-70.
- 1154. Ward TT, Rimland D, Kauffman C, Huycke M, Evans TG, Heifets L. Randomized, open-label trial of azithromycin plus ethambutol vs. clarithromycin plus ethambutol as therapy for *Mycobacterium avium* complex bacteremia in patient with human immunodeficiency virus infection. Clin Infect Dis 1998; 27: 1278-85.
- 1155. Truffot-Pernot C, Lounis N, Grosset JH, Ji B. Clarithromycin is inactive against *Mycobacterium tuberculosis*. Antimicrob Agents Chemother 1995; 39: 2827-8.
- 1156. Dautzenberg B, Truffot C, Legris S, Meyohas MC, Berlie HC, Mercat A, Chevret S, Grosset J. Activity of clarithromycin against *Mycobacterium avium* infection in patients with the acquired immune deficiency syndrome. A controlled clinical trial. Am Rev Respir Dis 1991; 144: 564-9.
- 1157. Bosi S, Da Ros T, Castellano S, Banfi E, Prato M. Antimycobacterial activity of ionic fullerene derivatives. Bioorganic Med Chem Lett 2000; 10: 1043-5.
- 1158. Agrawal KC, Bears KB, Sehgal RK, Brown JN, Rist PE, Rupp WD. Potential radiosensitizing agents. Dinitroimidazoles. J Med Chem 1979; 22: 583-5.
- 1159. Nagarajan K, Shankar RG, Rajapa S, Shenoy SJ, Costa-Pereira R. Nitroimidazoles XXI 2,3-dihydro-6-nitroimidazo [2,1-*b*] oxazoles with antitubercular activity. Eur J Med Chem 1989; 24: 631-3.
- 1160. Walsh JS, Wang R, Bagan E, Wang CC, Wislocki P, Miwa GT. Structural alterations that differentially affect the mutagenic and antitrichomonal activities of 5-nitroimidazoles. J Med Chem 1987; 30: 150-6.
- 1161. Stover CK, Warrener P, VanDevanter DR, Sherman DR, Arain TM, Langhorne MH, Anderson SW, Towell JA, Yuan Y, McMurray DN, Kreiswirth BN, Barry CE, Baker WR. A small-molecule nitroimidazopyran drug candidate for the treatment of tuberculosis. Nature 2000; 405: 962-6.
- 1162. Stewart GR, Ehrt S, Riley LW, Dale JW, McFadden J. Deletion of the putative antioxidant *noxR1* does not alter the virulence of *Mycobacterium tuberculosis* H37Rv. Tuber Lung Dis 2000; 80: 237-42.
- 1163. Slayden RA, Lee RE, Armour JW, Cooper AM, Orme IM, Brennan PJ, Besra GS. Antimycobacterial action of thiolactomycin: an inhibitor of fatty acid and mycolic acid synthesis. Antimicrob Agents Chemother 1996; 40: 2813-9.
- 1164. Murugasu-Oei B, Dick T. Bactericidal activity of nitrofurans against growing and dormant *Mycobacterium bovis* BCG. J Antimicrob Chemother 2000; 46: 917-9.

- 1165. Oleksijew A, Meulbroek J, Ewing P, Jarvis K, Mitten M, Paige L, Tovcimak A, Nukkula M, Chu D, Alder JD. In vivo efficacy of ABT-255 against drugsensitive and -resistant *Mycobacterium tuberculosis* strains. Antimicrob Agents Chemother 1998; 42: 2674-7.
- 1166. Cynamon MH, Klemens SP, Sharpe CA, Chase S. Activities of several novel oxazolidinones against *Mycobacterium tuberculosis* in a murine model. Antimicrob Agents Chemother 1999; 43: 1189-91.
- 1167. Eustice DC, Feldman PA, Zajac I, Slee AM. Mechanism of action of DuP 721: inhibition of an early event during initiation of protein synthesis. Antimicrob Agents Chemother 1988; 32: 1218-22.
- 1168. Lin AH, Murray RW, Vidmar TJ, Marotti KR. The oxazolidinone eperezolid binds to the 50S ribosomal subunit and competes with binding of chloramphenicol and lincomycin. Antimicrob Agents Chemother 1997; 41: 2127-31.
- 1169. Coffey GL, Anderson LE, Fisher MW, Galbraith MM, Hillegas AB, Kohberger DL, Thompson PA, Weston KS, Ehrlich J. Biological studies of paromomycin. Antibiotica et Chemotherapia 1959; 9: 730-80.
- 1170. Gilbert DN. Aminoglycosides. *In:* Mandell GL, Bennett JE, Dolin R, Eds. Principles and practice of infectious diseases. New York: Churchill Livingstone, 2000; 307-336.
- 1171. Donald PR, Sirgel FA, Kanyok TP, Danziger LH, Venter A, Botha FJ, Parkin DP, Seifart HI, Van de Wal BW, Maritz JS, Mitchison DA. Early bactericidal activity of paromomycin (aminosidine) in patients with smear-positive pulmonary tuberculosis. Antimicrob Agents Chemother 2000; 44: 3285-7.
- 1172. Kristiansen JE, Amaral L. The potential management of resistant infections with non-antibiotics. J Antimicrob Chemother 1997; 40: 319-27.
- 1173. Amaral L, Kristiansen JE. Phenothiazines: an alternative to conventional therapy for the initial management of suspected multidrug resistant tuberculosis. A call for studies. Int J Antimicrob Agents 2000; 14: 173-6.
- 1174. Kristiansen JE, Vergmann B. The antibacterial effect of selected phenothiazines and thioxanthines on slow-growing mycobacteria. Acta Pathol Microbiol Immunolog Scand Sect B 1986; 94: 393-8.
- 1175. Chakrabarty AN, Bhattacharya CP, Dastidar SG. Antimycobacterial activity of methidilazine (Md), an antimicrobic phenothiazine. APMIS 1993; 101: 449-54.
- 1176. Amaral L, Kristiansen JF, Abebe LS, Millett W. Inhibition of the respiration of multi-drug resistant clinical isolates of *Mycobacterium tuberculosis* by thoridazine: potential use for initial therapy of freshly diagnosed tuberculosis. J Antimicrob Chemother 1996; 38: 1049-53.
- 1177. Viveiros M, Amaral L. Enhancement of antibiotic activity against poly-drug resistant *Mycobacterium tuberculosis* by phenothiazines. Int J Antimicrob Agents 2001; 17: 225-8.

- 1178. Amaral L, Kristiansen JE, Viveiros M, Atouguia J. Activity of phenothiazines against antibiotic-resistant *Mycobacterium tuberculosis*: a review supporting further studies that may elucidate the potential use of thiridazine as anti-tuberculosis therapy. J Antimicrob Chemother 2001; 47: 505-11.
- 1179. Crowle AJ, Douvas GS, May MH. Chlorpromazine: a drug potentially useful for treating mycobacterial infections. Chemotherapy 1992; 38: 410-9.
- 1180. Nagata A, Ando T, Izumi R, Sakakibara H, Take T, Hayano K, Abe J. Studies on tuberactinomycin (tuberactin), a new antibiotic. I Taxonomy of producing strain, isolation and characterization. J Antibiotics 1968; 21: 681-7.
- 1181. Toyohara M, Nagata A, Havano K, Abe J. Study of the antituberculous activity of tuberactinomycin, a new antimicrobial drug. Am Rev Respir Dis 1969; 100: 228-30.
- 1182. Ohsato T, Toyohara M. Clinical study on tuberactinomycin, a new antibiotic. Kekkaku 1971; 46: 59-63.
- 1183. Ando T, Matsuura K, Izumi R, Noda T, Take T, Nagata A, Abe J. Studies on tuberactinomycin. II isolation and properties of tuberactinomycin-N, a new tuberactinomycin group antibiotic. J Antibiotics 1971; 24: 680-6.
- 1184. Toyohara M. An experimental study on the antituberculous activity of tuberactinomycin-N. Kekkaku 1972; 47: 181-7.
- 1185. Orme I. Beyond BCG: the potential for a more effective TB vaccine. Mol Med Today 1999; 5: 487-92.
- 1186. Young DB. Current tuberculosis vaccine development. Clin Infect Dis 2000; 30 (Suppl 3): S254-S256.
- 1187. Kaufmann SHE. Is the development of a new tuberculosis vaccine possible? (Commentary). Nature Med 2000; 6: 955-60.
- 1188. Dreher D, Kok M, Pechère JC, Nicod LP. New strategies against an old plague: genetically engineered tuberculosis vaccines. Schweiz Med Wochenschr 2000; 130: 1925-9.
- 1189. Stanford JL. Immunotherapy for leprosy and tuberculosis. Progr Drug Research 1989; 33: 415-47.
- 1190. Durban Immunotherapy Trial Group. Immunotherapy with *Mycobacterium vaccae* in patients with newly diagnosed pulmonary tuberculosis: a randomised controlled trial. Lancet 1999; 354: 116-9.
- 1191. Johnson JL, Kamya RM, Okwera A, Loughlin AM, Nyole S, Hom DL, Wallis RS, Hirsch CS, Wolski K, Foulds J, Mugerwa RD, Ellner JJ. Randomized controlled trial of *Mycobacterium vaccae* immunotherapy in non-human immunodeficiency virus-infeted Ugandan adults with newly diagnosed tuberculosis. J Infect Dis 2000; 181: 1304-12.
- 1192. Mayo REP, Stanford JL. Double-blind placebo-controlled trial of *Mycobacterium vaccae* immunotherapy for tuberculosis in KwaZulu, South Africa, 1991-97. Trans R Soc Trop Med Hyg 2000; 94: 563-8.

- 1193. Hondalus MK, Bardarov S, Russel R, Chan J, Jacobs WR, Jr., Bloom BR. Attenuation of and protection induced by a leucine auxotroph of *Mycobacterium tuberculosis*. Infect Immun 2000; 68: 1888-2898.
- 1194. Smith DA, Parish T, Stoker NG, Bancroft GJ. Characterization of auxotrophic mutants of *Mycobacterium tuberculosis* and their potential as vaccine candidates. Infect Immun 2001; 69: 1142-50.
- 1195. Lowrie DB, Tascon RE, Colston MJ, Silva CL. Towards a DNA vaccine against tuberculosis. Vaccine 1994; 12: 1537-40.
- 1196. Tascon RE, Colston MJ, Ragno S, Stavropoulos E, Gregory D, Lowrie DB. Vaccination against tuberculosis by DNA injection. Nature Med 1996; 2: 888-92.
- 1197. Kumar V, Sercarz E. Genetic vaccination: the advantage of going naked. (Editorial). Nature Med 1996; 2: 857-9.
- 1198. Huygen K, Content J, Denis O, Montgomery DL, Yawman AM, Deck RR, DeWitt CM, Orme IM, Baldwin S, D'Souza C, Drowart A, Lozes E, Vandenbussche P, Van Vooren JP, Liu MA, Ulmer JB. Immunogenicity and protective efficacy of a tuberculosis DNA vaccine. Nature Med 1996; 2: 893-8.
- 1199. Lowrie DB, Silva CL, Colston MJ, Ragno S, Tascon RE. Protection against tuberculosis by a plasmid DNA vaccine. Vaccine 1997; 15: 834-8.
- 1200. Lowrie DB, Silva CL, Tascon RE. DNA vaccines against tuberculosis. Immunol Cell Biol 1997; 75: 591-4.
- 1201. Ulmer JB, Montgomery DL, Tang A, Zhu L, Deck RR, DeWitt C, Denis O, Orme I, Content J, Huygen K. DNA vaccines against tuberculosis. Novartis Found Symp 1998; 217: 239-53.
- 1202. Young DB, Robertson BD. TB vaccines: global solutions for global problems. Science 1999; 284: 1479-80.
- 1203. Lowrie DB, Tascon RE, Bonato VLD, Lima VMF, Faccioli LH, Stavropoulos E, Colston MJ, Hewinson RG, Moelling K, Silva CL. Therapy of tuberculosis in mice by DNA vaccination. Nature 1999; 400: 269-71.
- 1204. Chambers MA, Vordermeier HM, Whelan A, Commander N, Tascon R, Lowrie D, Hewinson RG. Vaccination of mice and cattle with plasmid DNA encoding the *Mycobacterium tuberculosis* antigen MPB83. Clin Infect Dis 2000; 30(Suppl 3): S283-S287.
- 1205. Lowrie DB, Silva CL. Enhancement of immunocompetence in tuberculosis by DNA vaccination. Vaccine 2000; 18: 1712-6.
- 1206. Feng CG, Palendira U, Demangel C, Spratt JM, Malin AS, Britton WJ. Priming by DNA immunization augments protective efficacy of *Mycobacterium bovis* Bacille Calmette-Guerin against tuberculosis. Infect Immun 2001; 69: 4174-6.

- 1207. Zhu X, Venkataprasad N, Ivanyi J, Vordermeier HM. Vaccination with recombinant vaccinia viruses protects mice against *Mycobacterium tuberculosis* infection. Immunology 1997; 92: 6-9.
- 1208. Kumar M, Behera AK, Matsuse H, Lockey RF, Mohapatra SS. A recombinant BCG vaccine generates a Th1-like response and inhibits IgE synthesis in BALB/c mice. Immunology 2000; 97: 515-21.
- 1209. Luo Y, Chen X, Han R, O'Donnell MA. Recombinant bacille Calmette-Guérin (BCG) expressing human interferon-alpha 2B demonstrates enhanced immunogenicity. Clin Exp Immunol 2001; 123: 264-70.
- 1210. Cole ST, Brosch R, Parkhill J, Garnier T, Churcher C, Harris D, Gordon SV, Eiglmeier K, Gas S, Barry CE, III, Tekaia F, Badcock K, Basham D, Brown D, Chillingworth T, Connor R, Davies R, Devlin K, Feltwell T, Gentles S, Hamlin N, Holroyd S, Hornsby T, Jagels K, Krogh A, McLean J, Moule S, Murphy L, Oliver K, Osborne J, Quail MA, Rajandream MA, Rogers J, Rutter S, Seeger K, Skelton J, Squares R, Squares S, Sulston JE, Taylor K, Whitehead S, Barrell BG. Deciphering the biology of *Mycobacterium tuberculosis* from the complete genome sequence. Nature 1998; 393: 537-52.
- 1211. Horwitz MA, Harth G, Dillon BJ, Maslesa-Galic S. Recombinant bacillus Calmette-Guérin (BCG) vaccines expressing the *Mycobacterium tuberculosis* 30kDA major secretory protein induce greater protective immunity against tuberculosis than conventional BCG vaccines in a highly susceptible animal model. Proc Natl Acad Sci 2000; 97: 13853-8.
- 1212. Zügel U, Sponaas AM, Neckermann J, Schoel B, Kaufmann SHE. gp96-peptide vaccination of mice against intracellular bacteria. Infect Immun 2001; 69: 4164-7.
- 1213. Weinreich Olsen A, Hansen PR, Holm A, Andersen P. Efficient protection against *Mycobacterium tuberculosis* by vaccination with a single subdominant epitope from the ESAT-6 antigen. Eur J Immunol 2000; 30: 1724-32.
- 1214. Weinreich Olsen A, van Pinxteren LAH, Meng Okkels L, Birk Rasmussen P, Andersen P. Protection of mice with a tuberculosis subunit vaccine based on a fusion protein of antigen 85B and ESAT-6. Infect Immun 2001; 69: 2773-8.
- 1215. Letvin NL, Bloom BR, Hoffman SL. Prospects for vaccines to protect against AIDS, tuberculosis, and malaria. JAMA 2001; 285: 606-11.
- 1216. Brooks JV, Frank AA, Keen MA, Bellisle JT, Orme IM. Boosting vaccine for tuberculosis. Infect Immun 2001; 69: 2714-7.



IMPRESSION, BROCHAGE IMPRIMERIE CHIRAT 42540 ST-JUST-LA-PENDUE AVRIL 2002 DÉPÔT LÉGAL 2002 N° 4405

IMPRIMÉ EN FRANCE